Thermogelling biomimetic hydrogel supporting 3D neuronal networks

Donatella Di Lisa¹, Elena Dellacasa ¹, Lorenzo Muzzi ¹, Alberto Lagazzo ², Monica Frega³, Sergio Martinoia and Laura Pastorino ¹

¹University of Genova, Dept. of Informatics, Bioengineering, Robotics and System Engineering, Via Opera Pia 13, 16145 Genova, Italy

²University of Genova, Dept. of Civil, Chemical and Environmental, Via Opera Pia 15, 16145 Genova, Italy

³University of Twente, Dept. of Clinical neurophysiology, University of Twente, Drienerlolaan 5, 7522 NB Enschede, Netherlands

Engineered natural biomaterials, able to perform the sol-gel transition in the presence of cells under specific external stimuli, have gained great attention in the field of tissue engineering and regenerative medicine [1], despite of this, for the encapsulation of neurons and the development of 3D neuronal networks, are still limited.

In order to study neuronal (dys)functions and particularly neuronal connectivity, *in vitro* human brain models need to include not only a chemically and physically relevant microenvironment, but also structural network complexity [2]. To mimic the brain ECM environment, a porous and soft structure is preferred in the design of an artificial neural network.

The goal of this study was to demonstrate the versatility of thermosensitive chitosan-based scaffold as an artificial matrix for 3D neuronal networks for in vitro studies and as an injectable-hydrogel for the in-vivo applications or innovative ink for 3D bioprinter. Chitosan is a widely used biomaterial, well known for its biocompatibility, biodegradability, muco-adhesiveness as well as its antibacterial activity [3]. It is already demonstrated that chitosan is able to sustain the development of 3D scaffold for neuronal networks [4]. In this work, chitosan thermogels were fabricated and characterized by different techniques. The scaffold was used to encapsulate primary rat neurons and human-induced neurons cultures to carry out morphological characterization by immunofluorescence techniques and a preliminary electrophysiological characterization of spontaneous activity by Micro-Electrode Arrays.

References

- [1] DeVolder, R., & Kong, H. J. (2012). Wiley Interdisciplinary Reviews: Systems Biology and Medicine, 4(4), 351-365.
- [2] Bastiaens A, Xie S, Luttge R. Micromachines (Basel), (2019), Sep 23;10(10):638.
- [3] I.Y. Kim, S.J. Seo, H.S. Moon, M.K. Yoo, I.Y. Park, B.C. Kim, et al., Biotechnol. Adv. 26 (2008).

[4] Tedesco, M. T., Di Lisa, D., Massobrio, P., Colistra, N., Pesce, M., Catelani, T., ... & Pastorino, L. (2018), Biomaterials, 156, 159-171.