

Review Article

Effects of Curcumin on Aging: Molecular Mechanisms and Experimental Evidence

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Aging is characterized by a progressive inability to maintain homeostasis, self-repair, renewal, performance, and fitness of different tissues throughout the lifespan. Senescence is occurring following enormous intracellular or extracellular stress stimuli. Cellular senescence serves as an antiproliferative process that causes permanent cell cycle arrest and restricts the lifespan. Senescent cells are characterized by terminal cell cycle arrest, enlarged lysosome, and DNA double-strand breaks as well as lipofuscin granularity, senescence-associated heterochromatin foci, and activation of DNA damage response. Curcumin, a hydrophobic polyphenol, is a bioactive chemical constituent of the rhizomes of *Curcuma longa* Linn (turmeric), which has been extensively used for the alleviation of various human disorders. In addition to its pleiotropic effects, curcumin has been suggested to have antiaging features. In this review, we summarized the therapeutic potential of curcumin in the prevention and delaying of the aging process.

1. Introduction

Aging is identified by a progressive inability to maintain homeostasis, self-repair, renewal, performance, and fitness of different tissues with advancing age [1]. The picture of aging is characterized by genetic and environmental factors ultimately leading to gradual but persistent reduction in cellular proliferation, abnormal oxygen metabolism, and structural instability [2]. A complex gene network contributes to organism lifespan by regulating several critical pathways including protein synthesis and catabolism, energy metabolism, redox balance, intracellular communication, DNA repair, inflammation, cellular senescence, and death [3]. The aging process also involves the vascular system. In this context, cell senescence

involving either endothelial cells (ECs) or vascular smooth muscle cells (VSMCs) [4] determines structural and functional alterations resulting in development of endothelial dysfunction [5]. Previous researches identified several molecules and signaling pathways involved in the aging process: among them, growth hormone (GH)/insulin-like growth factor 1 (IGF1)/forkhead box O (FOXO) pathway, target of rapamycin (TOR)/ribosomal S6 kinase (S6K), sirtuins (Sirts), p38 mitogen-activated protein kinase (MAPK), and AMP-activated protein kinase (AMPK) [6–8]. Despite many efforts in clarifying the biology of aging and its cellular and molecular mechanisms, standardized biomarkers and therapeutic targets are scarce. Only several senotherapeutics, agents which inhibit senescence (senomorphics) and selectively kill senescent cells (senolytics), have

been proposed. Senolytics are drugs that particularly target senescent cells through promoting the apoptosis of senescence [9–11].

In this field of research, there is a growing interest towards the natural compound curcumin (CUR; diferuloylmethane), which is known as an active therapeutic compound against various human disorders owing to its numerous pharmacological actions [12–17]. In light of this, research groups worldwide are attempting to clarify biological pathways, pharmaceutical properties, and potential clinical application of CUR [18]. In this narrative review, we will summarize the therapeutic potential of CUR, especially focusing on prevention and delaying of the aging process.

2. Hallmarks of Aging

2.1. Oxidative Stress. A prooxidant environment certainly contributes to the aging process by sustaining oxidative modifications of cellular molecules [19–21]. Targets of oxidative stress (OS) include structural damage in cellular macromolecules such as nuclear and mitochondrial DNA, proteins, and lipids [22]. Nevertheless, the “free radical theory of aging” is no longer considered a primitive causal pathway. Free radicals and related oxidants are a subset of stressors with which all living beings must cope with over their lifespans. Rather, the concept of “defective adaptive homeostasis” better describes how aging organisms fail to dynamically expand the homeostatic range of stress defense and repair systems. Indeed, many signal transduction pathways contribute to best fit cellular response to a particular need.

2.2. Cellular Senescence. Cellular response to stressors includes three distinctive cellular processes: apoptosis, autophagy, and senescence [23–25]. The latter (from the latin term “senex”: growing old) occurs in response to enormous intracellular or extracellular stress stimuli [26]. Cellular senescence was firstly described by Hayflick and Moorhead [27] as an antiproliferative process leading to permanent cell cycle arrest lifespan reduction [25]. Such effect on the biological clock (Hayflick limit) is generally associated with progressive telomere attrition/dysfunction [28, 29], loss of proteostasis, induction of genes located in the INK4a/ARF locus [30], aberrant oncogene activation, DNA damage during cell division/replication, and apoptosis-resistance [31]. Leading mediators of cellular senescence include the p16^{INK4a/Rb} and tumor suppressor p53/p21^{CIP1/WAF1} families of cyclin-dependent kinase (CDK) [32]. Senescent cells endure futile growth, hypertrophy, and hyperfunctions, together with generation and release of inflammatory mediators named senescence-associated secretory phenotype (SASP) [33, 34]. SASP includes multiple inflammatory elements such as interleukin- (IL-) 6, IL-8, IL-1, tumor necrosis factor- α (TNF- α), nuclear factor kappa B (NF- κ B), and growth factors like insulin-like growth factor- (IGF-) 1, platelet-derived growth factor (PDGF), vascular endothelial growth factor (VEGF), and basic fibroblast growth factor (bFGF) [35, 36].

Alongside SASP, the core event in cellular senescence cell nucleus are the disturbances in DNA repair mechanisms, which determine DNA double-strand breaks senescence-associated heterochromatin foci (SAHF), terminal cell cycle

arrest with resistance to apoptosis, and loss of regeneration/resilience [37]. Additional features include enlarged lysosomes, overexpression of senescence associated β -galactosidase (SA- β -gal), and lipofuscin granularity as well. A relevant feature of aging is chronic low grade inflammation, referred to as “inflammaging” which is the age-related inflammatory status, results from immunosenescence, as it is found to be associated with the majority of age-related diseases sharing an inflammatory basis [38]. Together with immunological elements, cellular senescence and the SASP are the major contributors to inflammaging.

That cellular senescence may have a causative role in organismal aging [39]. During aging, senescent cells are possibly persistent, activated by random molecular damage and related with the activation of a DNA damage response [40]. The collection of senescent cells in animal organs may be involved in the aging process through reducing the renewal competence of tissues [30] and/or via reforming the tissue structure and activity by secretion of matrix metalloproteinases, epithelial growth factors, and inflammatory mediators which could intrude with the tissue microenvironment [41]. Therefore, tissue homeostasis will be compromised which finally will result to aging.

2.3. Sirtuins. Sirtuins are NAD⁺-dependent deacetylases, ubiquitously distributed in either prokaryote or eukaryote cells [42]. In mammals, 7 *Sirt* genes (Sirt1 to Sirt7) have been identified. Sirt1 belongs to the class III histone deacetylases (HDAC) with activity on various transcriptional factors (TFs), histones, and cytoplasmic proteins with acyl-lysine residues [43]. Antiaging properties of Sirt1 include the suppression of a typical senescent secretome through epigenetic gene modulation [44]. However, the antiaging effects of Sirt1 are far from being elucidated, potentially ranging from mitochondrial respiration to stress modulation, energy expenditure, and p53 deacetylation [37, 45].

3. Curcumin

Due to their ubiquitous distribution in food, phytochemicals attract more attention because of their obvious safety. Accumulating evidences reported how phytochemicals that can extend lifespan also enhance wellness in different heterotrophic organisms [46–49]. The hydrophobic yellow polyphenol CUR is a bioactive chemical constituent of the rhizome of *Curcuma longa* Linn, extensively used in cooking as food coloring and preservative. CUR is the main chief ingredient of turmeric representing nearly 2–5% of the plant [50]. Toxicity studies claimed it is a safe compound agent even at high doses [51]. Concerning effectiveness, several lines of evidence highlighted a pleiotropic potential of CUR towards several human diseases, such as malignancies, skin and immune-related disorders, cardiovascular diseases, pulmonary and renal fibrosis, nonalcoholic fatty liver disease (NAFLD), fatigue, neuropathic pain, bone and muscle loss, neurodegenerative disease, ocular diseases, leprosy, osteoporosis, leishmaniosis, and HIV infection [52–57]. Pleiotropic functions of CUR mainly rely on the inhibition of I κ B kinase (IKK) phosphorylation [58] and the consequent suppression of the nuclear translocation of the NF- κ B

p65 subunit [59]. As an alternative epigenetic modulator, CUR also enhances Sirt1 expression at both mRNA and protein levels, ultimately resulting in the suppression of histone acetyltransferase (HAT) activity and increased NAD⁺/NADH ratio [60, 61]. With the same mechanism, CUR modulated the expression of several types of microRNAs [62–65]. Through those mechanisms, CUR supplementation in human melanoma cells induces growth arrest in the G2/M phase and then apoptosis [66]. Other studies also reported that CUR may target oncogene expression, angiogenesis, invasion, and metastatic dissemination [67, 68] by interfering with several other intracellular pathways including hypoxia-inducible factor-1 α (HIF-1 α), mammalian sterile 20-like kinase 1 (MST1), enhancer of zeste homolog 2 (EZH2), platelet-derived growth factor (PDGF) receptor binding, Wnt/ β -catenin, transforming growth factor beta (TGF- β), Sonic Hedgehog, Notch, and phosphatidylinositol 3-kinase (PI3K)/Akt/mammalian target of rapamycin (mTOR) cascade [69–71]. Alongside with anti-tumorigenic activity, CUR was also shown to induce antimicrobial, antioxidant, antiglycemic, antiseptic, and analgesic effects [72–74]. This “pleiotropic” potential may be ascribed to the potent metal-chelating effects of CUR, which include the scavenging of the superoxide anion, hydroxyl radical, singlet oxygen, and nitrogen dioxide [75, 76]. In line with this, other studies demonstrated that CUR may reduce levels of malondialdehyde (MDA), protein carbonyls, thiols, and nitrotyrosines. With regard to inflammation, CUR stimulates a xenobiotic response with upregulation of defense genes (e.g., phase II enzymes and hemeoxygenase-1 [HO-1]) [77] and suppression of proinflammatory transcription factors (e.g., activator protein-1 [AP1]) and cytokines (e.g., TNF- α , IL-1b, IL-6, IL-8, and monocyte chemoattractant protein 1 [MCP-1]), signal transducer activator of transcription (STAT), peroxisome proliferator-activated receptor- γ (PPAR- γ), activating transcription factor 3 (ATF3), C/EBP homologous protein (CHOP), and the inducible inflammatory enzymes cyclooxygenase- (COX-) 2 and metalloproteinases [78].

Finally, as observed in human skin fibroblasts, CUR may activate cellular stress response by interacting with the thiol-disulfide redox system. Such stress determines a rise in cellular GSH amounts via HO-1 and nuclear factor E2-related factor 2 (NRF2) signaling [79], ultimately improving cellular antioxidant defenses [80, 81]. Moreover, several studies indicated that CUR and may be used as senolytic and anti-inflammatory agents for senescent cells [82, 83]. For instance, a CUR analog, EF24, promoted senescent cell apoptosis and showed protection effect against ionizing-stimulated senescent cells [83].

4. Effect of Curcumin on Aging/Longevity

4.1. Vascular Aging. Further enhancing a wide spectrum of activity, growing evidence indicates CUR as a promising anti-aging agent (Table 1; Figure 1) [84, 85]. The effects of CUR feeding have been largely investigated in animal models, unanimously reporting a suppression of intermediated oxidative stress (e.g., lipoxigenases [LPO], MDA, lipofuscin granules, and NO) and inflammation [3, 86]. By chelating nitrogen dioxide (NO₂), CUR administration in mice significantly attenuates nitric oxide- (NO-) associated vascular endothelial

dysfunction and generation of advanced glycation end-products (AGEs), leading determinants of age-related large elastic artery stiffening [87]. As an additional mechanism, CUR fixes lysosomal membranes and reduces the function of lysosomal acid hydrolases, thus preventing the aberrant deposition of different connective tissue components in aging endothelium. A similar upgrade in endothelial function was also observed in postmenopausal women after eight weeks of treatment [88], whereas in elderly with diabetes and cardiomyopathy, CUR mitigated hypertrophy in the aging heart via suppression of p300, the global transcription activator [89]. Beneficial effects of CUR on vascular aging also concern the development of age-related macular degeneration (AMD), one of the most important causes of blindness in elderly [90, 91]. CUR remarkably increases the viability of retinal pigment epithelial cells (RPEs) modulating their proliferation apoptosis and OS [92]. Overall, those evidences suggest potential application of CUR as an innovative approach to AMD, as for other ocular diseases (e.g., ocular dryness, conjunctivitis, uveitis, pterygium, and glaucoma) [93]. Even CUR has been found to prevent the development of cataract in diabetic rats by decreasing AGE accumulation and serum LPO [94, 95]. Aging-associated cerebrovascular endothelial dysfunction with consequent chronic cerebral ischemia also plays a critical role in stroke, as well as in cerebral amyloid angiopathy, cognitive impairments, and neurodegenerative disorders [96–98]. One of the main pathological mechanisms behind this effect is the generation of ROS, due to the suppression of mitochondrial uncoupling protein 2 (UCP2) [99] and the downregulation of AMPK. CUR reverses those effects in cultured ECs, whereas in experimental models, prolonged CUR feeding decreased ROS generation and promoted cerebrovascular endothelium-dependent relaxation, finally leading to improved cerebrovascular function [100–103]. Neuroprotective effects of CUR due to UCP2 overexpression suppression especially target hippocampal neurogenesis in the CA1 area, thus affecting spatial learning and memory. CUR also prevents detrimental effects of chronic cerebral hypoperfusion by maintaining cholesterol homeostasis. CUR also contributes to maintain cholesterol homeostasis, otherwise upset by chronic cerebral ischemia. Indeed, CUR promotes cholesterol efflux through the ATP-binding cassette transporter A1 (ABCA1) and the pathway involving apoA-I and the liver X receptor (LXR)/retinoic X receptor (RXR) [104].

4.2. Cognitive Impairments. With similar mechanisms, the reduction of circulating antioxidants is tightly associated with memory loss and cognitive impairment in the elderly [105]. It is then not surprising that CUR has been reported to improve neuropsychological functions. CUR has several inhibitory effects on combining aging and Alzheimer’s disease pathophysiology, such as the suppression of amyloid precursor protein (APP) and A β synthesis and the overexpression of *ApoE* and *Nrf2* gene, as well as the prohibition of p-mTOR and p-NF- κ B [106, 107]. CUR prevents D-gal-induced brain aging and cognitive impairment through increments of antioxidant enzymes and inhibition of apoptosis [108]. Beneficial effects of CUR on mental abilities and functional capacities are associated with a LPO reduction in brain tissue [109], especially in

TABLE 1: Antiaging effect of curcumin.

Compound	Animal model	Effect	Reference
Curcumin	(i) Aged female Wistar rats	(i) Decreasing the MDA and LPO levels in brain tissue	[109]
Curcumin (20, 40, and 80 μ M)	(i) Aging RPE cells	(i) Improvement of cell viability (ii) Reducing the apoptosis and OS (iii) Decreasing the expression of apoptosis-related proteins and OS biomarkers	[92]
Curcumin (0.2%)	(i) Male Sprague Dawley rats (ii) UCP2 knockout (UCP2 ^{-/-}) (iii) Matched wild-type mice	(i) Restoring the impaired cerebrovascular endothelium-dependent vasorelaxation (ii) Promoting eNOS and AMPK phosphorylation (iii) Overexpression of UCP2 and reduction of ROS generation	[103]
Curcumin (0.2%)	(i) Male C57BL/6N mice	(i) Ameliorates age-associated large elastic artery stiffening (ii) Improvement of NO-mediated vascular endothelial dysfunction (iii) Oxidative stress (iv) Decreasing the collagen I and AGEs in the arterial wall	[87]
Curcumin (100 μ M)	(i) Wild-type Canton-S flies	(i) Protective effect against radiation damage (ii) Decrement of the amount of protein carbonylation and γ H2Ax foci	[142]
Curcumin (100, 200, and 400 mg/kg BW)	(i) Female Wistar albino rats	(i) Increased the NO and MDA levels	[3]
Curcumin (50 mg/kg)	(i) Adult and aging male C57BL/6 mice	(i) Modulation of hippocampal redox status (ii) Restoring aging-related loss of synapse input specificity of HFS-LTP (i) Improving the spatial learning and memory (ii) Alleviating pathological change (iii) Reduction of the level of MDA (iv) Increment of the activity of SOD (v) Inducing HO-1 protein expression (vi) Increasing the protein levels of UCP2 (vii) Inhibiting OS induced by ischemia	[110]
Curcumin (50 and 100 mg/kg)	(i) Male Sprague Dawley rats	(i) Inhibition of $A\beta$ aggregation (ii) Amelioration of cognitive dysfunction (iii) Decrements of the amount of aggregated $A\beta$ and tau	[167]
PE859	(i) SAMP8	(i) Mitigated the H ₂ O ₂ -induced endothelial premature senescence (ii) Decrements of population of senescence-related β -galactosidase-positive cells (iii) Motivating cell division	[158]
Curcumin (5 to 100 μ M)	(i) HUVECs	(iv) Dwindling RNA amplification of senescence-related protein p21, OS, and apoptosis (v) Induction of the expression of the phosphorylation of eNOS (vi) Increments of the amount of NO (vii) Stimulation of the transcription, translation, and enzymatic activity of Sirt1	[159]

TABLE 1: Continued.

Compound	Animal model	Effect	Reference
Piperine (12 mg/kg)+curcumin (40 mg/kg)	(i) Adult male Wistar rats	(i) Improvement of spatial memory and serotonergic signaling (ii) Decrements of OS and lipofuscin deposition (iii) Higher hippocampal volume (iv) Hippocampal neuroprotection (v) Promotion of cognition (vi) Inhibition of senescence by the free radical quenching	[153]
Curcumin (50 mg/kg)	(i) SAMP8 mice	(i) Narrowing the hippocampal SOD activities (ii) Elevation of the amount of p-CaMKII in the stratum lucidum of hippocampal CA3 and p-NMDARI in the hippocampal membrane	[156]
Curcumin (0 to 500 mM)	(i) Two strains of <i>Drosophila</i> (Canton-S and Ives flies)	(i) Protection against oxidative stress (ii) Improvement in locomotion (iii) Modulating the expression of different aging-associated genes, including <i>mth</i> , <i>Thor</i> , <i>InR</i> , and <i>JNK</i>	[139]
Curcumin (0 to 200 mM)	(i) Normal-lived Ra strain (<i>Drosophila</i>)	(i) Induction of an extended longevity phenotype (ii) Slowing the aging rate (iii) Increases the adult animal's geotactic activity	[144]
Curcumin (0.5 to 1.0 mg/g of diet)	(i) Oregon-R strain (<i>Drosophila</i>)	Overexpression of Mn-SOD and CuZn-SOD genes (i) Downexpression of age-associated genes (<i>dInR</i> , <i>ATTD</i> , <i>Def</i> , <i>CecB</i> , and <i>DptB</i>) (ii) Modulating the gene expression of SOD (iii) Decrements of MDA and LPO	[143]
Galantamine (5 mg/kg) and curcumin (15 and 30 mg/kg)	(i) Old male LACA mice	(i) Postponing aging process (ii) Improving cognitive functions, locomotor activity, and antioxidant (iii) Decrements of acetylcholine esterase activity (iv) Restoring the mitochondrial enzyme complex execution	[154]
Curcumin	(i) Transgenic <i>Drosophila</i>	(i) Increments of amyloid fibril conversion by decreasing the prefibrillar/oligomeric species of A β	[148]
Curcumin and disulfiram/gram of media	(i) Male <i>D. melanogaster</i>	(i) Promotion of SOD activity	[168]

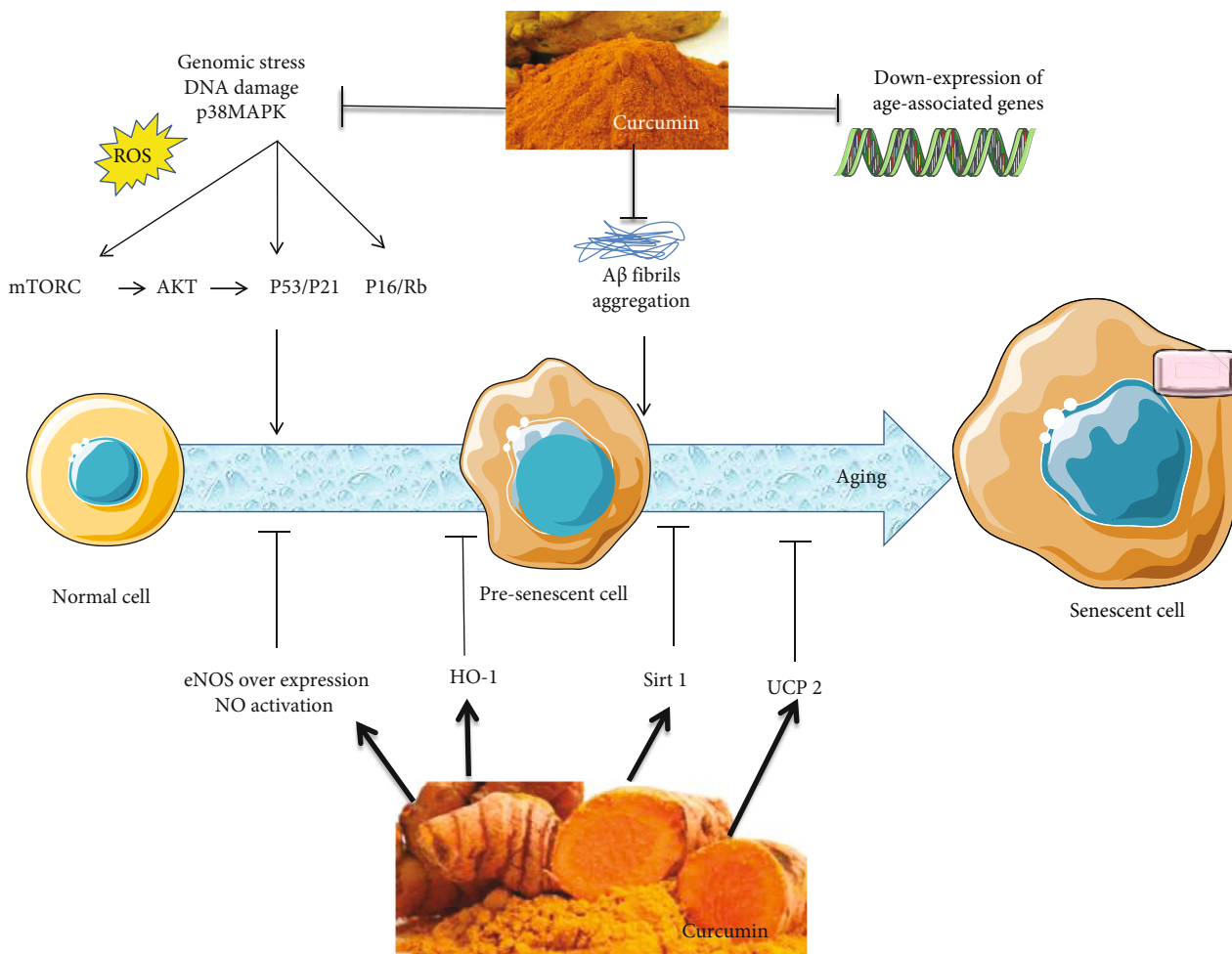


FIGURE 1: Mechanisms by which curcumin modulate aging process and senescence. Curcumin inhibited OS-stimulated p38MAPK activation, A β fibril aggregation, and expression of age-associated genes (*dInR*, *ATTD*, *Def*, *CecB*, *DptB*, *mth*, *thor*, *InR*, and *JNK*), although curcumin induced eNOS, NO, Sirt1, HO-1, and UCP2 expression. Curcumin also mitigates the SASP and its aging-induction consequences of senescent cell. Abbreviations: A β : amyloid- β ; eNOS: endothelial nitric-oxide synthase; HO-1: hemeoxygenase-1; mTORC 1: mammalian/mechanistic target of rapamycin complex 1; NO: nitric oxide; ROS: reactive oxygen species; SASP: senescence-associated secretory phenotype; Sirt: sirtuins; UCP2: uncoupling protein 2.

the hippocampal area. CUR improves the redox state in this area and prevents the decline of hippocampal long-term potentiation by maintaining synapse input specificity [110, 111]. Recently, Olesen et al. described that the dysfunction of synaptic mitochondria of the hippocampus causing memory loss during aging. They showed that curcumin feeding significantly improved integration and activity of the synaptic mitochondrial of the hippocampus, inhibiting mitochondrial swelling and enhancing the production of synapses surrounding the mitochondria in mice [112].

4.3. Evidence from Experimental Models

4.3.1. Study of Longevity in *Drosophila melanogaster* and *Caenorhabditis elegans*. *Drosophila melanogaster* (*D. melanogaster*) and *Caenorhabditis elegans* (*C. elegans*) are widely recognized models for the study of aging processes [113]. In particular, *D. melanogaster* represented a paradigm of experimental gerontology during the last century [114–118]

because of its complex biology and the ease of rearing and housing as well [119, 120]. More recently, in 1983, Klass isolated the first long-lived mutants of *C. elegans* [121], which rose to become a promising model for aging investigations due to the small size, anatomical simplicity, small genome, short life cycle, and inexpensive laboratory manipulation [122]. In *C. elegans*, longevity is widely determined by the expression of the Age-1 gene [123, 124]. As one of the main elements in the insulin/insulin-like growth factor-1 signaling (IIS) axis, Age-1 is a subunit of phosphoinositide 3-kinase (PI3K), which suppresses DAF-16 action [123–125]. Suppression of the IIS pathway activates the downstream gene DAF-16, which in turn promotes the transcription of genes associated with longevity, metabolism, and response to cellular stress [126–128]. In line, increased lifespan may also be obtained through TOR inhibition, another DAF-16 suppressor [129, 130]. By sharing the same downstream signaling of DAF-16, also the *FOXO3 A* gene is involved in lifespan extension, cell growth, and stress response through a direct

activity on DNA repair and transcription involving p21/p53 and β -catenin pathways [131–133]. Noteworthy, FOXO has a multistep regulation involving not only IGF-1 but also NAD⁺/Sirt1, 5' AMPK, and OS, all known as aging genes [134]. Due to these similarity with human beings, *C. elegans* became a genetic model organism already in 1965. Multiple pharmacological interventions have been found to prolong the survival of *D. melanogaster* and *C. elegans* [135–137]. Also, CUR was shown to increase the fecundity, reproductive lifespan, and child viability of *D. melanogaster* [85]. It has been shown that CUR supplementation at the larval stage of *D. melanogaster* elevated the developmental duration and longevity of adult *Drosophila* possibly through epigenetic programming of the pace of life [138].

CUR-mediated increased longevity was observed in two distinctive strains of *D. melanogaster* (Canton-S and Ivies flies) as a result of the delayed expression of aging genes (e.g., methuselah (*mth*), *thor*, insulin receptor [*InR*], and c-jun N-terminal kinase [*JNK*]), improved locomotion, and chemoprevention as well [139]. CUR was also shown to reduce OS, DNA damage, and number of mutagenic phenotypes induced via high-dose ionizing irradiation. These effects may be ascribed to ROS scavenging and transcriptional regulation of OS-related genes, which mainly involves γ H2Ax, a histone protein belonging to the H2A family and involved in DNA damage response [140–142]. Also, *in vivo* experiments on CUR-fed diets (0.5 and 1.0 mg/g of diet) were effective in extending the average lifespan in both females (6.2% and 25.8%, respectively) and males (15.5% and 12.6%, respectively), and this effect could be more likely attributed to the overexpression of Mn-SOD and CuZn-SOD genes and the downregulation of aging genes associated with the TOR pathway including *Drosophila* insulin receptor (*dInR*), attacin-D (*ATTD*), defensin (*Def*), cecropin B (*CecB*), and dipterin B (*DptB*) genes [143, 144]. Also, in *C. elegans*, CUR effectively improves lifespan and aging by lowering intracellular ROS and lipofuscin. The effects of CUR on *C. elegans* longevity are manifested by body size and pharyngeal pumping rate but not reproduction ability. Further studies revealed that the long-lived phenotype induced by CUR was maintained in *mev-1* and *daf-16* mutants but lost in *osr-1*, *sek-1*, *skn-1*, *unc-43*, *mek-1*, *sir-2.1*, and *age-1* ones [145]. This evidence indicates that CUR would exert its effects independently of the Age-1-DAF-16 pathway but rather through other constituents of the IIS pathway. With regard to cognitive impairment, the *in vivo* experiment demonstrated that CUR can improve learning and memory also reducing A β plaque formation in the context of Alzheimer disease (AD) [146]. *D. melanogaster* is a promising animal model for research in AD [147]. By increasing amyloid fibril conversion, CUR reduces the generation of prefibrillar/oligomeric species of A β , ultimately protecting against neurotoxicity [148]. The human β -amyloid precursor cleavage enzyme (BACE-1) is another critical enzyme targeted by CUR [149, 150] in the *D. melanogaster* model of AD [150].

4.3.2. Studies of Cell Senescence: Evidence from Mice and Rats. High doses of CUR (2.5–10 μ M) were shown to trigger senescence in cancer and vascular cells [151]. On the other

hand, low doses of CUR (0.1 and 1 μ M) failed to prevent early senescence in doxorubicin-treated (VSMC) and even slightly accelerated replicative senescence in endothelial cells [152]. It is therefore evident how the antiaging effect of CUR does not rely on delayed cellular senescence. As reported by Banji et al., CUR (40 mg/kg) and piperine (12 mg/kg), especially when combined, counteract D-gal-induced senescence in male Wistar rats by targeting OS and lipofuscin deposition, finally leading to higher hippocampal volume and function with improved spatial memory and serotonergic signaling [153]. Another study even reported how long-time CUR therapy may progressively reverse cognitive dysfunction in D-gal-induced senescent mice by delaying the aging process and improving cognitive functions and locomotor activity, as well as restoring the mitochondrial enzyme complex function [154]. In a recent study, CUR supplementation rejuvenates senescence-associated changes in thymus among D-gal-induced senescent mice through promotion of proliferating cells, preventing cells from apoptosis, and enhancing the transcription of the autoimmune regulator (Aire) [155].

CUR feeding (50 mg/kg) was also tested in senescence-accelerated mouse prone (SAMP) mice resulting in increased hippocampal SOD activity as well as upregulation of p-calcium/calmodulin-dependent kinase II (p-CaMKII) in the stratum lucidum and p-N-methyl-D-aspartate receptor subunit 1 (p-NMDAR1) in the hippocampal membrane [156]. Noteworthy, clinical benefits of the CUR analogue PE859 have been recently reported and associated with reduction of A β and tau aggregates in the mouse brain [157, 158]. Overall, these findings suggest a role of CUR in improving cognitive difficulties and the expression of hippocampal plasticity-associated proteins. With regard to vascular function, CUR administration significantly mitigated premature senescence in HUVECs, characterized by a reduction of senescence-related β -galactosidase-positive cells, cell division, levels of senescence-related protein p21 RNA, OS, and apoptosis. CUR is also associated with enhanced eNOS phosphorylation and NO generation, in addition to upregulating Sirt1 transcription, translation, and enzymatic activity [159]. In light of these mechanisms, diets containing tetrahydrocurcumin (THC), the main metabolite of CUR, were demonstrated to significantly extend mean lifespan in male C57BL/6 mice [160], whereas bisdemethoxycurcumin administration delayed the OS-caused premature senescence via Sirt1/AMPK cascade activation [161]. As recently demonstrated, Sirt1 signaling also mediates the anti-inflammatory effects of CUR in C57BL/6 mice fed with high fat diet [162] in addition to improved myocardial structure and function in streptozocin-induced diabetic mice fed with THC (120 mg/kg/d) [163]. Even more recently, it has been hypothesized that the antiaging effect of CUR may rely on the control of core clock genes on which Sirt1 belongs alongside *rBmal1*, *rCry1*, *rCry2*, *rPer1*, *rPer2*, and *rRev-erba*. CUR treatment in middle aged male Wistar rats restored the phase and daily pulse of *rCry1*, *rCry2*, *rPer1*, and *rPer2* as in the young, whereas only *rPer1* and partly *rBmal1*, *rCry1*, and *rCry2* were restored in the old ones [164]. Moreover, it has been shown that CUR mitigated mouse ovarian aging, upgraded embryonic development, promoted oocyte maturation and fertilization via improvement of ovarian hormones,

and elevated the amounts of *SIRT1* and 3 genes as well as attenuation of aging-associated oxidative stress and cell death [165]. Besides, CUR can reduce oxidative stress, inflammation status, and lipofuscin deposition in aged rat liver [166].

5. Conclusion

Aging and senescence are complex processes leading to organ dysfunction. Despite being permanent, delaying the occurrence of these processes is a reliable target, and CUR might be a promising candidate for this purpose. Nevertheless, evidence from clinical studies on the long-term effects of CUR on age-related pathological events remains largely understudied. While several strategies to enhance the systemic bioavailability of CUR have been suggested, the effects of long-term therapy with such bioavailability-boosted CUR preparations is not fully known, and increased concentrations may even lead to opposite results. Pleiotropic benefits of CUR supplementation involve the control of aging genes, OS, and inflammation in both the vascular system and the central nervous system. Further studies are warranted to clarify the mechanisms of CUR function for potential clinical application.

Abbreviations

A β :	Amyloid- β
AGEs:	Advanced glycation end-products
ATTD:	Attacin-D
CecB:	Cecropin B
Def:	Defensin
dInR:	<i>Drosophila</i> insulin receptor
DptB:	Dipteracin B
HO-1:	Hemeoxygenase-1
HFS:	High-frequency stimulation
H ₂ O ₂ :	Hydrogen peroxide
HUVECs:	Human umbilical vein endothelial cells
GSH:	Glutathione
LTP:	Long-term potentiation
MDA:	Malondialdehyde
NO:	Nitric oxide
RPE:	Retinal pigment epithelial
p-CaMKII:	p-Calcium/calmodulin-dependent kinase II
SAMP8:	p-N-Methyl-D-aspartate receptor subunit 1 (p-NMDAR1), senescence-accelerated mouse prone 8
SOD:	Superoxide dismutase
UCP2:	Uncoupling protein 2.

Conflicts of Interest

The authors declare that no competing interests exist.

References

- [1] C.-W. Hung, Y.-C. Chen, W.-L. Hsieh, S.-H. Chiou, and C.-L. Kao, "Ageing and neurodegenerative diseases," *Ageing Research Reviews*, vol. 9, pp. S36–S46, 2010.
- [2] W. A. Van Voorhies and S. Ward, "Genetic and environmental conditions that increase longevity in *Caenorhabditis* ele-

- gans decrease metabolic rate," *Proceedings of the National Academy of Sciences*, vol. 96, no. 20, pp. 11399–11403, 1999.
- [3] M. Shailaja, K. M. Damodara Gowda, K. Vishakh, and N. Suchetha Kumari, "Anti-aging role of curcumin by modulating the inflammatory markers in albino Wistar rats," *Journal of the National Medical Association*, vol. 109, no. 1, pp. 9–13, 2017.
- [4] T. Minamino and I. Komuro, "Vascular cell Senescence," *Circulation Research*, vol. 100, no. 1, pp. 15–26, 2007.
- [5] E. G. Lakatta and D. Levy, "Arterial and cardiac aging: major shareholders in cardiovascular disease Enterprises," *Circulation*, vol. 107, no. 1, pp. 139–146, 2003.
- [6] L. Fontana, L. Partridge, and V. D. Longo, "Extending healthy life span—from yeast to humans," *Science*, vol. 328, no. 5976, pp. 321–326, 2010.
- [7] M. C. Haigis and D. A. Sinclair, "Mammalian sirtuins: biological insights and disease relevance," *Annu Rev Pathol Mech Dis*, vol. 5, no. 1, pp. 253–295, 2010.
- [8] C. J. Kenyon, "The genetics of ageing," *Nature*, vol. 464, no. 7288, pp. 504–512, 2010.
- [9] H. Fuhrmann-Stroissnigg, Y. Y. Ling, J. Zhao et al., "Identification of HSP90 inhibitors as a novel class of senolytics," *Nature Communications*, vol. 8, no. 1, 2017.
- [10] M. Xu, T. Pirtskhalava, J. N. Farr et al., "Senolytics improve physical function and increase lifespan in old age," *Nature Medicine*, vol. 24, no. 8, pp. 1246–1256, 2018.
- [11] L. J. Hickson, L. G. P. Langhi Prata, S. A. Bobart et al., "Senolytics decrease senescent cells in humans: preliminary report from a clinical trial of Dasatinib plus quercetin in individuals with diabetic kidney disease," *eBioMedicine*, vol. 47, pp. 446–456, 2019.
- [12] M. Ghandadi and A. Sahebkar, "Curcumin: an effective inhibitor of interleukin-6," *Current Pharmaceutical Design*, vol. 23, no. 6, pp. 921–931, 2017.
- [13] F. Ghasemi, M. Shafiee, Z. Banikazemi et al., "Curcumin inhibits NF- κ B and Wnt/ β -catenin pathways in cervical cancer cells," *Pathology Research and Practice*, vol. 215, no. 10, p. 152556, 2019.
- [14] K. Mortezaee, E. Salehi, H. Mirtavoos-mahyari et al., "Mechanisms of apoptosis modulation by curcumin: implications for cancer therapy," *Journal of Cellular Physiology*, vol. 234, no. 8, pp. 12537–12550, 2019.
- [15] Y. Panahi, N. Khalili, E. Sahebi et al., "Effects of curcuminoids plus piperine on glycemic, hepatic and inflammatory biomarkers in patients with type 2 diabetes mellitus: a randomized double-blind placebo-controlled trial," *Drug Research*, vol. 68, no. 7, pp. 403–409, 2018.
- [16] M. Mohajeri, V. Bianconi, M. F. Ávila-Rodríguez et al., "Curcumin: a phytochemical modulator of estrogens and androgens in tumors of the reproductive system," *Pharmacological Research*, vol. 156, p. 104765, 2020.
- [17] M. Sadeghian, S. Rahmani, T. Jamialahmadi, T. P. Johnston, and A. Sahebkar, "The effect of oral curcumin supplementation on health-related quality of life: a systematic review and meta-analysis of randomized controlled trials," *Journal of Affective Disorders*, vol. 278, pp. 627–636, 2021.
- [18] S. Hewlings and D. Kalman, "Curcumin: a review of its effects on human health," *Food*, vol. 6, no. 10, p. 92, 2017.
- [19] M. Zhao, P. Zhu, M. Fujino et al., "Oxidative stress in hypoxic-ischemic encephalopathy: molecular mechanisms

- and therapeutic strategies,” *International Journal of Molecular Sciences*, vol. 17, no. 12, p. 2078, 2016.
- [20] I. Bellezza, “Oxidative stress in age-related macular degeneration: Nrf2 as therapeutic target,” *Frontiers in Pharmacology*, vol. 9, 2018.
- [21] G. Barrera, S. Pizzimenti, M. Daga et al., “Lipid peroxidation-derived aldehydes, 4-hydroxynonenal and malondialdehyde in aging-related disorders,” *Antioxidants*, vol. 7, no. 8, p. 102, 2018.
- [22] O. Ighodaro and O. Akinloye, “First line defence antioxidants-superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPX): their fundamental role in the entire antioxidant defence grid,” *Alexandria Journal of Medicine*, vol. 54, no. 4, pp. 287–293, 2018.
- [23] J. M. Vicencio, L. Galluzzi, N. Tajeddine et al., “Senescence, apoptosis or autophagy?,” *Gerontology*, vol. 54, no. 2, pp. 92–99, 2008.
- [24] R. G. Faragher, A. McArdle, A. Willows, and E. L. Ostler, “Senescence in the aging process,” *F1000Research*, vol. 6, 2017.
- [25] S. Yanagi, H. Tsubouchi, A. Miura, A. Matsuo, N. Matsumoto, and M. Nakazato, “The impacts of cellular senescence in elderly pneumonia and in age-related lung diseases that increase the risk of respiratory infections,” *International journal of molecular sciences.*, vol. 18, no. 3, p. 503, 2017.
- [26] N. N. Hooten and M. K. Evans, “Techniques to induce and quantify cellular senescence,” *JoVE (Journal of Visualized Experiments)*, vol. 123, article e55533, 2017.
- [27] L. Hayflick and P. S. Moorhead, “The serial cultivation of human diploid cell strains,” *Experimental cell research.*, vol. 25, no. 3, pp. 585–621, 1961.
- [28] G. von Figura, D. Hartmann, Z. Song, and K. L. Rudolph, “Role of telomere dysfunction in aging and its detection by biomarkers,” *Journal of molecular medicine.*, vol. 87, no. 12, pp. 1165–1171, 2009.
- [29] J. Karlseder, A. Smogorzewska, and T. de Lange, “Senescence induced by altered telomere state, not telomere loss,” *Science*, vol. 295, no. 5564, pp. 2446–2449, 2002.
- [30] W. Y. Kim and N. E. Sharpless, “The Regulation of *_INK4_ / _ARF_* in Cancer and Aging,” *Cell*, vol. 127, no. 2, pp. 265–275, 2006.
- [31] C. Wang, D. Jurk, M. Maddick, G. Nelson, C. Martin-Ruiz, and T. Von Zglinicki, “DNA damage response and cellular senescence in tissues of aging mice,” *Aging Cell*, vol. 8, no. 3, pp. 311–323, 2009.
- [32] U. Herbig, W. A. Jobling, B. P. Chen, D. J. Chen, and J. M. Sedivy, “Telomere Shortening Triggers Senescence of Human Cells through a Pathway Involving ATM, p53, and p21^{CIP1}, but Not p16^{INK4a},” *Molecular Cell*, vol. 14, no. 4, pp. 501–513, 2004.
- [33] M. V. Blagosklonny, “Hypoxia, MTOR and autophagy,” *Autophagy*, vol. 9, no. 2, pp. 260–262, 2013.
- [34] J.-P. Coppé, C. K. Patil, F. Rodier et al., “Senescence-associated secretory phenotypes reveal cell-nonautonomous functions of oncogenic RAS and the p53 tumor suppressor,” *PLoS biology.*, vol. 6, no. 12, article e301, 2008.
- [35] H. L. Pahl, “Activators and target genes of Rel/NF- κ B transcription factors,” *Oncogene*, vol. 18, no. 49, pp. 6853–6866, 1999.
- [36] M. Demaria, N. Ohtani, S. A. Youssef et al., “An essential role for senescent cells in optimal wound healing through secretion of PDGF-AA,” *Developmental cell*, vol. 31, no. 6, pp. 722–733, 2014.
- [37] W. Grabowska, E. Sikora, and A. Bielak-Zmijewska, “Sirtuins, a promising target in slowing down the ageing process,” *Biogerontology*, vol. 18, no. 4, pp. 447–476, 2017.
- [38] F. Prattichizzo, V. De Nigris, L. La Sala, A. D. Procopio, F. Olivieri, and A. Ceriello, ““Inflammaging” as a druggable target: a senescence-associated secretory phenotype—centered view of type 2 diabetes,” *Oxidative medicine and cellular longevity.*, vol. 2016, article 1810327, p. 10, 2016.
- [39] J.-H. Chen, C. N. Hales, and S. E. Ozanne, “DNA damage, cellular senescence and organismal ageing: causal or correlative?,” *Nucleic acids research.*, vol. 35, no. 22, pp. 7417–7428, 2007.
- [40] D. Munoz-Espin and M. Serrano, “Cellular senescence: from physiology to pathology,” *Nature reviews Molecular cell biology*, vol. 15, no. 7, pp. 482–496, 2014.
- [41] J. Campisi, “Senescent cells, tumor suppression, and organismal aging: good citizens, bad neighbors,” *Cell*, vol. 120, no. 4, pp. 513–522, 2005.
- [42] B. J. North and E. Verdin, “Sirtuins: Sir2-related NAD-dependent protein deacetylases,” *Genome biology*, vol. 5, no. 5, p. 224, 2004.
- [43] T. Liu, P. Y. Liu, and G. M. Marshall, “The critical role of the class III histone deacetylase SIRT1 in cancer,” *Cancer research*, vol. 69, no. 5, pp. 1702–1705, 2009.
- [44] T. Hayakawa, M. Iwai, S. Aoki et al., “SIRT1 suppresses the senescence-associated secretory phenotype through epigenetic gene regulation,” *PLoS One*, vol. 10, no. 1, article e0116480, 2015.
- [45] C. Cantó and J. Auwerx, “Caloric restriction, SIRT1 and longevity,” *Trends in Endocrinology & Metabolism*, vol. 20, no. 7, pp. 325–331, 2009.
- [46] K. B. Harikumar and B. B. Aggarwal, “Resveratrol: a multitargeted agent for age-associated chronic diseases,” *Cell Cycle*, vol. 7, no. 8, pp. 1020–1035, 2008.
- [47] D. Vauzour, “Dietary polyphenols as modulators of brain functions: biological actions and molecular mechanisms underpinning their beneficial effects,” *Oxidative medicine and cellular longevity*, vol. 2012, Article ID 914273, 2012.
- [48] M. Adrian, P. Jeandet, J. Veneau, L. A. Weston, and R. Bessis, “Biological activity of resveratrol, a stilbenic compound from grapevines, against *Botrytis cinerea*, the causal agent for gray mold,” *Journal of Chemical Ecology*, vol. 23, no. 7, pp. 1689–1702, 1997.
- [49] A. Murakami, “Modulation of protein quality control systems by food phytochemicals,” *Journal of clinical biochemistry and nutrition*, vol. 52, no. 3, pp. 215–217, 2013.
- [50] G. Kapakos, V. Youreva, and A. K. Srivastava, *Cardiovascular Protection by Curcumin: Molecular Aspects*, vol. 49, no. 5, pp. 306–315, 2012.
- [51] V. Soleimani, A. Sahebkar, and H. Hosseinzadeh, “Turmeric (*Curcuma longa*) and its major constituent (curcumin) as nontoxic and safe substances: Review,” *Phytotherapy Research*, vol. 32, no. 6, pp. 985–995, 2018.
- [52] N. Pescosolido, R. Giannotti, A. M. Plateroti, A. Pascarella, and M. Nebbioso, “Curcumin: therapeutical potential in ophthalmology,” *Planta Medica*, vol. 80, no. 4, pp. 249–254, 2014.
- [53] D. Saleheen, S. A. Ali, K. Ashfaq, A. A. Siddiqui, A. Agha, and M. M. Yasinzi, “Latent activity of curcumin against

- leishmaniasis in vitro," *Biological and Pharmaceutical Bulletin*, vol. 25, no. 3, pp. 386–389, 2002.
- [54] C. D. Gomes, L. V. Alegrio, L. Leon, and C. Araújo, "Synthetic derivatives of curcumin and their activity against *Leishmania amazonensis*," *Arzneimittel-Forschung*, vol. 52, no. 2, pp. 120–124, 2002.
- [55] W. Jordan and C. Drew, "Curcumin—a natural herb with anti-HIV activity," *Journal of the National Medical Association*, vol. 88, no. 6, p. 333, 1996.
- [56] Z. Chen, J. Xue, T. Shen, S. Mu, and Q. Fu, "Curcumin alleviates glucocorticoid-induced osteoporosis through the regulation of the Wnt signaling pathway," *International journal of molecular medicine*, vol. 37, no. 2, pp. 329–338, 2016.
- [57] T. Koide, M. Nose, Y. Ogihara, Y. Yabu, and N. Ohta, "Leishmanicidal effect of curcumin in vitro," *Biological and Pharmaceutical Bulletin*, vol. 25, no. 1, pp. 131–133, 2002.
- [58] S. Singh and B. B. Aggarwal, "Activation of Transcription Factor NF- κ B Is Suppressed by Curcumin (Diferuloylmethane) (*)," *Journal of Biological Chemistry*, vol. 270, no. 42, pp. 24995–25000, 1995.
- [59] G.-Y. Kim, K.-H. Kim, S.-H. Lee et al., "Curcumin inhibits immunostimulatory function of dendritic cells: MAPKs and translocation of NF- κ B as potential targets," *The Journal of Immunology*, vol. 174, no. 12, pp. 8116–8124, 2005.
- [60] E. Zendedel, A. E. Butler, S. L. Atkin, and A. Sahebkar, "Impact of curcumin on sirtuins: a review," *Journal of Cellular Biochemistry*, vol. 119, no. 12, pp. 10291–10300, 2018.
- [61] T. Morimoto, Y. Sunagawa, T. Kawamura et al., "The dietary compound curcumin inhibits p300 histone acetyltransferase activity and prevents heart failure in rats," *The Journal of clinical investigation*, vol. 118, no. 3, pp. 868–878, 2008.
- [62] S. C. Gupta, G. Kismali, and B. B. Aggarwal, "Curcumin, a component of turmeric: from farm to pharmacy," *BioFactors*, vol. 39, no. 1, pp. 2–13, 2013.
- [63] S. S. Boyanapalli and A.-N. T. Kong, "Curcumin, the king of spices: epigenetic regulatory mechanisms in the prevention of cancer, neurological, and inflammatory diseases," *Current pharmacology reports*, vol. 1, no. 2, pp. 129–139, 2015.
- [64] M. Remely, L. Lovrecic, A. L. de la Garza et al., "Therapeutic perspectives of epigenetically active nutrients," *British journal of pharmacology*, vol. 172, no. 11, pp. 2756–2768, 2015.
- [65] S. Reuter, S. C. Gupta, B. Park, A. Goel, and B. B. Aggarwal, "Epigenetic changes induced by curcumin and other natural compounds," *Genes & nutrition*, vol. 6, no. 2, pp. 93–108, 2011.
- [66] M. Zheng, S. Ekmekcioglu, E. T. Walch, C.-H. Tang, and E. A. Grimm, "Inhibition of nuclear factor- κ B and nitric oxide by curcumin induces G2/M cell cycle arrest and apoptosis in human melanoma cells," *Melanoma research*, vol. 14, no. 3, pp. 165–171, 2004.
- [67] C. Schaaf, B. Shan, M. Buchfelder et al., "Curcumin acts as anti-tumorigenic and hormone-suppressive agent in murine and human pituitary tumour cells in vitro and in vivo," *Endocrine-related cancer*, vol. 16, no. 4, pp. 1339–1350, 2009.
- [68] L. Hamzehzadeh, S. L. Atkin, M. Majeed, A. E. Butler, and A. Sahebkar, "The versatile role of curcumin in cancer prevention and treatment: a focus on PI3K/AKT pathway," *Journal of cellular physiology*, vol. 233, no. 10, pp. 6530–6537, 2018.
- [69] T. S. Ramasamy, A. Z. Ayob, H. H. L. Myint, S. Thiagarajah, and F. Amini, "Targeting colorectal cancer stem cells using curcumin and curcumin analogues: insights into the mechanism of the therapeutic efficacy," *Cancer cell international*, vol. 15, no. 1, p. 96, 2015.
- [70] B. Bao, S. Ali, S. Banerjee et al., "Curcumin analogue CDF inhibits pancreatic tumor growth by switching on suppressor microRNAs and attenuating EZH2 expression," *Cancer research*, vol. 72, no. 1, pp. 335–345, 2012.
- [71] J. H. Seo, K. J. Jeong, W. J. Oh et al., "Lysophosphatidic acid induces STAT3 phosphorylation and ovarian cancer cell motility: their inhibition by curcumin," *Cancer letters*, vol. 288, no. 1, pp. 50–56, 2010.
- [72] V. P. Menon and A. R. Sudheer, "Antioxidant and anti-inflammatory properties of curcumin," in *The molecular targets and therapeutic uses of curcumin in health and disease*, pp. 105–125, Springer, 2007.
- [73] Y. K. Han, S. H. Lee, H. J. Jeong, M. S. Kim, M. H. Yoon, and W. M. Kim, "Analgesic effects of intrathecal curcumin in the rat formalin test," *The Korean journal of pain*, vol. 25, no. 1, pp. 1–6, 2012.
- [74] Y. Wang, Z. Lu, H. Wu, and F. Lv, "Study on the antibiotic activity of microcapsule curcumin against foodborne pathogens," *International journal of food microbiology*, vol. 136, no. 1, pp. 71–74, 2009.
- [75] S. Daniel, J. L. Limson, A. Dairam, G. M. Watkins, and S. Daya, "Through metal binding, curcumin protects against lead- and cadmium-induced lipid peroxidation in rat brain homogenates and against lead-induced tissue damage in rat brain," *Journal of inorganic biochemistry*, vol. 98, no. 2, pp. 266–275, 2004.
- [76] A. Bielak-Zmijewska, W. Grabowska, A. Ciolko et al., "The role of curcumin in the modulation of ageing," *International journal of molecular sciences*, vol. 20, no. 5, p. 1239, 2019.
- [77] E. Balogun, M. Hoque, P. Gong et al., "Curcumin activates the haem oxygenase-1 gene via regulation of Nrf2 and the antioxidant-responsive element," *Biochemical Journal*, vol. 371, no. 3, pp. 887–895, 2003.
- [78] C. T. Tu, Q.-y. Yao, B.-l. Xu, J.-y. Wang, C.-h. Zhou, and S.-c. Zhang, "Protective effects of curcumin against hepatic fibrosis induced by carbon tetrachloride: modulation of high-mobility group box 1, toll-like receptor 4 and 2 expression," *Food and chemical toxicology*, vol. 50, no. 9, pp. 3343–3351, 2012.
- [79] C. F. Lima, C. Pereira-Wilson, and S. I. Rattan, "Curcumin induces heme oxygenase-1 in normal human skin fibroblasts through redox signaling: relevance for anti-aging intervention," *Molecular nutrition & food research*, vol. 55, no. 3, pp. 430–442, 2011.
- [80] J. R. Mein, D. R. James, and S. Lakkanna, "Induction of phase 2 antioxidant enzymes by broccoli sulforaphane: perspectives in maintaining the antioxidant activity of vitamins A, C, and E," *Frontiers in genetics*, vol. 3, p. 7, 2012.
- [81] A. I. Rojo, O. N. Medina-Campos, P. Rada et al., "Signaling pathways activated by the phytochemical nordihydroguaiaretic acid contribute to a Keap1-independent regulation of Nrf2 stability: role of glycogen synthase kinase-3," *Free Radical Biology and Medicine*, vol. 52, no. 2, pp. 473–487, 2012.
- [82] H. Cherif, D. G. Bisson, P. Jarzem, M. Weber, J. A. Ouellet, and L. Haglund, "Curcumin and o-vanillin exhibit evidence of senolytic activity in human IVD cells in vitro," *Journal of Clinical Medicine*, vol. 8, no. 4, p. 433, 2019.

- [83] W. Li, Y. He, R. Zhang, G. Zheng, and D. Zhou, "The curcumin analog EF24 is a novel senolytic agent," *Aging*, vol. 11, no. 2, pp. 771–782, 2019.
- [84] M. C. Fadus, C. Lau, J. Bikhchandani, and H. T. Lynch, "Curcumin: an age-old anti-inflammatory and anti-neoplastic agent," *Journal of traditional and complementary medicine*, vol. 7, no. 3, pp. 339–346, 2017.
- [85] K. Chandrashekar, S. Popli, and M. Shakarad, "Curcumin enhances parental reproductive lifespan and progeny viability in *Drosophila melanogaster*," *Age*, vol. 36, no. 5, p. 9702, 2014.
- [86] P. Sarvalkar, M. Walvekar, and L. Bhopale, "Antioxidative effect of curcumin (*Curcuma longa*) on lipid peroxidation and lipofuscinogenesis in submandibular gland of D-galactose-induced aging male mice," *Journal of Medicinal Plants Research*, vol. 5, no. 20, pp. 5191–5193, 2011.
- [87] B. S. Fleenor, A. L. Sindler, N. K. Marvi et al., "Curcumin ameliorates arterial dysfunction and oxidative stress with aging," *Experimental gerontology*, vol. 48, no. 2, pp. 269–276, 2013.
- [88] N. Akazawa, Y. Choi, A. Miyaki et al., "Curcumin ingestion and exercise training improve vascular endothelial function in postmenopausal women," *Nutrition research*, vol. 32, no. 10, pp. 795–799, 2012.
- [89] W. Wongcharoen and A. Phrommintikul, "The protective role of curcumin in cardiovascular diseases," *International journal of cardiology*, vol. 133, no. 2, pp. 145–151, 2009.
- [90] N. M. Bressler, "Age-related macular degeneration is the leading cause of blindness," *Journal of the American Medical Association*, vol. 291, no. 15, pp. 1900–1901, 2004.
- [91] M. Zhuang, J. Shao, C. Tan, and Y. Yao, "Effects of transthyretin on biological behavior of retinal pigment epithelial cells and retinal microvascular epithelial cells," [*Zhonghua yan ke za zhi*] *Chinese journal of ophthalmology*, vol. 52, no. 11, pp. 856–860, 2016.
- [92] W. Zhu, Y. Wu, Y.-F. Meng et al., "Effect of curcumin on aging retinal pigment epithelial cells," *Drug design, development and therapy*, vol. 9, p. 5337, 2015.
- [93] S. Sundar Dhilip Kumar, N. Houreld, and H. Abrahamse, "Therapeutic potential and recent advances of curcumin in the treatment of aging-associated diseases," *Molecules*, vol. 23, no. 4, p. 835, 2018.
- [94] P. A. Kumar, P. Suryanarayana, P. Y. Reddy, and G. B. Reddy, "Modulation of alpha-crystallin chaperone activity in diabetic rat lens by curcumin," *Molecular Vision*, vol. 11, pp. 561–568, 2005.
- [95] P. Suryanarayana, M. Saraswat, T. Mrudula, T. P. Krishna, K. Krishnaswamy, and G. B. Reddy, "Curcumin and turmeric delay streptozotocin-induced diabetic cataract in rats," *Investigative ophthalmology & visual science*, vol. 46, no. 6, pp. 2092–2099, 2005.
- [96] R.-L. Chen, J. S. Balami, M. M. Esiri, L.-K. Chen, and A. M. Buchan, "Ischemic stroke in the elderly: an overview of evidence," *Nature Reviews Neurology*, vol. 6, no. 5, pp. 256–265, 2010.
- [97] H. K. Shin, P. B. Jones, M. Garcia-Alloza et al., "Age-dependent cerebrovascular dysfunction in a transgenic mouse model of cerebral amyloid angiopathy," *Brain*, vol. 130, no. 9, pp. 2310–2319, 2007.
- [98] V. Vasilevko, G. Passos, D. Quiring et al., "Aging and cerebrovascular dysfunction: contribution of hypertension, cerebral amyloid angiopathy, and immunotherapy," *Annals of the New York Academy of Sciences*, vol. 1207, no. 1, pp. 58–70, 2010.
- [99] Y.-W. C. Fridell, A. Sánchez-Blanco, B. A. Silvia, and S. L. Helfand, "Targeted expression of the human uncoupling protein 2 (hUCP2) to adult neurons extends life span in the fly," *Cell metabolism*, vol. 1, no. 2, pp. 145–152, 2005.
- [100] W. Winder, B. Holmes, D. Rubink, E. Jensen, M. Chen, and J. Holloszy, "Activation of AMP-activated protein kinase increases mitochondrial enzymes in skeletal muscle," *Journal of Applied Physiology*, vol. 88, no. 6, pp. 2219–2226, 2000.
- [101] R. M. Reznick, H. Zong, J. Li et al., "Aging-associated reductions in AMP-activated protein kinase activity and mitochondrial biogenesis," *Cell metabolism*, vol. 5, no. 2, pp. 151–156, 2007.
- [102] A. Salminen and K. Kaarniranta, "AMP-activated protein kinase (AMPK) controls the aging process via an integrated signaling network," *Ageing research reviews*, vol. 11, no. 2, pp. 230–241, 2012.
- [103] Y. Pu, H. Zhang, P. Wang et al., "Dietary curcumin ameliorates aging-related cerebrovascular dysfunction through the AMPK/uncoupling protein 2 pathway," *Cellular Physiology and Biochemistry*, vol. 32, no. 5, pp. 1167–1177, 2013.
- [104] M. Tian, X. Zhang, L. Wang, and Y. Li, "Curcumin induces ABCA1 expression and apolipoprotein A-I-Mediated cholesterol transmembrane in the chronic cerebral hypoperfusion aging rats," *The American journal of Chinese medicine*, vol. 41, no. 5, pp. 1027–1042, 2013.
- [105] K. Reeta, J. Mehla, and Y. K. Gupta, "Curcumin is protective against phenytoin-induced cognitive impairment and oxidative stress in rats," *Brain research*, vol. 1301, pp. 52–60, 2009.
- [106] I.-J. Su, H.-Y. Chang, H.-C. Wang, and K.-J. Tsai, "A curcumin analog exhibits multiple biologic effects on the pathogenesis of Alzheimer's disease and improves behavior, inflammation, and β -amyloid accumulation in a mouse model," *International journal of molecular sciences*, vol. 21, no. 15, p. 5459, 2020.
- [107] T. Benameur, R. Soleti, M. A. Panaro et al., "Curcumin as prospective anti-aging natural compound: focus on brain," *Molecules*, vol. 26, no. 16, p. 4794, 2021.
- [108] J. Lee, Y. S. Kim, E. Kim, Y. Kim, and Y. Kim, "Curcumin and hesperetin attenuate D-galactose-induced brain senescence in vitro and in vivo," *Nutrition Research and Practice*, vol. 14, no. 5, pp. 438–452, 2020.
- [109] M. Belviranlı, N. Okudan, K. Atalık, and M. Öz, "Curcumin improves spatial memory and decreases oxidative damage in aged female rats," *Biogerontology*, vol. 14, no. 2, pp. 187–196, 2013.
- [110] Y.-F. Cheng, L. Guo, Y.-S. Xie et al., "Curcumin rescues aging-related loss of hippocampal synapse input specificity of long term potentiation in mice," *Neurochemical Research*, vol. 38, no. 1, pp. 98–107, 2013.
- [111] M. Taghizadeh, S. A. Talaei, A. Djazayeri, and M. Salami, "Vitamin D supplementation restores suppressed synaptic plasticity in Alzheimer's disease," *Nutritional neuroscience*, vol. 17, no. 4, pp. 172–177, 2014.
- [112] M. A. Olesen, A. K. Torres, C. Jara, M. P. Murphy, and C. Tapia-Rojas, "Premature synaptic mitochondrial dysfunction in the hippocampus during aging contributes to memory loss," *Redox Biology*, vol. 34, p. 101558, 2020.

- [113] M. S. Grotewiel, I. Martin, P. Bhandari, and E. Cook-Wiens, "Functional senescence in *Drosophila melanogaster*," *Ageing research reviews*, vol. 4, no. 3, pp. 372–397, 2005.
- [114] S. Libert, J. Zwiener, X. Chu, W. VanVoorhies, G. Roman, and S. D. Pletcher, "Regulation of *Drosophila* life span by olfaction and food-derived odors," *Science*, vol. 315, no. 5815, pp. 1133–1137, 2007.
- [115] J. M. Toivonen and L. Partridge, "Endocrine regulation of aging and reproduction in *Drosophila*," *Molecular and cellular endocrinology*, vol. 299, no. 1, pp. 39–50, 2009.
- [116] M. E. Giannakou and L. Partridge, "Role of insulin-like signalling in *Drosophila* lifespan," *Trends in biochemical sciences*, vol. 32, no. 4, pp. 180–188, 2007.
- [117] F. Demontis and N. Perrimon, "FOXO/4E-BP Signaling in *Drosophila* Muscles Regulates Organism-wide Proteostasis during Aging," *Cell*, vol. 143, no. 5, pp. 813–825, 2010.
- [118] J. Karpac, A. Younger, and H. Jasper, "Dynamic coordination of innate immune signaling and insulin signaling regulates systemic responses to localized DNA damage," *Developmental cell*, vol. 20, no. 6, pp. 841–854, 2011.
- [119] T. Osterwalder, K. S. Yoon, B. H. White, and H. Keshishian, "A conditional tissue-specific transgene expression system using inducible GAL4," *Proceedings of the National Academy of Sciences*, vol. 98, no. 22, pp. 12596–12601, 2001.
- [120] T. Lee and L. Luo, "Mosaic analysis with a repressible cell marker (MARCM) for *Drosophila* neural development," *Trends in neurosciences*, vol. 24, no. 5, pp. 251–254, 2001.
- [121] M. R. Klass, "A method for the isolation of longevity mutants in the nematode *Caenorhabditis elegans* and initial results," *Mechanisms of ageing and development*, vol. 22, no. 3-4, pp. 279–286, 1983.
- [122] G. Taormina, F. Ferrante, S. Vieni, N. Grassi, A. Russo, and M. G. Mirisola, "Longevity: lesson from model organisms," *Genes*, vol. 10, no. 7, p. 518, 2019.
- [123] D. B. Friedman and T. E. Johnson, "A mutation in the age-1 gene in *Caenorhabditis elegans* lengthens life and reduces hermaphrodite fertility," *Genetics*, vol. 118, no. 1, pp. 75–86, 1988.
- [124] D. B. Friedman and T. E. Johnson, "Three mutants that extend both mean and maximum life span of the nematode, *Caenorhabditis elegans*, define the age-1 gene," *Journal of gerontology*, vol. 43, no. 4, pp. B102–B109, 1988.
- [125] C. Kenyon, "The plasticity of aging: insights from long-lived mutants," *Cell*, vol. 120, no. 4, pp. 449–460, 2005.
- [126] S. S. Lee, S. Kennedy, A. C. Tolonen, and G. Ruvkun, "DAF-16 target genes that Control *C. elegans* Life-Span and metabolism," *science*, vol. 300, no. 5619, pp. 644–647, 2003.
- [127] C. T. Murphy, S. A. McCarroll, C. I. Bargmann et al., "Genes that act downstream of DAF-16 to influence the lifespan of *Caenorhabditis elegans*," *Nature*, vol. 424, no. 6946, pp. 277–283, 2003.
- [128] M. Tatar, A. Bartke, and A. Antebi, "The endocrine regulation of aging by insulin-like signals," *Science*, vol. 299, no. 5611, pp. 1346–1351, 2003.
- [129] M. Kaeberlein, R. W. Powers, K. K. Steffen, E. A. Westman, D. Hu, and N. Dang, "Regulation of yeast replicative life span by TOR and Sch 9 in response to nutrients," *Science*, vol. 310, no. 5751, pp. 1193–1196, 2005.
- [130] D. E. Harrison, R. Strong, Z. D. Sharp et al., "Rapamycin fed late in life extends lifespan in genetically heterogeneous mice," *Nature*, vol. 460, no. 7253, pp. 392–395, 2009.
- [131] Y. Li, W.-J. Wang, H. Cao et al., "Genetic association of FOXO1A and FOXO3A with longevity trait in Han Chinese populations," *Human molecular genetics*, vol. 18, no. 24, pp. 4897–4904, 2009.
- [132] C. V. Anselmi, A. Malovini, R. Roncarati et al., "Association of the FOXO3A Locus with extreme longevity in a southern Italian centenarian study," *Rejuvenation research*, vol. 12, no. 2, pp. 95–104, 2009.
- [133] M. Soerensen, S. Dato, K. Christensen et al., "Replication of an association of variation in the FOXO3A gene with human longevity using both case-control and longitudinal data," *Ageing Cell*, vol. 9, no. 6, pp. 1010–1017, 2010.
- [134] A. Brunet, L. B. Sweeney, J. F. Sturgill, K. F. Chua, P. L. Greer, and Y. Lin, "Stress-dependent regulation of FOXO transcription factors by the SIRT1 deacetylase," *science*, vol. 303, no. 5666, pp. 2011–2015, 2004.
- [135] G. J. Lithgow, M. S. Gill, A. Olsen, and J. N. Sampayo, "Pharmacological intervention in invertebrate aging," *Age*, vol. 27, no. 3, pp. 213–223, 2005.
- [136] H.-L. Kang, S. Benzer, and K.-T. Min, "Life extension in *Drosophila* by feeding a drug," *Proceedings of the National Academy of Sciences*, vol. 99, no. 2, pp. 838–843, 2002.
- [137] M. Jafari, J. S. Felgner, I. I. Bussel et al., "Rhodiola: a promising anti-aging Chinese herb," *Rejuvenation research*, vol. 10, no. 4, pp. 587–602, 2007.
- [138] A. Pisaruk, N. Koshel, L. Mekhova, O. Zabuga, and S. Ivanov, "Influence of curcumin on lifespan if it is applied at the larval stage of *Drosophila melanogaster*," *Ageing and longevity*, vol. 1, no. 2, pp. 89–96, 2020.
- [139] K.-S. Lee, B.-S. Lee, S. Semnani et al., "Curcumin extends life span, improves health span, and modulates the expression of age-associated aging genes in *Drosophila melanogaster*," *Rejuvenation Research*, vol. 13, no. 5, pp. 561–570, 2010.
- [140] T.-S. Huang, S.-C. Lee, and J.-K. Lin, "Suppression of c-Jun/AP-1 activation by an inhibitor of tumor promotion in mouse fibroblast cells," *Proceedings of the National Academy of Sciences*, vol. 88, no. 12, pp. 5292–5296, 1991.
- [141] L. Korutla and R. Kumar, "Inhibitory effect of curcumin on epidermal growth factor receptor kinase activity in A431 cells," *Biochimica et Biophysica Acta (BBA) - Molecular Cell Research*, vol. 1224, no. 3, pp. 597–600, 1994.
- [142] K. M. Seong, M. Yu, K.-S. Lee, S. Park, Y. W. Jin, and K.-J. Min, "Curcumin mitigates accelerated aging after irradiation in *Drosophila* by reducing oxidative stress," *Bio Med research international*, vol. 2015, pp. 1–8, 2015.
- [143] L.-R. Shen, F. Xiao, P. Yuan et al., "Curcumin-supplemented diets increase superoxide dismutase activity and mean life-span in *Drosophila*," *Age*, vol. 35, no. 4, pp. 1133–1142, 2013.
- [144] J.-W. Soh, N. Marowsky, T. J. Nichols et al., "Curcumin is an early-acting stage-specific inducer of extended functional longevity in *Drosophila*," *Experimental gerontology*, vol. 48, no. 2, pp. 229–239, 2013.
- [145] V. H.-C. Liao, C.-W. Yu, Y.-J. Chu, W.-H. Li, Y.-C. Hsieh, and T.-T. Wang, "Curcumin-mediated lifespan extension in *Caenorhabditis elegans*," *Mechanisms of ageing and development*, vol. 132, no. 10, pp. 480–487, 2011.
- [146] J. E. Kim, A. C. Shrestha, H. S. Kim et al., "WS-5 Extract of *Curcuma longa*, *Chaenomeles sinensis*, and *Zingiber officinale* Contains Anti-AChE Compounds and Improves Amyloid-Induced Memory Impairment in Mice," *Evidence-Based*

- Complementary and Alternative Medicine*, vol. 2019, 16 pages, 2019.
- [147] C. W. Wittmann, M. F. Wszolek, J. M. Shulman, P. M. Salvaterra, J. Lewis, and M. Hutton, "Tauopathy in *Drosophila*: neurodegeneration without neurofibrillary tangles," *Science*, vol. 293, no. 5530, pp. 711–714, 2001.
- [148] I. Caesar, M. Jonson, K. P. R. Nilsson, S. Thor, and P. Hammarström, "Curcumin promotes A-beta fibrillation and reduces neurotoxicity in transgenic *Drosophila*," *PLoS One*, vol. 7, no. 2, article e31424, 2012.
- [149] J. Hardy and D. J. Selkoe, "The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics," *science*, vol. 297, no. 5580, pp. 353–356, 2002.
- [150] Y.-H. Suh and F. Checler, "Amyloid precursor protein, presenilins, and alpha -Synuclein: molecular pathogenesis and pharmacological applications in Alzheimer's disease," *Pharmacological Reviews*, vol. 54, no. 3, pp. 469–525, 2002.
- [151] W. Grabowska, K. Kucharewicz, M. Wnuk et al., "Curcumin induces senescence of primary human cells building the vasculature in a DNA damage and ATM-independent manner," *Age*, vol. 37, no. 1, 2015.
- [152] W. Grabowska, M. Suszek, M. Wnuk et al., "Curcumin elevates sirtuin level but does not postpone vitrosenescence of human cells building the vasculature," *Oncotarget*, vol. 7, no. 15, pp. 19201–19213, 2016.
- [153] D. Banji, O. J. Banji, S. Dasaraju, and A. Annamalai, "Piperine and curcumin exhibit synergism in attenuating D-galactose induced senescence in rats," *European journal of pharmacology*, vol. 703, no. 1-3, pp. 91–99, 2013.
- [154] A. Kumar, A. Prakash, and S. Dogra, "Protective effect of curcumin (*Curcuma longa*) against d-galactose-induced senescence in mice," *Journal of Asian natural products research*, vol. 13, no. 1, pp. 42–55, 2011.
- [155] J.-h. Li, T.-t. Wei, L. Guo et al., "Curcumin protects thymus against D-galactose-induced senescence in mice," *Naunyn-Schmiedeberg's Archives of Pharmacology*, vol. 394, no. 2, pp. 411–420, 2021.
- [156] C. Y. Sun, S. S. Qi, P. Zhou et al., "Neurobiological and pharmacological validity of curcumin in ameliorating memory performance of senescence-accelerated mice," *Pharmacology Biochemistry and Behavior*, vol. 105, pp. 76–82, 2013.
- [157] K. Iijima, H.-P. Liu, A.-S. Chiang, S. A. Hearn, M. Konsolaki, and Y. Zhong, "Dissecting the pathological effects of human A40 and A42 in *Drosophila*: a potential model for Alzheimer's disease," *Proceedings of the National Academy of Sciences*, vol. 101, no. 17, pp. 6623–6628, 2004.
- [158] M. Okuda, Y. Fujita, I. Hijikuro et al., "PE859, a novel curcumin derivative, inhibits amyloid- β and tau aggregation, and ameliorates cognitive dysfunction in senescence-accelerated mouse prone 8," *Journal of Alzheimer's Disease*, vol. 59, no. 1, pp. 313–328, 2017.
- [159] Y. Sun, X. Hu, G. Hu, C. Xu, and H. Jiang, "Curcumin attenuates hydrogen peroxide-induced premature senescence via the activation of SIRT1 in human umbilical vein endothelial cells," *Biological and Pharmaceutical Bulletin*, vol. 38, no. 8, pp. 1134–1141, 2015.
- [160] K. Kitani, T. Osawa, and T. Yokozawa, "The effects of tetrahydrocurcumin and green tea polyphenol on the survival of male C57BL/6 mice," *Biogerontology*, vol. 8, no. 5, pp. 567–573, 2007.
- [161] Y.-B. Li, Z.-F. Zhong, M.-W. Chen et al., "Bisdemethoxycurcumin Increases Sirt1 to Antagonize t-BHP-Induced Premature Senescence in WI38 Fibroblast Cells," *Evidence-Based Complementary and Alternative Medicine*, vol. 2013, article 851714, pp. 1–9, 2013.
- [162] K. Takano, J. Tatebe, N. Washizawa, and T. Morita, "Curcumin inhibits age-related vascular changes in aged mice fed a high-fat diet," *Nutrients*, vol. 10, no. 10, p. 1476, 2018.
- [163] K. Li, M. Zhai, L. Jiang et al., "Tetrahydrocurcumin ameliorates diabetic cardiomyopathy by attenuating high glucose-induced oxidative stress and fibrosis via activating the SIRT1 pathway," *Oxidative medicine and cellular longevity*, vol. 2019, Article ID 6746907, 2019.
- [164] K. Kukkemann and A. Jagota, "Therapeutic effects of curcumin on age-induced alterations in daily rhythms of clock genes and Sirt 1 expression in the SCN of male Wistar rats," *Biogerontology*, vol. 20, no. 4, pp. 405–419, 2019.
- [165] S. H. Azami, H. Nazarian, M. A. Abdollahifar, F. Eini, M. A. Farsani, and M. G. Novin, "The antioxidant curcumin postpones ovarian aging in young and middle-aged mice," *Reproduction, Fertility and Development*, vol. 32, no. 3, pp. 292–303, 2020.
- [166] A. M. Selim, M. M. Nooh, M. M. El-Sawalhi, and N. A. Ismail, "Amelioration of age-related alterations in rat liver: Effects of curcumin C3 complex, Astragalus membranaceus and blueberry," *Experimental Gerontology*, vol. 137, p. 110982, 2020.
- [167] L. Liu, P. Zhang, Y. Li, and G. Yu, "Curcumin protects brain from oxidative stress through inducing expression of UCP2 in chronic cerebral hypoperfusion aging-rats," *Molecular neurodegeneration*, vol. 7, no. 1, 2012.
- [168] B. K. Suckow and M. A. Suckow, "Lifespan extension by the antioxidant curcumin in *Drosophila melanogaster*," *International journal of biomedical science: IJBS*, vol. 2, no. 4, p. 402, 2006.