Article type : Mini-Review

Ageing and longevity genes in cardiovascular diseases

Luca Liberale*^{1,2}, Simon Kraler*¹, Giovanni G. Camici^{1,3,4}, Thomas F. Lüscher^{1,5}

¹ Center for Molecular Cardiology, University of Zürich, Schlieren, Switzerland;

² First Clinic of Internal Medicine, Department of Internal Medicine, University of Genoa, Genoa, Italy;

³ University Heart Center, Department of Cardiology, University Hospital Zurich, Zurich, Switzerland;

⁴ Department of Research and Education, University Hospital Zurich, Zurich, Switzerland;

⁵ Heart Division, Royal Brompton and Harefield Hospitals and National Heart and Lung Institute, Imperial College, London, United Kingdom.

*these authors contributed equally to this article.

Correspondence to: Thomas F. Lüscher. Center for Molecular Cardiology, University of Zurich, Wagistrasse 12, Schlieren CH-8952, Switzerland. E-mail address: thomas.luescher@zhh.ch, phone: +41 44 250 40 97, fax: +41 44 635 68 27. Article

Article

¹ Cente

² First

¹ Cente

² First

¹ Lente

² First

¹ Depa

⁵ Hear

⁵ Hear

⁵ Hear

⁵ Hear

⁴ Depa

⁵ Hear

⁴ Depa

⁵ Hear

⁴ Depa

⁵ Hear

⁴ Depa

⁵ Hear

phone:

Running title: ageing and longevity genes in CV function.

Keywords: ageing, cardiovascular diseases, cardiovascular ageing, oxidative stress, inflammation.

This article has been accepted for publication and undergone full peer review but has not been through the copyediting, typesetting, pagination and proofreading process, which may lead to differences between this version and the [Version of Record.](https://doi.org/10.1111/BCPT.13426) Please cite this article as [doi:](https://doi.org/10.1111/BCPT.13426) [10.1111/BCPT.13426](https://doi.org/10.1111/BCPT.13426)

Abstract

Over the last century, Western societies experienced a demographic shift driven by increased lifespan and decreased fertility, resulting in the subversion of the world's demographic pyramid. In ageing societies, cardiovascular diseases are the major cause of morbidity and mortality, thus representing a major societal and economic burden. Indeed, ageing associates with the deterioration of a genetic network implicated in senescence and longevity, orchestrating deleterious cellular processes that converge in the structural and functional decline of both the myocardium and the vasculature. In this review, we revise a compendium of genes involved in these processes and delineate possible strategies to interfere with them. Dietary interventions (e.g. intermittent fasting) and sirtuin-activating compounds are among the most promising interventions shown to promote protective effects on the ageing cardiovascular system. We conclude that ageing and longevity genes modulate cardiovascular function by acting on deleterious downstream processes such as inflammation and oxidative stress, thus representing promising targets for the prevention and treatment of age-related cardiovascular dysfunction.

Example 1
 Accepted Article
 Accepted Article
 Accepted Article
 Accepted Article
 Accepted Articles
 Accepted Articles
 Accepted Articles
 Accepted Articles
 Accepted Articles
 Accepted Articles

Introduction

Advances of contemporary medicine critically contributed to extend life expectancy; as a result worldwide population is ageing at a unprecedented pace [1]. People over 65 years of age (currently the elderly definition) will be expected to represent one quarter of total population by 2050. Of interest, the oldest segments of the 65-plus population have dramatically grown and centenarians have more than doubled from 2.9 every 10,000 over-65 individuals in 1990 to currently 7.4/10,000. Also, projections foresee a further increment to 23.6/10,000 by 2050 [2]. Despite being extremely positive from an individual perspective, these demographic data raise major concerns for social and health care systems. Indeed, ageing associates with a progressive decline in physiological function of several organs and in line with this, the elderly represent a high-risk population burdened with high morbidity and disability [3].

Ageing is the most important determinant of lifespan, and in rodents ageing *per se* is associated with a decline in cardiac and vascular function even without risk factors suggesting that a genetically determined biological clock governs lifespan and does so by promoting adverse changes causing organ dysfunction [4]. Yet, few insights are available with respect to the molecular pathways determining age-dependent cardiovascular (CV) and cerebrovascular (CBV) diseases and this is also reflected by the lack of direct effective therapies. Ageing, vascular dysfunction and age-dependent CV and CBV disease are mediated by similar molecular mechanisms which predominantly involve production of free radicals and surge of inflammation [5]. Indeed, several animal and human studies showed an association between ischaemic stroke, myocardial infarction and increased systemic and local production of reactive oxygen species (ROS) and inflammatory mediators [6,7]. Of interest, ageing associates with a chronic dysfunctional activation of the inflammatory system occurring in the absence of appropriate infective signals known as inflamm-ageing and thought to be involved in different chronic agerelated afflictions [8]. Different genes regulating life span and modulating ageing (i.e. ageing and longevity genes) are major regulator of oxidative stress and inflammation. Accordingly, some of these genes are critically involved in the development of age-dependent CV and CBV diseases as well as being effective molecular targets for their treatment [9] (Figure 1). This review article builds on previously published articles from our research group to delineate a compendium of promising genes implicated in ageing and age-related CV diseases. Moreover, it discusses their potential as targets for future therapeutic interventions. Advan

worldv

(currer

2050.

centen

current

Despit

major

decline

high-ri

Ageing

with a

genetic

change

molecu

disease

dysfun

mecha

[5]. In

myoca

(ROS)

dysfun

infecti

related

longev

these g

well a

bu

Ageing genes

The Adaptor Protein p66^{Shc}: The adaptor protein p66^{Shc} is considered to play a key role in driving cellular senescence by regulating mitochondrial reactive oxygen species (ROS) formation and thus, cellular apoptosis [10]. Along with $p46^{Shc}$ and $p52^{Shc}$, $p66^{Shc}$ is encoded by the human ShcA locus with major roles in different cellular signalling pathways [11]. Mechanistically, p66^{Shc} translocates into the mitochondrion upon protein kinase C-β-mediated Ser36 phosphorylation. Mitochondrial p66^{Shc} initiates the oxidation of cytochrome c, thus resulting in enhanced hydrogen peroxide production and increased oxidative stress, eventually leading to the activation of proapoptotic pathways [10,12,13]. With ageing, vascular ROS production increases, thus reducing protective nitric oxide (NO) bioavailability and leading to endothelial dysfunction and CV disease $[14,15]$. In accordance with its master role in regulating ROS production, tissue levels of $p66^{Shc}$ are increased in vessels from elderly patients as compared to young individuals [16]. Hence, p66Shc gained increasing attention as a promising molecular target to reduce the burden of age and age-related CV diseases [10]. Indeed, mice lacking *p66Shc* show an increased lifespan of up to 30% which goes along with decreased levels of intracellular ROS and cellular resistance to paraquatinduced oxidative stress [17]. Notably, *p66Shc-/-* mice also exhibit preserved endothelial NO bioavailability and stable aortic superoxide levels during the course of ageing, suggesting that these animals are protected from age-dependent endothelial dysfunction mediated by oxidative stress [18]. In line with the recognized role of dysfunctional endothelium as *primum movens* of atherosclerosis [19], the expression and activity of p66Shc positively correlates with different CV risk factors (i.e. hyperglycaemia, hyperlipidaemia, and hyperhomocysteinaemia) in humans [20– 22]. The diameter of the retinal microvasculature has been proposed to serve as a marker of vascular health and, being associated with CV risk. Recently, wider retinal arterioles and narrower venules – indicators of good vascular health – have been reported to associate with blunted *p66Shc* mononuclear cell expression and lower levels of oxidative stress markers in healthy individuals being physically active as compared to sedentary subjects (with or without CV risk factors). Of note, these differences have been linked to hypomethylation of the *p66Shc* promoter suggesting once more epigenetics as an important modulator of vascular ageing [21]. In accordance with its association with CV risk factors, p66^{Shc} has been shown to be implicated in atherosclerotic plaque formation. Napoli *et al.* [22] found that high-fat fed mice lacking p66^{Shc} are less susceptible to develop early atherosclerotic lesions while exhibiting reduced levels of systemic oxidative stress. Agein_H

The Accellula

thus, c

clocus

translo

Mitoch

peroxic

apopto

protect

[14,15]

are inc

p66^{She}

age-rel

which

induce

bioava

these age-rel

stress

afteros

risk fa

22]. T

vascul:

venule

monon

bein

The detrimental role of p66^{Shc} was further investigated using male mice deficient for ApoE and p66Shc, revealing that p66Shc-deprivation attenuates atherosclerotic plaque formation also in advanced stages [23]. As a further step, the putative role of p66^{Shc} has been investigated in the tremendous clinical sequelae of advanced atherosclerotic lesions, such as myocardial infarction and ischemic stroke. Of interest, monocytes of patients presenting with acute coronary syndrome (i.e. either ST-elevation or non-ST-elevation myocardial infarction) showed augmented *p66Shc* transcript levels, suggesting a potential role of this protein in myocardial ischaemia [24]. Despite an early preclinical study employing a non-recirculating Langendorff approach suggested *p66Shc-/* hearts to be less susceptible to ischemia-reperfusion (I/R) injury [25], transient *in vivo* ligation of the left anterior coronary artery in *p66Shc-/-* mice resulted in increased myocardial injury due to blunted activation of protective pathways (i.e. Akt and Stat 3) thus increasing mitochondrial swelling and cellular apoptosis [26]. In the brain, on the other hand, mounting evidence points towards a detrimental role of p66Shc in the setting of I/R injury. Of interest, *p66Shc-/-* mice subjected to transient middle cerebral artery occlusion exhibit reduced stroke size and blunted neurological dysfunction after stroke, which is paralleled by attenuated ROS production locally and systemically [27]. Intriguingly, these effects are also present when small interfering RNA targeting p66Shc is injected intravenously after the ischaemic event upon reperfusion, an approach with high translational value. In line with the above, peripheral blood monocytes obtained from patients with ischaemic stroke display transiently increased *p66Shc* mRNA levels which correlate with shortterm neurological outcome if thrombolytic treatment is initiated [28]. p66^{She}, advanted trement and ise (i.e. ei transcription and ise (i.e. ei transcription and ise (i.e. ei transcription and ise of the left blunted swelling toward to tran dysfun system p66^{She} transla ischaer term n Mito

Mitochondrial ROS formation is actively involved in hypertension-induced vascular dysfunction where cyclic stretch stress disrupts the cellular redox balance and impairs the endothelial homeostasis [29]. Accordingly, human aortic endothelial cells exposed to pathologic cyclic stress exhibit enhanced p66^{Shc} phosphorylation, ROS production and blunted NO bioavailability [30]. Similarly, aortic endothelial cells isolated from spontaneously hypertensive rats show augmented Ser36 phosphorylation of p66^{Shc}, which associates with enhanced superoxide production in aortic homogenates [30]. At the cardiac level, long-standing hypertension-related pressure overload causes left-ventricular dysfunction and myocardial remodelling, leading to dilated cardiomyopathy and heart failure. Notably, recent evidence supports the notion that p66^{Shc} may represent an important nexus that orchestrates oxidative stress and cardiac dysfunction in a murine model of pressure overload-induced heart failure [31]. In patients with CV risk factors, Angiotensin-II

(AngII) is considered to be a critical mediator promoting cardiac remodelling and ageing. Notably, mice lacking p66Shc are protected against cardiac AngII-induced hypertrophic remodelling via reduced apoptotis [32].

Taken together, these data suggest that p66^{Shc} is an important player implicated in the regulation of CV (dys-)function in ageing and highlight its potential as a therapeutic target to reduce the burden of age-related CV diseases. Yet, given the different effects of p66Shc modulations in different organs, tissue selective inhibition would be required. While its short-term activation might protect the infarcted myocardium, p66Shc inhibition may reduce cerebral ischaemic injury and prevent endothelial dysfunction, atherosclerosis, and diabetic vascular disease. This complexity rose safety concern on the potential use of p66Shc modulators in the clinical setting and to date no approved therapeutic targets this gene.

Mammalian target of rapamycin: The mammalian target of rapamycin (mTOR) is a highly conserved serine/threonine kinase embedded in a sophisticated network governing cellular growth and metabolism upon nutrient-sensing and hormonal stimuli [33]. mTOR exerts its effects by forming two structurally and functionally distinct multiprotein complexes, namely mTORC1 and mTORC2, thus regulating different downstream processes [34]. These protein complexes emerged as critical players of CV integrity during embryonic development as well as in adulthood [34,35]. Indeed, while adult mice lacking cardiac-specific mTORC1 activity develop fatal dilated cardiomyopathy at baseline, these animals display impaired hypertrophic response and accelerated heart failure progression upon pressure-induced overload stress [36]. Similarly, deprivation of mTORC2 in the murine heart via *RPTOR Independent Companion Of MTOR Complex 2* (*Rictor)* deletion causes cardiomyocyte death and promotes left-ventricular dilation and fibrosis, resulting in profound cardiac dysfunction [37]. On the opposite, partial genetic or pharmacological inhibition (through rapamycin) of mTORC1 has consistently been linked to longevity, and confers protection against multiple age-related CV diseases by modulating various downstream pathways, including autophagy, oxidative stress and inflammation [36,38–40]. While mTORC2-activity is less sensitive to rapamycin treatment and requires long-term administration to be effectively blocked, mTORC1 activity was shown to be immediately inhibited following acute application [40]. Traditionally being used as an immunosuppressant, beneficial effects of rapamycin on CV function were first described in renal allograft patients withdrawn from cyclosporine, in which add-on treatment with rapamycin resulted in lower blood pressure levels despite the use of less mice 1

reduce

Taken

CV (d<u>y</u>

of age

organs

the inf

endoth

concer

therape
 Mamm

conser

and m

formin

mTOR

as criticand article

cardion

heart 1

mTOR

deletio

in pro

in pro

in pro

in pro

in linkibit

pr

antihypertensive drugs [41]. Likewise, age-related mTORC1 activation in mice is thought to be involved in the development of endothelial dysfunction via increased ROS formation and blunted NO bioavailability, effects partially reversed by rapamycin treatment [15,42,43]. With age, arterial walls lose elastic tissue in favour of collagen deposition, a process termed arterial stiffening, which is characterized by increased pulse wave velocity, representing a strong predictor of CV events [44]. Notably, Lesniewski *et al.* observed attenuated arterial stiffening in mice upon treatment with dietary rapamycin via decreased vascular collagen content, while content of elastin did not differ between groups [43]. In line with this notion, kidney transplant recipients whose therapy regimen is switched to rapamycin display improved carotid-femoral pulse-wave velocity, proposing mTOR-inhibition as a potential therapeutic strategy against arterial stiffening in selected patients [45]. Besides, Flynn *et al.* reported lately that mTOR-inhibition through rapamycin exerts beneficial effects on cardiac function and is capable to prevent age-related hypertrophic remodelling under unstressed conditions, even if administered late in life, suggesting a protective role independent of arterial stiffening [39]. In this context, low-grade sterile inflammation (i.e. "*inflamm-ageing"*) is a hallmark of ageing deeply linked to various age-related CV diseases, including atherosclerosis [46,47]. Very recently, the endothelial-specific deletion of the endogenous mTORC-1 inhibitor PRAS40 was reported to promote atherogenesis in atherosclerosis-prone animals via induction of pro-inflammatory signalling [48]. Despite several lines of evidence proposing an important role for mTORC1 in the modulation of inflammation, in a recent study rapamycin administration failed to ameliorate inflamm-ageing markers in a mouse model of accelerated ageing despite confirming its protective role on frailty and health span in general [49]. Yet, mTORC1 modulation through rapamycin or derivatives has been proven effective in reducing experimental atherosclerosis, and these drugs are currently applied locally in daily clinical practice to reduce arterial restenosis after stenting [49,50]. **Access Transferred Article**
 Access Transferred Article
 Access Scheef Expansion
 Access Scheef Expansion
 Access Transferred Article Article Scheef Article Scheef Article Scheef Article Scheef Article Scheef Artic

Albeit a bulk of evidence suggests a beneficial role of partial mTOR inhibition in maintaining CV health and integrity during the course of ageing, studies assessing selective blockade of mTORC signalling or its downstream targets (e.g. S6K1) are scarce. Thus, future studies are warranted to investigate the specific modulation of mTORC-1 and mTORC2-dependent pathways using genetic or pharmacological approaches.

Longevity genes

Sirtuins: The sirtuin family comprises seven proteins (SIRT1 through 7) governing nicotinamide adenine dinucleotide (NAD⁺)-dependent deacylase reactions (such as deacetylation, demalonylation, desuccinylation) and, therefore represent important players regulating key aspects of ageing [51]. In mammals, various genes acting in nutrient-sensing pathways, including *FOXO3* (for details see below) and *sirtuins*, have been identified as crucial regulators of CV senescence, and variants within these genes have been linked to exceptional longevity across numerous studies [52]. Of interest, most sirtuins have been shown to play important roles in the modulation of oxidative stress, inflammation, autophagy, cellular senescence, and apoptosis, processes deeply involved in the pathophysiology of CV diseases [53]. As vascular levels of NAD⁺ and sirtuin activity steadily decline during the course of ageing [54,55], unravelling their role in age-related CV afflictions is of particular interest. In *ApoE-/-* mice fed on a high-fat diet, endothelial cellspecific Sirt1 overexpression as well as the administration of its pharmacological activator SRT3025 dampens atherosclerosis [56,57]. Oppositely, Sirt1 inhibition associates with increased plaque formation in atherosclerosis-prone experimental models [56,57]. Notably, *ApoE-/-* mice display increased endothelial superoxide formation and enhanced NF-kB-dependent expression of adhesion molecules upon *Sirt1* deprivation, suggesting that *Sirt1* may protect from atherosclerosis by blunting inflammation and oxidative stress [58]. Similarly, while mice lacking *Sirt1* specifically in cardiomyocytes display exacerbated myocardial injury upon I/R injury, its cardiac overexpression associates with reduced infarct size via blunted apoptosis [59]. Likewise, *in vitro* studies found that the upregulation of the mitochondrial sirtuins (*Sirt3*, *4* and *5*, respectively) maintain cardiomyocyte structure and function in the setting of I/R, proposing a protective role of these proteins during I/R in the heart [60–62]. For example, *Sirt3* deficient animals show increased cardiac damage [63], profound coronary microvascular dysfunction as well as impaired remodelling upon myocardial ischaemia [64]. Additionally, sirtuins have been hypothesized to be implicated in the pathophysiology of cerebral I/R injury. Recently, we showed that mice lacking endothelial-specific *Sirt6* display increased infarct volumes, mortality and brain extravasation of large molecules following transient middle cerebral artery occlusion, indicating that SIRT6 plays a protective role in ischaemic stroke by preserving blood-brain barrier integrity [65]. We further showed that post-ischaemic SIRT6 overexpression reduces stroke size by 50% and ameliorates neurologic outcome, highlighting its potential as a therapeutic target in this context [65]. Confirmatory *in vitro* experiments employing primary human cerebral microvascular endothelial cells increased the translational relevance of our findings, which has been further strengthened by **Accession**
 Access definition
 Access definition
 Article
 Article
 Article
 Article
 Article
 Article
 Article
 Actes definition
 Actes definition
 Article
 Article
 Article
 Article
 Artic

the assessment of *Sirt6* expression in peripheral blood mononuclear cells (PBMCs) obtained from stroke patients. Of interest, we found that its expression in PBMCs closely correlates with stroke outcome [65]. On the contrary, we previously studied the detrimental role of *Sirt5* in the context of ischaemic stroke [66]. Specifically, our findings suggest that PBMCs and cerebrovascular tissue obtained from stroke patients and mice following experimental I/R brain injury, respectively, display enhanced Sirt5 gene expression. Likewise, SIRT5 gene deletion or silencing using smallinterfering RNA resulted in reduced infarct size, post-stroke neuromotor deficit and BBB permeability through blunted occludin degradation, effects reproduced *in vitro* in human brain microvascular endothelial cells exposed to hypoxia/reoxygenation [66].

Increased fibrosis, left-ventricular hypertrophy and dilation are hallmarks of cardiac ageing and key causes of heart failure. Notably, SIRT3 levels steadily decline during hypertrophic remodelling irrespective of the model used [67,68]. Accordingly, ageing mice deficient for cardiac-specific *Sirt3* develop cardiac hypertrophy and fibrosis, while its overexpression protects from hypertrophic stimuli, at least in part, by enhanced FOXO3a activation [69]. A bulk of evidence suggests that the effects mediated by SIRT1 and SIRT3 may be also beneficial in the setting of hypertension, a potent risk factor for numerous CV diseases. Indeed, in mice with Angiotensin-II-induced hypertension, *Sirt1* overexpression alleviates vascular remodelling, abrogates ROS formation and exerts antihypertensive effects [70]. Furthermore, mice lacking *Sirt3* develop spontaneous pulmonary arterial hypertension (PAH) which associates with aberrant mitochondrial function. Interestingly, in rats with monocrotaline-induced PAH, disease phenotype can be reversed upon the rescue of *Sirt3* deficiency using adenovirus gene therapy, implying a causative role of SIRT3 in PAH. In line with this notion, a loss-of-function SIRT3 polymorphism associates with PAH in humans, which underscores the protective role of this protein in PAH [71]. On the contrary, the role of Sirt5 in the setting of cardiac hypertrophy remains to be fully characterized since opposite findings have been reported so far by using genetically deficient mice undergoing transverse aortic constriction [72,73]. stroke

outcon

ischaer

obtaine

display

interfe

permea

microv

Increas

key c

remode

cardiaα

from l

eviden

setting

Angiot

develo

mitoch

can be

causati

develo

mitoch

mitoch

mitoch

mitoch

mitoch

mitoch

In summary, with a few exceptions these findings point towards a protective role of most sirtuins in atherosclerosis, cardiac I/R injury, cardiac remodelling, hypertension and stroke. Few exceptions need to be considered with respect to specific sirtuins, their targets, route of administration (i.e. systemic vs local) or dosage. These effects may, at least in part, be mediated

by the activation of different downstream pathways crucially involved in ageing and longevity (such as FOXO3 and $p66^{Shc}$).

Beclin 1: Macroautophagy (hereafter referred to as autophagy) is a cytoprotective process that maintains cardiac structure and function during the course of ageing [74]. Physiological (e.g. caloric restriction), pharmacological (e.g. spermidine) or genetic induction of this process promotes longevity and exerts cardioprotective effects across different species, including mice [75–77]. Accordingly, *Beclin 1* knock-down has been associated with defective autophagy and shortened lifespan [77]. Conversely, knock-in mice bearing a *Phe121Ala* mutation in *Beclin 1* (*Becn1F121A/F121A)* which reduces the interaction with the negative regulator Bcl-2 and results in enhanced autophagy, associates with blunted age-related structural changes of the heart (i.e. cardiac fibrosis) and improved health- and lifespan [78]. Of interest, *Beclin 1* overexpression exerts deleterious effects in mice subjected to thoracic aortic constriction, as evidenced by impaired systolic function and accentuated pathological remodelling, indicating that enhanced autophagy may be detrimental in the setting of pressure-related cardiac remodelling [79]. Furthermore, Razani *et al.* showed that Beclin 1 protein levels are not significantly altered in aortic lysates obtained from atherosclerosis-prone *ApoE-/-* mice, indicating that atherosclerotic plaque formation may not depend on upstream components of the autophagic pathway. Notably, the complete loss of macrophage autophagy, however, promotes compromised cholesterol efflux capacity and enhanced foam cell development, resulting in overt plaque formation, implying a protective role of basal autophagy in atherogenesis [80]. Hence, Beclin 1, among other proteins regulating autophagy (e.g. mTORC1), may represent a promising target for cardioprotective interventions during ageing. Yet, the effects of its modulation seem complex and might depend based on the disease and the magnitude of the intervention. More studies are required to investigate the specific role of Beclin 1 in the different age-related CV diseases. **Example 18**
 Accelin mainta
 Caloric promotor
 Acceling mainta
 Caloric promotor
 CALORIC 19
 CALORIC PROMOTER CONSERVING PURSUARY (Promotor)
 ACCELE 2013
 ACCELE PROMOTER CONSERVING PURSUARY EXPOSED EXPIDE

Klotho: So far, two isoforms (i.e. αKlotho and βKlotho) of the Klotho protein have been described, both important components of the fibroblast growth factor receptor complex. The constitutively-occurring ectodomain-shedding of αKlotho liberates a soluble form, which acts as a circulating hormone regulating a plethora of cellular processes including the inhibition of growth factors, such as insulin-like growth factor 1 (IGF1) and transforming growth factor β1 (TGFβ1) [81]. Interestingly, mice bearing a mutation in the *Klotho* gene display ageing-like phenotypes across different tissues with an average life-span of 60.7 days [82]. Conversely, *Klotho*

overexpression increases lifespan by 20-30% and alleviates ageing-like phenotypes [83]. Furthermore, it promotes cardioprotection [84], and blunts oxidative stress [85]. On the other side, *Klotho^{+/-}* mice display impaired aortic eNOS activity and attenuated endothelial SIRT1 expression, resulting in arterial stiffening and hypertension. Interestingly, treatment with a SIRT1 activator (i.e. SRT1720) was able to rescue the deleterious effect of *Klotho*-deprivation on vascular structure and function [86]. In line with the fact that mice partially defective for *Klotho* are more prone to develop atherosclerosis [82], humans bearing a functional variant for *KLOTHO* display an increased risk of early-onset coronary artery disease [87].

Forkhead Transcription Factors: The forkhead transcription factors (FOXOs) represent a subfamily of highly-conserved proteins regulating longevity by modulating insulin/IGF-1 signalling [88]. In humans, the gene encoding the transcription factor FOXO3 has been consistently linked to longevity [89,90]. In Drosophila, modest overexpression of *Foxo* exerts cardioprotective effects and ameliorates the age-related decline in cardiac function [91]. Accordingly, *Foxo3-/-* mice display a more pronounced hypertrophic response upon pressureinduced overload as compared to their control littermates. Likewise, cultured cardiomyocytes expressing *Foxo1* or *Foxo3* show attenuated Angiotensin-II induced hypertrophic growth, suggesting that *Foxo*s regulate cardiomyocyte growth [92]. On the other hand, overexpression of both, *Foxo1* and *Foxo3a* results in reduced eNOS protein expression. In line with this notion, aortic lysates obtained from *FOXO3a-/-* mice have decreased endothelial eNOS protein expression [93]. Yet, FOXO factors positively regulate the expression of key ROS-scavenging enzymes such as MnSOD (manganese superoxide dismutase), catalase, and GADD45 [94,95] and their inactivation associates with increased intracellular ROS and accelerated atherosclerosis [96]. Albeit accumulating evidence points towards a beneficial role of FOXOs within the longevity network, more research is warranted to understand the processes governed by these proteins in the CV system during health and disease. Furthe
 Klotho

resultin

(i.e. S

structu

prone

an incr
 Forkhe

subfan

signall

consist

cardioq

accerd induce

expres

sugges

both, *l*

aortic

[93]. Y

as Mr inactiv

Albeit

networ

CV sys

Agein_i

The cl

Ageing and longevity genes as targets to improve CV outcome: future perspectives

The clear pre-clinical evidence pointing at ageing and longevity genes as pivotal mediators of agedependent CV disease together with the global burden of an ever-ageing society call for great efforts in translating these results into effective clinical strategies. While the "*bench-to-bedside*" translation occurred smoothly for mTOR and its inhibitors, this is not the case for other potential therapeutic targets in which activity modulation in patients is not easy to obtain. Sirtuins represent a paradigmatic example and major advances have been obtained as different clinical trials evaluating sirtuin boosters showed protective effects in cardiovascular and metabolic diseases. Different approaches can be used to increase sirtuin activity, such as dietary interventions (e.g. fasting, intermittent fasting or ketogenic diet) and administration of NAD⁺ boosting or sirtuinactivating compounds (Small-Molecule Allosteric Activators of Sirtuins or STACs) [97–99] (Figure 2).

Beyond its fundamental role in redox balance, NAD⁺ is the essential substrate of sirtuin enzymatic activity. Accordingly, NAD⁺ was shown to play pivotal roles in cell signalling and increased cellular supplies of this mediator mimic caloric restriction and associate with stress resistance and prolonged lifespan [98]. Also, cellular NAD⁺ is significantly down-regulated in ageing while its supplementation associates with several protective effects on metabolism and CV function in different experimental models [55,100]. Besides the supplementation of NAD⁺ precursors, such as nicotinamide or nicotinamide mononucleotide (NMN), enzyme induction of the NAD⁺ synthesis pathways or blockage of NAD⁺ consuming pathways (i.e. inhibition of Poly [ADP-ribose] polymerase (PARP1) via INO-1001) may represent feasible strategies to increase intracellular NAD⁺ levels [100–102]. Among STACs, resveratrol and other natural phenols have been longterm described and characterized [103,104]. Despite being a potent activator of SIRT1 and having shown ability to extend lifespan in yeast [105], resveratrol as well as other phenols lacks specificity and can induce the activity of other SIRTs as well as non-sirtuin targets [106,107]. Then, different STACs have been generated with increased activity and specificity all acting as allosteric activator of SIRT1. Both NAD⁺ boosters and STACs have proven beneficial actions in different animal models of disease. Treatment with the SIRT1 activator SRT1720 was able to reverse ageing-associated endothelial dysfunction in mice through decreased oxidative stress and vascular inflammation [108]. Similarly, in monkeys, resveratrol treatment blunted arterial stiffening and inflammation. Treatment with NMN and other NAD⁺ boosters showed similar beneficial effects on vascular function in rat models of disease as well as increasing angiogenesis ability of the aged vasculature [109]. Increased cardiac NAD⁺ levels also showed protective features on I/R myocardial injury as well as being indicated as possible mediators of the beneficial effect of preconditioning [110,111]. Similarly, several studies suggest that NAD⁺ boosters may protect against hypertrophy and cardiac remodelling in several experimental models [112,113], while the role of STACs in this setting remain to be fully elucidated with opposite effects of the over activation of different sirtuin at different level being reported [114]. **Procedure Contains and Article**
 Accept Article
 Accept Article
 Accept Article
 Accept Article
 Accept Article
 Accept Article
 ACCEP Article
 ACCEP Article
 ACCEP Article
 ACCEP Article
 ACCEP Artic

Among STACs, resveratrol and SRT2104 have been investigated in several randomized clinical trials with metabolic or cardiovascular outcome. Although contrasting results have been so far reported by different clinical trials, treatment with resveratrol has shown modest beneficial evidence on metabolic parameters (e.g. insulin sensitivity, body weight and circulating lipids) in obese, diabetic and metabolic syndrome patients, while these effects were not reported in healthy people [115–118]. Of interest, in patients with coronary artery disease, resveratrol could improve endothelial and diastolic function [119]. Only few trials investigated NAD⁺ boosters as sirtuin activator in the contest of CV disease. NRPT, NR, NMN and INO-1001 showed good safety profiles [120–122] and promising preliminary results, many trials investigating their effect on heart failure, atherosclerosis and other CV outcome are currently underway [114].

Despite of the impressive amount of evidence, none of the approaches described above have so far translated into an effective strategy to counteract CV ageing and disease. Potentially divergent roles for different SIRTs in different CV conditions and the lack of specificity of the available pharmacological approaches might partly account for this discrepancy. Similarly, other antioxidants have failed in proving efficacy in large clinical trials. However, the lack of benefit of antioxidants in clinical trials does not disprove the central role of oxidative stress in cardiovascular ageing which is rather well-defined and proven by a plethora of experimental and clinical findings. Many explanations have been proposed to address this paradox: (i) the choice of antioxidants tested in current trials was mostly driven by their availability and safety profiles rather than their efficiency as antioxidants (e.g. vitamins); (ii) the trials may not have lasted long enough: oxidative stress acts on the vessels for decades, thus, months or a few years of antioxidant therapy may not be enough for reversing its deleterious effects; (iii) preventing ROS production as mostly done in the experimental setting rather than increase ROS scavenging by using non-specific antioxidants may lead to better results, particularly as ROS may sequester in cell compartments and are rapid in their mode of action. Thus, it might be more promising (i) to test genetic approaches (e.g. RNA interference, antisense technology) to prevent the formation of ROS, (ii) selecting an appropriate population with markedly increased oxidative stress and (iii) with long-term follow-up. Accordingly, understanding the molecular mechanisms regulating the age-dependent modulation of ageing and longevity genes is of utmost importance to generate novel specific and effective therapeutic approaches [5]. Standing at the crossroad between genetic background and environmental factors, epigenetic modifications have emerged as significant modulator of gene expression during the life course with crucial role on vascular and heart function [123]. A careful Fried and the expected of age theraption and the expected article in the expected of age theraption and the expected article in the expected article in the expected article in the expected article in the expected article i

description of the epigenetic changes characterizing the elderly might pave the way for novel strategies aiming at reprogramming the genetic maladaptive changes occurring with ageing thus reducing their burden on the CV system.

In recent years, senescence-associated secretory phenotype (SASP), a cellular process characterized by the production of extracellular modulators, such as pro-inflammatory cytokines, aroused increasing attention in ageing research [124]. Interestingly, novel findings indicate that Klotho suppresses the maladaptive activation of the endoplasmic reticulum and Golgi in senescent monocytes, which may, in turn affect the acquisition of SASP [125]. To our knowledge, however, so far no direct link between the genes previously outlined and SASP in the setting of CV diseases has been established, although a complex intersection among different axes, specifically NF-κBdependent signalling, may exist [126]. The term "senolytics" comprises a broad family of small molecules that selectively promote death of senescent cells and target a variety of subcellular pathways, including FOXO4-dependent signalling, and have been shown to be effective under conditions of accelerated ageing and may therefore represent a feasible strategy to maintain or even restore tissue homeostasis during ageing [127,128]. Albeit preliminary results on humans obtained during a clinical trial using a combination of senolytic drugs show a clear decline in adipose tissue senescent cells in subjects with diabetic kidney disease, large-scale studies are warranted to investigate their efficacy and safety in the setting of CV diseases [129,130]). strateg

reducin

In rec

charace

arouse

Klotho

monoc

so far i

has be

depend

molecu

pathwa

conditi

even r

obtains

adipos

warran

Conclu

By act

Longev

extens:

has er

of such

such of such

adipos

extens:

Conclusions

By acting on common cellular mechanisms such as oxidative stress and inflammation, ageing and longevity genes hold clear roles in the pathophysiology of different CV diseases. Specifically, extensive preclinical evidence exists for a protective role of most sirtuins and *Klotho*, while *p66Shc* has emerged as a potential mediator of CV dysfunction. Modulating the expression or the activity of such targets will be vital to prevent and treat age-dependent CV conditions.

Acknowledgement

GGC received support for this work from: the Swiss National Science Foundation [310030_175546], the Swiss Heart Foundation, the Alfred and Annemarie von Sick Grants for Translational and Clinical Research Cardiology and Oncology and the Foundation for Cardiovascular Research–Zurich Heart House. GGC is also the recipient of a Sheikh Khalifa's Foundation Ass. Professorship at the Faculty of Medicine, University of Zurich.

Conflict of interests

Accession Conflit Cardio
Franslic Cardio
Found:
None.
None. None. Acc

References

- 1. Camici GG, Liberale L. Aging: the next cardiovascular disease? Eur Heart J 2017;38:1621– 3. doi: 10.1093/eurheartj/ehx239
- 2. United Nations, Department of Economic and Social Affairs PD. World population ageing 2017 - highlights [Internet]. 2017 [cited 2020 Jan 15];
	- 3. Camici GG, Savarese G, Akhmedov A, Lüscher TF. Molecular mechanism of endothelial and vascular aging: implications for cardiovascular disease. Eur Heart J 2015;36:3392–403. doi: 10.1093/eurheartj/ehv587
- 4. North BJ, Sinclair DA. The intersection between aging and cardiovascular disease. Circ Res 2012;110:1097–108. doi: 10.1161/CIRCRESAHA.111.246876
- 5. Paneni F, Diaz Cañestro C, Libby P, Lüscher TF, Camici GG. The aging cardiovascular system. J Am Coll Cardiol 2017;69:1952–67. doi: 10.1016/j.jacc.2017.01.064
- 6. Montecucco F, Liberale L, Bonaventura A, Vecchiè A, Dallegri F, Carbone F. The role of inflammation in cardiovascular outcome. Curr Atheroscler Rep 2017;19:11. doi: 10.1007/s11883-017-0646-1
- 7. Bonaventura A, Liberale L, Vecchié A, Casula M, Carbone F, Dallegri F, et al. Update on inflammatory biomarkers and treatments in ischemic stroke. Int J Mol Sci 2016;17:1967. doi: 10.3390/ijms17121967
	- Liberale L, Montecucco F, Tardif J-C, Libby P, Camici GG. Inflamm-ageing: the role of inflammation in age-dependent cardiovascular disease. Eur Heart J 2020;1–14. doi: 10.1093/eurheartj/ehz961
- 9. Liberale L, Camici GG. The role of vascular aging in atherosclerotic plaque development and vulnerability. Curr Pharm Des 2019;25:1–14. doi: 10.2174/1381612825666190830175424
- 10. Cosentino F, Francia P, Camici GG, Pelicci PG, Lüscher TF. Final common molecular pathways of aging and cardiovascular disease: role of the p66shc protein. Arterioscler Thromb Vasc Biol 2008;28:622–8. doi: 10.1161/ATVBAHA.107.156059
- 11. Ahmed SBM, Prigent SA. Insights into the shc family of adaptor proteins. J Mol Signal 2017;12:1–17. doi: 10.5334/1750-2187-12-2
- 12. Giorgio M, Migliaccio E, Orsini F, Paolucci D, Moroni M, Contursi C, et al. Electron transfer between cytochrome c and p66shc generates reactive oxygen species that trigger mitochondrial apoptosis. Cell 2005;122:221–33. doi: 10.1016/j.cell.2005.05.011

Accepted Article

- 13. Pinton P, Rimessi A, Marchi S, Orsini F, Migliaccio E, Giorgio M, et al. Protein kinase c and prolyl isomerase 1 regulate mitochondrial effects of the life-span determinant p66shc. Science (80-) 2007;315:659–63. doi: 10.1126/science.1135380
- 14. Tschudi MR, Barton M, Bersinger NA, Moreau P, Cosentino F, Noll G, et al. Effect of age on kinetics of nitric oxide release in rat aorta and pulmonary artery. J Clin Invest 1996;98:899–905. doi: 10.1172/JCI118872
	- 15. Van Der Loo B, Labugger R, Skepper JN, Bachschmid M, Kilo J, Powell JM, et al. Enhanced peroxynitrite formation is associated with vascular aging. J Exp Med 2000;192:1731–43. doi: 10.1084/jem.192.12.1731
- 16. Pandolfi S, Bonafè M, Di Tella L, Tiberi L, Salvioli S, Monti D, et al. P66Shc is highly expressed in fibroblasts from centenarians. Mech Ageing Dev 2005;126:839–44. doi: 10.1016/j.mad.2005.03.004
- 17. Migliaccio E, Giogio M, Mele S, Pelicci G, Reboldi P, Pandolfi PP, et al. The p66(shc) adaptor protein controls oxidative stress response and life span in mammals. Nature 1999;402:309–13. doi: 10.1038/46311
- 18. Francia P, Delli Gatti C, Bachschmid M, Martin-Padura I, Savoia C, Migliaccio E, et al. Deletion of p66shc gene protects against age-related endothelial dysfunction. Circulation 2004;110:2889–95. doi: 10.1161/01.CIR.0000147731.24444.4D
- 19. Davignon J, Ganz P. Role of endothelial dysfunction in atherosclerosis. Circulation 2004;109. doi: 10.1161/01.cir.0000131515.03336.f8
- 20. Costantino S, Paneni F, Mitchell K, Mohammed SA, Hussain S, Gkolfos C, et al. Hyperglycaemia-induced epigenetic changes drive persistent cardiac dysfunction via the adaptor p66 shc. Int J Cardiol 2018;268:179–86. doi: 10.1016/j.ijcard.2018.04.082
- 21. Streese L, Khan AW, Deiseroth A, Hussain S, Suades R, Tiaden A, et al. Physical activity may drive healthy microvascular ageing via downregulation. 2019;doi: 10.1177/2047487319880367
- 22. Napoli C, Martin-Padura I, De Nigris F, Giorgio M, Mansueto G, Somma P, et al. Deletion of the p66shc longevity gene reduces systemic and tissue oxidative stress, vascular cell apoptosis, and early atherogenesis in mice fed a high-fat diet. Proc Natl Acad Sci U S A 2003;100:2112–6. doi: 10.1073/pnas.0336359100 **Accepted Article**
	- 23. Martin-Padura I, de Nigris F, Migliaccio E, Mansueto G, Minardi S, Rienzo M, et al. P66Shc deletion confers vascular protection in advanced atherosclerosi in

hypercholesterolemic apolipoprotein e knockout mice. Endothel J Endothel Cell Res 2008;15:276–87. doi: 10.1080/10623320802487791

- 24. Franzeck FC, Hof D, Spescha RD, Hasun M, Akhmedov A, Steffel J, et al. Expression of the aging gene p66shc is increased in peripheral blood monocytes of patients with acute coronary syndrome but not with stable coronary artery disease. Atherosclerosis 2012;220:282–6. doi: 10.1016/j.atherosclerosis.2011.10.035
- 25. Carpi A, Menabò R, Kaludercic N, Pelicci PG, Di Lisa F, Giorgio M. The cardioprotective effects elicited by p66shc ablation demonstrate the crucial role of mitochondrial ros formation in ischemia/reperfusion injury. Biochim Biophys Acta - Bioenerg 2009;1787:774–80. doi: 10.1016/j.bbabio.2009.04.001
- 26. Akhmedov A, Montecucco F, Braunersreuther V, Camici GG, Jakob P, Reiner MF, et al. Genetic deletion of the adaptor protein p66shc increases susceptibility to short-term ischaemic myocardial injury via intracellular salvage pathways. Eur Heart J 2015;36:516– 26. doi: 10.1093/eurheartj/ehu400
- 27. Spescha RD, Shi Y, Wegener S, Keller S, Weber B, Wyss MM, et al. Deletion of the ageing gene p66shc reduces early stroke size following ischaemia/reperfusion brain injury. Eur Heart J 2013;34:96–103. doi: 10.1093/eurheartj/ehs331
- 28. Spescha RD, Klohs J, Semerano A, Giacalone G, Derungs RS, Reiner MF, et al. Postischaemic silencing of p66shc reduces ischaemia/reperfusion brain injury and its expression correlates to clinical outcome in stroke. Eur Heart J 2015;36:1590–600. doi: 10.1093/eurheartj/ehv140 **Accepted Article**
	- 29. Hishikawa K, Lüscher TF. Pulsatile stretch stimulates superoxide production in human aortic endothelial cells. Circulation 1997 [cited 2019 Nov 18];96:3610–6. doi: 10.1161/01.CIR.96.10.3610
	- 30. Spescha RD, Glanzmann M, Simic B, Witassek F, Keller S, Akhmedov A, et al. Adaptor protein p66shc mediates hypertension-associated, cyclic stretch-dependent, endothelial damage. Hypertension 2014;64:347–53. doi: 10.1161/HYPERTENSIONAHA.113.02129
	- 31. Wang Y, Qu H, Liu J. P66Shc deletion ameliorates oxidative stress and cardiac dysfunction in pressure overload-induced heart failure. J Card Fail 2019;00:1–11. doi: 10.1016/j.cardfail.2019.09.003
	- 32. Graiani G, Lagrasta C, Migliaccio E, Spillmann F, Meloni M, Madeddu P, et al. Genetic deletion of the p66shc adaptor protein protects from angiotensin ii-induced myocardial

damage. Hypertension 2005;46:433–40. doi: 10.1161/01.HYP.0000174986.73346.ba

- 33. Liu GY, Sabatini DM. MTOR at the nexus of nutrition, growth, ageing and disease. Nat Rev Mol Cell Biol 2020;260:148–55. doi: 10.1038/s41580-019-0199-y
- 34. Johnson SC, Rabinovitch PS, Kaeberlein M. MTOR is a key modulator of ageing and agerelated disease. Nature 2013;493:338–45. doi: 10.1038/nature11861
- 35. Zhu Y, Pires KMP, Whitehead KJ, Olsen CD, Wayment B, Zhang YC, et al. Mechanistic target of rapamycin (mtor) is essential for murine embryonic heart development and growth. PLoS One 2013;8. doi: 10.1371/journal.pone.0054221
- 36. Zhang D, Contu R, Latronico MVG, Zhang JL, Rizzi R, Catalucci D, et al. MTORC1 regulates cardiac function and myocyte survival through 4e-bp1 inhibition in mice. J Clin Invest 2010;120:2805–16. doi: 10.1172/JCI43008
- 37. Sciarretta S, Zhai P, Maejima Y, DelRe DP, Nagarajan N, Yee D, et al. MTORC2 regulates cardiac response to stress by inhibiting mst1. Cell Rep 2015;11:125–36. doi: 10.1016/j.celrep.2015.03.010
- 38. Harrison DE, Strong R, Sharp ZD, Nelson JF, Astle CM, Flurkey K, et al. Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. Nature 2009;460:392–5. doi: 10.1038/nature08221
- 39. Flynn JM, O'Leary MN, Zambataro CA, Academia EC, Presley MP, Garrett BJ, et al. Latelife rapamycin treatment reverses age-related heart dysfunction. Aging Cell 2013;12:851– 62. doi: 10.1111/acel.12109
- 40. Kennedy BK, Lamming DW. The mechanistic target of rapamycin: the grand conductor of metabolism and aging. Cell Metab 2016;23:990–1003. doi: 10.1016/j.cmet.2016.05.009
- 41. Johnson RWG, Kreis H, Oberbauer R, Brattström C, Claesson K, Eris J. Sirolimus allows early cyclosporine withdrawal in renal transplantation resulting in improved renal function and lower blood pressure. Transplantation 2001;72:777–86. doi: 10.1097/00007890- 200109150-00007
- 42. Reho JJ, Guo DF, Rahmouni K. Mechanistic target of rapamycin complex 1 signaling modulates vascular endothelial function through reactive oxygen species. J Am Heart Assoc 2019;8. doi: 10.1161/JAHA.118.010662
- 43. Lesniewski LA, Seals DR, Walker AE, Henson GD, Blimline MW, Trott DW, et al. Dietary rapamycin supplementation reverses age-related vascular dysfunction and oxidative stress, while modulating nutrient-sensing, cell cycle, and senescence pathways. Aging Cell

Accepted Articles

2017;16:17–26. doi: 10.1111/acel.12524

- 44. Mitchell GF, Hwang SJ, Vasan RS, Larson MG, Pencina MJ, Hamburg NM, et al. Arterial stiffness and cardiovascular events: the framingham heart study. Circulation 2010;121:505– 11. doi: 10.1161/CIRCULATIONAHA.109.886655
- 45. Joannidès R, Monteil C, De Ligny BH, Westeel PF, Iacob M, Thervet E, et al. Immunosuppressant regimen based on sirolimus decreases aortic stiffness in renal transplant recipients in comparison to cyclosporine. Am J Transplant 2011;11:2414–22. doi: 10.1111/j.1600-6143.2011.03697.x
- 46. Franceschi C, Garagnani P, Parini P, Giuliani C, Santoro A. Inflammaging: a new immune– metabolic viewpoint for age-related diseases. Nat Rev Endocrinol 2018;14:576–90. doi: 10.1038/s41574-018-0059-4
- 47. Ross R. Atherosclerosis--an inflammatory disease. N Engl J Med 1999;340:115–26. doi: 10.1056/NEJM199901143400207
- 48. Zhang KS, Schecker J, Krull A, Riechert E, Jürgensen L, Kamuf-Schenk V, et al. PRAS40 suppresses atherogenesis through inhibition of mtorc1-dependent pro-inflammatory signaling in endothelial cells. Sci Rep 2019;9:16787. doi: 10.1038/s41598-019-53098-1 49. Correia-Melo C, Birch J, Fielder E, Rahmatika D, Taylor J, Chapman J, et al. Rapamycin improves healthspan but not inflammaging in nfκb1 −/− mice. Aging Cell 2019;18:e12882. doi: 10.1111/acel.12882 **Accepted Articles**
 Accepted Articles
 Accepted Articles
 Accepted Articles
 Accepted Articles
 Accepted Articles
	- 50. Kurdi A, De Meyer GRY, Martinet W. Potential therapeutic effects of mtor inhibition in atherosclerosis. Br J Clin Pharmacol 2016;1267–79. doi: 10.1111/bcp.12820
	- 51. Kane AE, Sinclair DA. Sirtuins and nad+ in the development and treatment of metabolic and cardiovascular diseases. Circ Res 2018;123:868–85. doi: 10.1161/CIRCRESAHA.118.312498
	- 52. Giuliani C, Garagnani P, Franceschi C. Genetics of human longevity within an ecoevolutionary nature-nurture framework. Circ Res 2018;123:745–72. doi: 10.1161/CIRCRESAHA.118.312562
	- 53. Houtkooper RH, Pirinen E, Auwerx J. Sirtuins as regulators of metabolism and healthspan. Nat Rev Mol Cell Biol 2012;13:225–38. doi: 10.1038/nrm3293
	- 54. Ungvari Z, Tarantini S, Donato AJ, Galvan V, Csiszar A. Mechanisms of vascular aging. Circ Res 2018;123:849–67. doi: 10.1161/CIRCRESAHA.118.311378
	- 55. Rajman L, Chwalek K, Sinclair DA. Therapeutic potential of nad-boosting molecules: the

in vivo evidence. Cell Metab 2018;27:529–47. doi: 10.1038/nature01578

- 56. Zhang QJ, Wang Z, Chen HZ, Zhou S, Zheng W, Liu G, et al. Endothelium-specific overexpression of class iii deacetylase sirt1 decreases atherosclerosis in apolipoprotein edeficient mice. Cardiovasc Res 2008;80:191–9. doi: 10.1093/cvr/cvn224
- 57. Miranda MX, Van Tits LJ, Lohmann C, Arsiwala T, Winnik S, Tailleux A, et al. The sirt1 activator srt3025 provides atheroprotection in apoe-/- mice by reducing hepatic pcsk9 secretion and enhancing ldlr expression. Eur Heart J 2015;36:51–9. doi: 10.1093/eurheartj/ehu095
- 58. Stein S, Schäfer N, Breitenstein A, Besler C, Winnik S, Lohmann C, et al. SIRT1 reduces endothelial activation without affecting vascular function in apoe-/-mice. Aging (Albany NY) 2010;2:344–52.
- 59. Hsu CP, Zhai P, Yamamoto T, Maejima Y, Matsushima S, Hariharan N, et al. Silent information regulator 1 protects the heart from ischemia/reperfusion. Circulation 2010;122:2170–82. doi: 10.1161/CIRCULATIONAHA.110.958033
- 60. Sundaresan NR, Samant SA, Pillai VB, Rajamohan SB, Gupta MP. SIRT3 is a stressresponsive deacetylase in cardiomyocytes that protects cells from stress-mediated cell death by deacetylation of ku70. Mol Cell Biol 2008;28:6384–401. doi: 10.1128/mcb.00426-08
- 61. Liu B, Che W, Xue J, Zheng C, Tang K, Zhang J, et al. SIRT4 prevents hypoxia-induced apoptosis in h9c2 cardiomyoblast cells. Cell Physiol Biochem 2013;32:655–62. doi: 10.1159/000354469
- 62. Liu B, Che W, Zheng C, Liu W, Wen J, Fu H, et al. SIRT5: a safeguard against oxidative stress-induced apoptosis in cardiomyocytes. Cell Physiol Biochem 2013;32:1050–9. doi: 10.1159/000354505 **Accepted Articles**

Contract Con
	- 63. Klishadi MS, Zarei F, Hejazian SH, Moradi A, Hemati M, Safari F. Losartan protects the heart against ischemia reperfusion injury: sirtuin3 involvement. J Pharm Pharm Sci 2015;18:112. doi: 10.18433/J3XG7T
	- 64. He X, Zeng H, Chen J-X. Ablation of sirt3 causes coronary microvascular dysfunction and impairs cardiac recovery post myocardial ischemia. Int J Cardiol 2016;215:349–57. doi: 10.1016/j.ijcard.2016.04.092
	- 65. Liberale L, Gaul DS, Akhmedov A, Bonetti NR, Nageswaran V, Costantino S, et al. Endothelial sirt6 blunts stroke size and neurological deficit by preserving blood–brain barrier integrity: a translational study. Eur Heart J 2019;1–13. doi:

10.1093/eurheartj/ehz712

- 66. Diaz-Cañestro C, Merlini M, Bonetti NR, Liberale L, Wüst P, Briand-Schumacher S, et al. Sirtuin 5 as a novel target to blunt blood–brain barrier damage induced by cerebral ischemia/reperfusion injury. Int J Cardiol 2018;260:148–55. doi: 10.1016/j.ijcard.2017.12.060 **Accepted Articles**
	- 67. Martin AS, Abraham DM, Hershberger KA, Bhatt DP, Mao L, Cui H, et al. Nicotinamide mononucleotide requires sirt3 to improve cardiac function and bioenergetics in a friedreich's ataxia cardiomyopathy model. JCI insight 2017;2. doi: 10.1172/jci.insight.93885
	- 68. Pillai VB, Bindu S, Sharp W, Fang YH, Kim G, Gupta M, et al. Sirt3 protects mitochondrial dna damage and blocks the development of doxorubicin-induced cardiomyopathy in mice. Am J Physiol - Hear Circ Physiol 2016;310:H962–72. doi: 10.1152/ajpheart.00832.2015
	- 69. Sundaresan NR, Gupta M, Kim G, Rajamohan SB, Isbatan A, Gupta MP. Sirt3 blocks the cardiac hypertrophic response by augmenting foxo3a-dependent antioxidant defense mechanisms in mice. J Clin Invest 2009;119. doi: 10.1172/JCI39162
	- 70. Gao P, Xu TT, Lu J, Li L, Xu J, Hao DL, et al. Overexpression of sirt1 in vascular smooth muscle cells attenuates angiotensin ii-induced vascular remodeling and hypertension in mice. J Mol Med 2014;92:347–57. doi: 10.1007/s00109-013-1111-4
	- 71. Paulin R, Dromparis P, Sutendra G, Gurtu V, Zervopoulos S, Bowers L, et al. Sirtuin 3 deficiency is associated with inhibited mitochondrial function and pulmonary arterial hypertension in rodents and humans. Cell Metab 2014;20:827–39. doi:
		- 10.1016/j.cmet.2014.08.011
	- 72. Hershberger KA, Abraham DM, Martin AS, Mao L, Liu J, Gu H, et al. Sirtuin 5 is required for mouse survival in response to cardiac pressure overload. J Biol Chem 2017;292:19767– 81. doi: 10.1074/jbc.M117.809897
	- 73. Zhang M, Wu J, Sun R, Tao X, Wang X, Kang Q, et al. SIRT5 deficiency suppresses mitochondrial atp production and promotes ampk activation in response to energy stress. PLoS One 2019;14:e0211796. doi: 10.1371/journal.pone.0211796
	- 74. Taneike M, Yamaguchi O, Nakai A, Hikoso S, Takeda T, Mizote I, et al. Inhibition of autophagy in the heart induces age-related cardiomyopathy. Autophagy 2010;6:600–6. doi: 10.4161/auto.6.5.11947

- **Accepted Articles** Articles Arti
- 75. Abdellatif M, Sedej S, Carmona-Gutierrez D, Madeo F, Kroemer G. Autophagy in cardiovascular aging. Circ Res 2018;123:803–24. doi: 10.1161/CIRCRESAHA.118.312208 76. Eisenberg T, Abdellatif M, Schroeder S, Primessnig U, Stekovic S, Pendl T, et al.
	- Cardioprotection and lifespan extension by the natural polyamine spermidine. Nat Med 2016;22:1428–38. doi: 10.1038/nm.4222
	- 77. Meléndez A, Tallóczy Z, Seaman M, Eskelinen EL, Hall DH, Levine B. Autophagy genes are essential for dauer development and life-span extension in c. elegans. Science (80-) 2003;301:1387–91. doi: 10.1126/science.1087782
	- 78. Fernández ÁF, Sebti S, Wei Y, Zou Z, Shi M, McMillan KL, et al. Disruption of the beclin 1-bcl2 autophagy regulatory complex promotes longevity in mice. Nature 2018;558:136– 40. doi: 10.1038/s41586-018-0162-7
	- 79. Zhu H, Tannous P, Johnstone JL, Kong Y, Shelton JM, Richardson J a, et al. Cardiac autophagy is a maladaptive response to hemodynamic stress. J Clin Invest 2007;117:1782– 93. doi: 10.1172/JCI27523
	- 80. Razani B, Feng C, Coleman T, Emanuel R, Wen H, Hwang S, et al. Autophagy links inflammasomes to atherosclerotic progression. Cell Metab 2012;15:534–44. doi: 10.1016/j.cmet.2012.02.011
	- 81. Kuro-o M. The klotho proteins in health and disease. Nat Rev Nephrol 2019;15:27–44. doi: 10.1038/s41581-018-0078-3
	- 82. Kuro-o M, Matsumura Y, Aizawa H, Kawaguchi H, Suga T, Utsugi T, et al. Mutation of the mouse klotho gene leads to a syndrome resembling ageing. Nature 1997;390:45–51. doi: 10.1038/36285
	- 83. Kurosu H, Yamamoto M, Clark JD, Pastor J V., Nandi A, Gurnani P, et al. Physiology: suppression of aging in mice by the hormone klotho. Science (80-) 2005;309:1829–33. doi: 10.1126/science.1112766
		- 84. Xie J, Cha SK, An SW, Kuro-O M, Birnbaumer L, Huang CL. Cardioprotection by klotho through downregulation of trpc6 channels in the mouse heart. Nat Commun 2012;3. doi: 10.1038/ncomms2240
	- 85. Hsieh CC, Kuro-o M, Rosenblatt KP, Brobey R, Papaconstantinou J. The ask1-signalosome regulates p38 mapk activity in response to levels of endogenous oxidative stress in the klotho mouse models of aging. Aging (Albany NY) 2010;2:597–611. doi: 10.18632/aging.100194
- 86. Gao D, Zuo Z, Tian J, Ali Q, Lin Y, Lei H, et al. Activation of sirt1 attenuates klotho deficiency-induced arterial stiffness and hypertension by enhancing amp-activated protein kinase activity. Hypertension 2016;68:1191–9. doi: 10.1161/HYPERTENSIONAHA.116.07709
- 87. Arking DE, Becker DM, Yanek LR, Fallin D, Judge DP, Moy TF, et al. KLOTHO allele status and the risk of early-onset occult coronary artery disease. Am J Hum Genet 2003;72:1154–61. doi: 10.1086/375035
- 88. Martins R, Lithgow GJ, Link W. Long live foxo: unraveling the role of foxo proteins in aging and longevity. Aging Cell 2016;15:196–207. doi: 10.1111/acel.12427
- 89. Willcox BJ, Donlon TA, He Q, Chen R, Grove JS, Yano K, et al. FOXO3A genotype is strongly associated with human longevity. Proc Natl Acad Sci U S A 2008;105:13987–92. doi: 10.1073/pnas.0801030105
- 90. Helgadottir A, Thorleifsson G, Gretarsdottir S, Stefansson OA, Tragante V, Thorolfsdottir RB, et al. Genome-wide analysis yields new loci associating with aortic valve stenosis. Nat Commun 2018;9:987. doi: 10.1038/s41467-018-03252-6
- 91. Blice-Baum AC, Zambon AC, Kaushik G, Viswanathan MC, Engler AJ, Bodmer R, et al. Modest overexpression of foxo maintains cardiac proteostasis and ameliorates ageassociated functional decline. Aging Cell 2017;16:93–103. doi: 10.1111/acel.12543 **Accepted Articles**
	- 92. Ni YG, Berenji K, Wang N, Oh M, Sachan N, Dey A, et al. Foxo transcription factors blunt cardiac hypertrophy by inhibiting calcineurin signaling. Circulation 2006;114:1159–68. doi: 10.1161/CIRCULATIONAHA.106.637124
	- 93. Potente M, Urbich C, Sasaki KI, Hofmann WK, Heeschen C, Aicher A, et al. Involvement of foxo transcription factors in angiogenesis and postnatal neovascularization. J Clin Invest 2005;115:2382–92. doi: 10.1172/JCI23126
	- 94. Kops GJPL, Dansen TB, Polderman PE, Saarloos I, Wirtz KWA, Coffer PJ, et al. Forkhead transcription factor foxo3a protects quiescent cells from oxidative stress. Nature 2002;419:316–21. doi: 10.1038/nature01036
	- 95. Nemoto S. Redox regulation of forkhead proteins through a p66shc-dependent signaling pathway. Science (80-) 2002;295:2450–2. doi: 10.1126/science.1069004
	- 96. Tsuchiya K, Westerterp M, Murphy AJ, Subramanian V, Ferrante AW, Tall AR, et al. Expanded granulocyte/monocyte compartment in myeloid-specific triple foxo knockout increases oxidative stress and accelerates atherosclerosis in mice. Circ Res 2013;112:992–

1003. doi: 10.1161/CIRCRESAHA.112.300749

- 97. de Cabo R, Mattson MP. Effects of intermittent fasting on health, aging, and disease. N Engl J Med 2019;381:2541–51. doi: 10.1056/NEJMra1905136
- 98. Anderson RM, Bitterman KJ, Wood JG, Medvedik O, Sinclair DA. Nicotinamide and pnc1 govern lifespan extension by calorie restriction in saccharomyces cerevisiae. Nature 2003;423:181–5. doi: 10.1038/nature01578 97.

98.

99.

99.

100.

100.
	- 99. Imai S, Guarente L. NAD+ and sirtuins in aging and disease. Trends Cell Biol 2014;24:464–71. doi: 10.1002/cphy.c160029.
	- 100. Carbone F, Liberale L, Bonaventura A, Vecchiè A, Casula M, Cea M, et al. Regulation and function of extracellular nicotinamide phosphoribosyltransferase/visfatin. Compr Physiol 2017;603–21. doi: 10.2174/138161209787185788
	- 101. Chini E. CD38 as a regulator of cellular nad: a novel potential pharmacological target for metabolic conditions. Curr Pharm Des 2009;15:57–63. doi: 10.2147/CMAR.S200524
	- 102. Jiang X, Li W, Li X, Bai H, Zhang Z. Current status and future prospects of parp inhibitor clinical trials in ovarian cancer. Cancer Manag Res 2019;Volume 11:4371–90. doi: 10.2147/CMAR.S200524
	- 103. Liberale L, Bonaventura A, Montecucco F, Dallegri F, Carbone F. Impact of red wine consumption on cardiovascular health. Curr Med Chem 2019;26:3542–66. doi: 10.2174/0929867324666170518100606
	- 104. Dai H, Sinclair DA, Ellis JL, Steegborn C. Sirtuin activators and inhibitors: promises, achievements, and challenges. Pharmacol Ther 2018;188:140–54. doi: 10.1016/j.pharmthera.2018.03.004
	- 105. Howitz KT, Bitterman KJ, Cohen HY, Lamming DW, Lavu S, Wood JG, et al. Small molecule activators of sirtuins extend saccharomyces cerevisiae lifespan. Nature 2003;425:191–6. doi: 10.1038/nature01960
	- 106. Sajish M, Schimmel P. A human trna synthetase is a potent parp1-activating effector target for resveratrol. Nature 2015;519:370–3. doi: 10.1038/nature14028
	- 107. Cantó C, Gerhart-Hines Z, Feige JN, Lagouge M, Noriega L, Milne JC, et al. AMPK regulates energy expenditure by modulating nad+ metabolism and sirt1 activity. Nature 2009;458:1056–60. doi: 10.1038/nature07813
	- 108. Gano LB, Donato AJ, Pasha HM, Hearon CM, Sindler AL, Seals DR. The sirt1 activator srt1720 reverses vascular endothelial dysfunction, excessive superoxide production, and

inflammation with aging in mice. Am J Physiol Circ Physiol 2014;307:H1754–63. doi: 10.1152/ajpheart.00377.2014

- 109. Das A, Huang GX, Bonkowski MS, Longchamp A, Li C, Schultz MB, et al. Impairment of an endothelial nad+-h2s signaling network is a reversible cause of vascular aging. Cell 2018;173:74-89.e20. doi: 10.1371/journal.pone.0098972
- 110. Yamamoto T, Byun J, Zhai P, Ikeda Y, Oka S, Sadoshima J. Nicotinamide mononucleotide, an intermediate of nad+ synthesis, protects the heart from ischemia and reperfusion. PLoS One 2014;9:e98972. doi: 10.1371/journal.pone.0098972
- 111. Hsu C-P, Oka S, Shao D, Hariharan N, Sadoshima J. Nicotinamide phosphoribosyltransferase regulates cell survival through nad + synthesis in cardiac myocytes. Circ Res 2009;105:481–91. doi: 10.1161/CIRCRESAHA.109.203703
- 112. Lee CF, Chavez JD, Garcia-Menendez L, Choi Y, Roe ND, Chiao YA, et al. Normalization of nad + redox balance as a therapy for heart failure. Circulation 2016;134:883–94. doi: 10.1161/CIRCULATIONAHA.116.022495
- 113. Pillai VB, Sundaresan NR, Kim G, Gupta M, Rajamohan SB, Pillai JB, et al. Exogenous nad blocks cardiac hypertrophic response via activation of the sirt3-lkb1-amp-activated kinase pathway. J Biol Chem 2010;285:3133–44. doi: 10.1074/jbc.M109.077271
- 114. Kane AE, Sinclair DA. Sirtuins and nad + in the development and treatment of metabolic and cardiovascular diseases. Circ Res 2018;123:868–85. doi: 10.1161/CIRCRESAHA.118.312498
- 115. Timmers S, Konings E, Bilet L, Houtkooper RH, van de Weijer T, Goossens GH, et al. Calorie restriction-like effects of 30 days of resveratrol supplementation on energy metabolism and metabolic profile in obese humans. Cell Metab 2011;14:612–22. doi: 10.1016/j.cmet.2011.10.002 **Accepted Articles** Articles Arti
	- 116. Méndez-del Villar M, González-Ortiz M, Martínez-Abundis E, Pérez-Rubio KG, Lizárraga-Valdez R. Effect of resveratrol administration on metabolic syndrome, insulin sensitivity, and insulin secretion. Metab Syndr Relat Disord 2014;12:497–501. doi: 10.1089/met.2014.0082
	- 117. Poulsen MM, Vestergaard PF, Clasen BF, Radko Y, Christensen LP, Stodkilde-Jorgensen H, et al. High-dose resveratrol supplementation in obese men: an investigator-initiated, randomized, placebo-controlled clinical trial of substrate metabolism, insulin sensitivity, and body composition. Diabetes 2013;62:1186–95. doi: 10.2337/db12-0975
- 118. Chen S, Zhao X, Ran L, Wan J, Wang X, Qin Y, et al. Resveratrol improves insulin resistance, glucose and lipid metabolism in patients with non-alcoholic fatty liver disease: a randomized controlled trial. Dig Liver Dis 2015;47:226–32. doi: 10.1016/j.dld.2014.11.015
- 119. Magyar K, Halmosi R, Palfi A, Feher G, Czopf L, Fulop A, et al. Cardioprotection by resveratrol: a human clinical trial in patients with stable coronary artery disease. Clin Hemorheol Microcirc 2012;50:179–87. doi: 10.3233/CH-2011-1424
- 120. Dellinger RW, Santos SR, Morris M, Evans M, Alminana D, Guarente L, et al. Repeat dose nrpt (nicotinamide riboside and pterostilbene) increases nad+ levels in humans safely and sustainably: a randomized, double-blind, placebo-controlled study. npj Aging Mech Dis 2017;3:17. doi: 10.1038/s41514-017-0016-9 **Accepted Articles** 123.
 Accepted Articles 123.
 Accepted Articles 124.

124.

125.

128.
	- 121. Trammell SAJ, Schmidt MS, Weidemann BJ, Redpath P, Jaksch F, Dellinger RW, et al. Nicotinamide riboside is uniquely and orally bioavailable in mice and humans. Nat Commun 2016;7:12948. doi: 10.1038/ncomms12948
	- 122. Ren J, Esnouf R, Hopkins A, Ross C, Jones Y, Stammers D, et al. The structure of hiv-1 reverse transcriptase complexed with 9-chloro-tibo: lessons for inhibitor design. Structure 1995;3:915–26. doi: 10.1016/S0969-2126(01)00226-X
	- 123. Costantino S, Camici GG, Mohammed SA, Volpe M, Lüscher TF, Paneni F. Epigenetics and cardiovascular regenerative medicine in the elderly. Int J Cardiol 2018;250:207–14. doi: 10.1016/j.ijcard.2017.09.188
	- 124. Stojanović SD, Fiedler J, Bauersachs J, Thum T, Sedding DG. Senescence-induced inflammation: an important player and key therapeutic target in atherosclerosis. Eur Heart J 2020;1–14. doi: 10.1093/eurheartj/ehz919
	- 125. Mytych J, Sołek P, Będzińska A, Rusinek K, Warzybok A, Tabęcka-Łonczyńska A, et al. Towards age-related anti-inflammatory therapy: klotho suppresses activation of er and golgi stress response in senescent monocytes. Cells 2020;9:261. doi: 10.3390/cells9020261
	- 126. Salminen A, Kauppinen A, Kaarniranta K. Emerging role of nf-κb signaling in the induction of senescence-associated secretory phenotype (sasp). Cell Signal 2012;24:835– 45. doi: 10.1016/j.cellsig.2011.12.006
	- 127. Baar MP, Brandt RMC, Putavet DA, Klein JDD, Derks KWJ, Bourgeois BRM, et al. Targeted apoptosis of senescent cells restores tissue homeostasis in response to chemotoxicity and aging. Cell 2017;169:132-147.e16. doi: 10.1016/j.cell.2017.02.031
	- 128. Chang J, Wang Y, Shao L, Laberge R-M, Demaria M, Campisi J, et al. Clearance of

senescent cells by abt263 rejuvenates aged hematopoietic stem cells in mice. Nat Med 2016;22:78–83. doi: 10.1038/nm.4010

- 129. Hickson LJ, Langhi Prata LGP, Bobart SA, Evans TK, Giorgadze N, Hashmi SK, et al. Senolytics decrease senescent cells in humans: preliminary report from a clinical trial of dasatinib plus quercetin in individuals with diabetic kidney disease. EBioMedicine 2019;47:446–56. doi: 10.1016/j.ebiom.2019.08.069
- 130. Hickson LJ, Langhi Prata LGP, Bobart SA, Evans TK, Giorgadze N, Hashmi SK, et al. Corrigendum to "Senolytics decrease senescent cells in humans: preliminary report from a clinical trial of dasatinib plus quercetin in individuals with diabetic kidney disease" ebiomedicine 47 (2019) 446-456. EBioMedicine 2020;52:102595. doi: 10.1016/j.ebiom.2019.12.004

Figure legends

Figure 1. The hallmarks of the ageing cardiovasculature. During the course of ageing, the imbalance of genes involved in ageing (red) and longevity (green) fuels deleterious cellular processes that coincide with the functional and structural decline of the arterial system (left) and the heart (right). Note that the aged myocardium displays ventricular hypertrophy and fibrosis, whereas the ageing vasculature exhibits endothelial dysfunction, arterial stiffening, and atherosclerosis, culminating in increased risk of myocardial infarction, stroke and heart failure. * indicates direct or indirect involvement of the ageing genes mentioned in disease initiation or progression. For details see main text.

Figure 2. Dysregulation of the outlined signalling network which impacts longevity and ageing converges on the activation of pro-inflammatory pathways and perturbated redox signalling. Dietary interventions (e.g intermittent fasting or caloric restriction) act on downstream targets such as sirtuins (which deacetylate FOXO) and mTOR, resulting in enhanced expression of genes implicated in stress resistance and mitochondrial biogenesis. Likewise, the administration of sirtuin-activating compounds, such as STACs, may represent feasible strategies to combat the burden of age-related cardiovascular maladies. Substrate boosting through NAD+ precursor supplementation, enzyme induction or the inhibition of degrading pathways may be a novel approach to activate downstream targets such as sirtuins which impact many cellular functions, including inflammation and stress resistance. Finally, epigenetic reprogramming of circulating or bone marrow-derived cells followed by autologous transplantation may be beneficial to rescue maladaptive changes of the human genome occurring with ageing. **Access 12**
 Access 13
 Access 13
 Access 14
 Access 14

The Aging Cardiovasculature

Aging and Longevity Genes

