

Allergen Immunotherapy

History and Future Developments

Giovanni Passalacqua, MD*, Giorgio Walter Canonica, MD

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KEYWORDS

- Subcutaneous immunotherapy • Sublingual immunotherapy • Indications
- Allergen immunotherapy • Efficacy • Safety • History • Molecular diagnosis

KEY POINTS

- Allergen immunotherapy (AIT) is a cornerstone in the management of respiratory allergic diseases because it is allergen-specific and immunomodulating and may affect disease progression.
- Sublingual immunotherapy (SLIT) represents a significant advance, offering patients an excellent safety and acceptance profile.
- From a historical viewpoint, in the past three decades there has been an impressive development in this form of treatment, which has lasted more than 100 years.
- The most promising fields are the use of AIT in food allergy, preventative effects, and improvement of routes of administration and standardization of extracts and protocols.

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THE HISTORICAL PERSPECTIVE

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AIT was introduced into clinical practice more than a century ago by Leonard Noon,¹ with the aim of “vaccinating” against hypothetical “aerogenic toxins” (Fig. 1). Despite the wrong rationale, the subcutaneous immunotherapy (SCIT) of pollen extracts was effective in reducing hay fever symptoms. Subsequently, the use of SCIT gradually increased and was progressively extended to other allergens. SCIT remained the only mode of administration for more than 70 years, and its use remained totally empirical until 1965 when IgE was discovered.² The first randomized controlled study on AIT was published in 1954 by Frankland and Augustin,³ and a few years later, Johnstone and Dutton⁴ suggested that AIT could modify the natural history of respiratory allergy, but this fact was not considered for another 40 years. In 1978, the first randomized, double-blinded, placebo-controlled (RDBPC) trial with AIT for hymenoptera venom allergy appeared,⁵ showing the superiority of purified venoms over whole-body extracts. This was followed by numerous other trials substantially confirming the efficacy

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Allergy and Respiratory Diseases, IRCCS San Martino-IST, University of Genoa, Genoa, Italy
* Corresponding author. Allergy and Respiratory Diseases, DIMI, Padiglione Maragliano, Largo Rosanna Benzi 10, Genoa 16132, Italy.
E-mail address: passalacqua@unige.it

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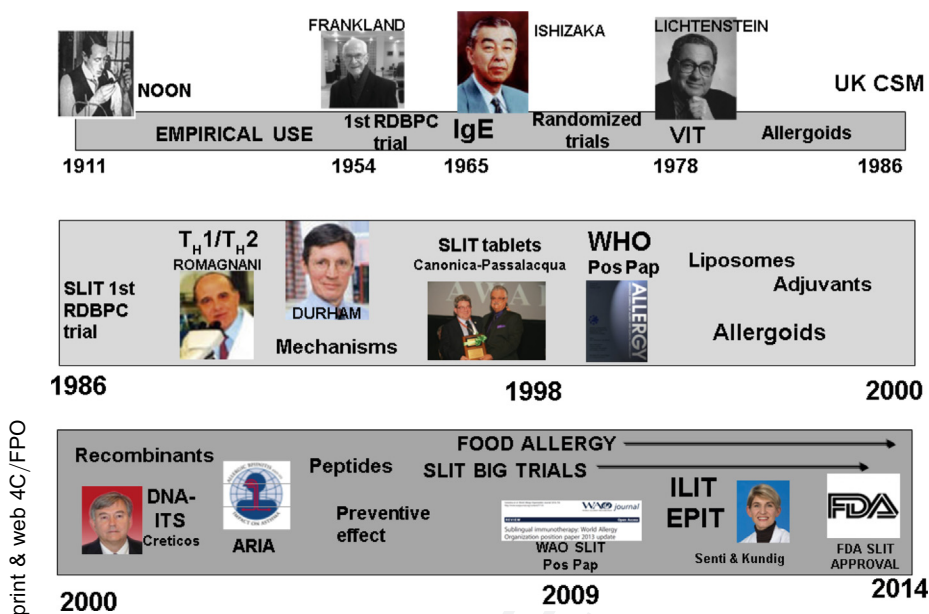


Fig. 1. The history of AIT.

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and safety of venom immunotherapy (VIT),⁶ now widely used and well standardized in procedures.

It became clear that SCIT with respiratory allergens involved a certain risk of severe or even fatal adverse events,⁷ as established by the UK Committee on Safety of Medicines in 1986.⁸ Many AIT adverse events are due to human errors, but some adverse events are unpredictable and unavoidable.^{9,10} This fact prompted the search for safer routes of administration of AIT. Among the proposed routes, SLIT rapidly established scientific credibility and soon remained the most viable alternative to SCIT. Other routes of administration had been proposed: the local bronchial during the 1950s, the local nasal during the 1970s, and the oral at the beginning of the 1980s (for review see Canonica and Passalacqua¹¹). The results of clinical trials demonstrated that the efficacy of oral and bronchial routes is unproved and the risk/benefit ratio is unfavorable; thus, these routes of administration were abandoned, although there is currently a renewed interest for the oral route in the desensitization for food allergy. The local nasal immunotherapy proved effective for allergic rhinitis but because of the impractical administration technique, its clinical use rapidly declined.

The first randomized, double-blind, placebo-controlled trial with SLIT appeared in 1986,¹² and it was followed by numerous other trials which, although conducted in small samples, substantially confirmed the efficacy of this route. SLIT was first mentioned as a possible alternative to SCIT in a World Health Organization position paper¹³ in 1998, and its role in clinical practice was confirmed in the subsequent official documents.^{14,15}

In the meanwhile, other relevant advances about AIT appeared. Among the most important were the discovery of the helper T cell (T_H1/T_H2) system,¹⁶ the re-evaluation of the role of IgG4 as blocking antibodies,¹⁷ and the description of the regulatory T cells.^{18,19} The improved knowledge of the mechanisms of action²⁰ allowed for the introduction of new approaches, such as the use of adjuvants (currently some

products are commercialized) and the use of antigenic peptides and the recombinant allergens. In parallel, other specific aspects began to be investigated, namely the preventive effect on the development of asthma, that was demonstrated for both SCIT and SLIT, although in open trials and with relatively small populations.^{21–23}

In the past decade, the efficacy of SLIT was clearly confirmed in the so-called big trials, which included hundreds (usually from 250 to more than 800) of patients. Some of those trials involved a dose-ranging design^{24–29} and therefore allowed identification of the optimal maintenance dose for each of the tested products, at least for the relevant allergens (grass, mite, and ragweed). There is 1 single dose-ranging large trial performed with SCIT.³⁰ The introduction of fast-dissolving tablets for SLIT further improved the convenience. The official acceptance of SLIT culminated in 2009 with the publication of a first position paper prepared by the World Allergy Organization,³¹ including 60 RDBPC trials, followed by an updated version with 77 trials.³² Approximately 1 year ago, the Food and Drug Administration (FDA), approved 3 SLIT tablet products to be marketed in the United States.³³

THE PRESENT SITUATION

Practical Aspects

To date, the practice of AIT is standardized, and numerous official position papers and practice parameters are available worldwide (Table 1). In particular, hymenoptera VIT, although there are different extracts available, is well standardized and its practice is uniform.

Table 1
The main position papers and guidelines on allergen immunotherapy

Year	Organization	Type of Allergen Immunotherapy	Reference
1998	World Health Organization	SCIT/SLIT	Ann Allergy Asthma Immunol 1998;81(5 Pt 1):401–5.
1998	European Academy of Allergy and Clinical Immunology	Non injection routes	Allergy 1998;53:933–44.
2001	Allergic Rhinitis and its Impact on Asthma	SCIT/SLIT	J Allergy Clin Immunol 2001;108(5 Suppl):S147–334.
2005	European Academy of Allergy and Clinical Immunology	VIT	Allergy 2005;60:1459–70.
2007	American Academy of Allergy, Asthma & Immunology/ American College of Allergy, Asthma & Immunology	SCIT	J Allergy Clin Immunol 2007;120(Suppl):S25–85, IV.
2008	Allergic Rhinitis and its Impact on Asthma	SCIT/SLIT	Allergy 2008;63(Suppl 86):8–160.
2009	World Allergy Organization	SLIT	Allergy 2009;64(Suppl 91):1–59.
2011	American Academy of Allergy, Asthma & Immunology/ American College of Allergy, Asthma & Immunology	SCIT	J Allergy Clin Immunol 2011;127(1 Suppl):S1–55.
2011	British Society for Allergy and Clinical Immunology	VIT	Clin Exp Allergy 2011;41:1201–20.
2013	World Allergy Organization	SLIT	World Allergy Organ J 2014;7(1):6.

151 At variance with SCIT, which is standardized in regimens and protocols, SLIT is
152 affected by numerous variables. It can be administered as drops, monodose vials,
153 or tablets and with variable timings and doses. In particular, the maintenance dose
154 is strictly dependent on the method of standardization, which varies from one manu-
155 facturer to another. It is also true that all the products that are officially approved
156 (eg, by the FDA or European Medicines Agency) display the content in micrograms Q15
157 of major allergen(s) per dose. At present, tablets that were first introduced in 1998
158 as monomeric allergoids³⁴ seem to represent the preferred SLIT formulation because
159 of ease of use. Also, the time interval between each maintenance dose varies from one
160 producer to another (daily, on alternate days, or twice weekly), but the current attitude
161 is to prefer once-a-day administration.³⁵ For pollen allergies, the pre-coseasonal
162 protocol is the most largely used, because its efficacy does not differ from that of Q16
163 the continuous (all-year-long) administration.^{36,37}

164 Another important and unresolved debate concerns the use of mixtures of allergens.
165 The European view is that AIT is given for no more than 3 allergens in the same pa-
166 tient,³⁸ and the dose of each allergen is given separately. In the United States, the
167 usual practice is multiple allergens mixed together in a single preparation with atten-
168 tion to not mixing allergens that can degradate other proteins.³⁹ This dichotomy has
169 cultural and historical reasons and is attributable to different concentrations of allergen
170 solutions, which are usually higher in the United States products.⁴⁰ There are few well Q17
171 designed studies that have evaluated and demonstrated the efficacy of allergen mix-
172 tures.⁴¹ On the contrary, it is now accepted that AIT with a single allergen is effective in Q18
173 polysensitized patients, provided the allergen chosen is responsible for the disease.⁴²
174 In this regard, the molecular-based diagnosis (molecular allergy) has become a useful
175 tool to refine the prescription of AIT (discussed later).

176 Other current fields of research in AIT are pharmacoeconomic aspects and adher-
177 ence. Looking at the published studies, it seems that in the long term both SCIT and
178 SLIT produce economic savings for both patients and health providers.⁴³ This is a
179 result of a combination of reduced drug consumption and health care utilization (direct
180 costs) as well as improvement of the quality of life (indirect costs). In contrast, adher-
181 ence is a major problem, particularly for SLIT, which is self-administered: although
182 structured studies provided overall favorable results in terms of adherence,⁴⁴ real-
183 life adherence is reported to be poor,⁴⁵ although more frequent follow-up of patients
184 seems to increase compliance.⁴⁶

185 ***The Role of Molecular Diagnosis***

187 The IgE response is not generically directed toward an allergenic source but rather
188 to specific proteins (or epitopes) that are contained into the raw material. For
189 instance, the IgE response to “grasses” is directed to a few proteins (Phl p 1, Phl Q19
190 p 5, an Phl p 6), and the IgE response to mite is specific for the proteins Der p 1,
191 Der p 2, Der f 1, Der f 2, and so forth.⁴⁷ Such molecules are considered the genuine
192 sensitizers. On the other hand, there are also highly conserved, are present in Q20
193 different species (eg, profilins, lipid transfer proteins, and storage proteins). They
194 are called pan-allergens or cross-reacting proteins⁴⁷ and are often responsible for
195 multiple positivities on the standard diagnostic tests. The relevant implications of
196 pan-allergen sensitization may be particularly pertinent in AIT. The molecular diag-
197 nosis allows distinction of genuine sensitizations from the positivities due to cross-
198 reacting proteins, thereby refining the choice of the allergen to be used for AIT.⁴⁸
199 Several studies have shown that molecular diagnosis significantly modifies the pre-
200 scription of AIT in polysensitized patients.^{49,50} Many recombinant or purified molec- Q21
201 ular components for skin testing and immunoassay are available in multiplexed

202 systems that allow detecting in a single analysis specific IgE toward approximately
203 130 allergenic molecules.⁵¹

204 **Regulatory Aspects**

206 Despite the amount of clinical and mechanistic data on AIT and its consolidated use,
207 the regulatory aspects (pharmacologic classification of products, marketing authori-
208 zation, national and supranational approval, and deputy regulatory authorities) remain
209 vague and largely differ among countries. Although in the United States and in the
210 European Community (EC), there are well-defined regulatory authorities (FDA, Euro-
211 pean Medicines Agency, and Paul Ehrlich Institute), in other countries, such as those
212 in Latin America, there is no uniform regulation.⁵²

213 In Europe, numerous official regulatory documents have been released (for review,
214 see Kaul and colleagues⁵³ and Bonini⁵⁴), mainly concerning Good Manufacturing
215 Practice. Those documents impose on all members of the EC specific standards for
216 the production of allergen extracts. Within the EC, apart from a few exceptions,
217 allergen extracts are considered named patient products (NPPs), prepared individu-
218 ally according to a physician's prescription, but almost all extracts are manufactured
219 by industrial procedures. There is a general effort to abolish NPPs, with exceptions of
220 rare allergens or special sensitization profiles, whereas a single preparation should
221 contain in the near future only allergens from homologous groups (trees, grasses,
222 mites, and so forth).⁵³ In addition, for each new product, a registration dossier (from
223 phase I to III) is required for the marketing authorization.

225 **THE NEAR FUTURE: PERSPECTIVES**

227 After the introduction of SLI and recent mechanistic studies, there was an impressive
228 advancement in the clinical research on AIT, and new opportunities rapidly appeared
229 (Table 2).

230 The current indication for AIT is allergic rhinoconjunctivitis with/without allergic
231 asthma and hymenoptera venom allergy,^{13,38,39} but for the SLIT tablets approved
232 in the United States, asthma is not an indication. In recent years, many clinical trials
233 have suggested that the indications of AIT can be expanded. In terms of amount of
234 clinical data, the most promising application is food allergy. As discussed elsewhere,
235 there are many clinical trials proving the efficacy of desensitization for cow's milk,
236 peanut, egg, and some other allergenic foods (for review, see Albin and Nowak-
237 Węgrzyn⁵⁵ and Jones and colleagues⁵⁶). Whether administration of gradually
238 increasing amounts of an offending food represents a true AIT or, better, a simple
239 oral induction of tolerance is still not clear. Latex allergy is not an official indication
240 for AIT, although SLIT products are available and commercialized, based on the re-
241 sults of clinical trials.⁵⁷ The same is true for atopic dermatitis, for which both SLIT
242 and SCIT were demonstrated partially effective, especially if a sensitization to dust
243 mite is present.^{58,59}

244 According to current knowledge, the goal of AIT is to take the allergen into contact
245 with antigen-presenting cells to develop an immunologic desensitization. This can be
246 made, in addition to the subcutaneous or sublingual route, by administering an Q22
247 allergen directly into lymph nodes. An innovative clinical trial⁶⁰ supports this rationale,
248 showing that the intralymphatic immunotherapy (ILIT) requires much lower doses of Q23
249 allergen and fewer injections than the traditional SCIT modality, while maintaining
250 the same efficacy.⁶¹ Also, skin is a suitable site for presenting antigens. Epicutaneous
251 immunotherapy (EPIT) has been tested with good results for both aeroallergens and
252 food allergens.⁶² This route seems particularly suitable in children.

Table 2
The future developments of allergen immunotherapy

Advancement	Description	Comments
Route of administration	ILIT Epicutaneous Intradermal	The ILIT allows short courses of administration with lower doses of antigens. EPIT is totally noninvasive and, therefore, particularly suitable for children.
Formulation	Nanoparticles Slow release/mucoadhesive	At early experimental stage, with positive results in animal models
Extract + adjuvants	Bacteria-derived adjuvants DNA-derived adjuvants	Bacterial adjuvants already are commercially available for SCIT. Low number of injections. DNA-adjuvants are under experimental investigation, with a single human trial.
Peptides	Long or short peptides	Under investigation, mainly with Fel d 1 allergen
Molecules	Recombinant/highly purified sensitizing molecules	Some trials available in humans. The single molecules seem not to perform better than the crude extracts.
New indications	Food allergy Atopic dermatitis Latex allergy Nickel allergy?	Despite the existence of numerous trials with positive results, none of these indications is currently approved for clinical practice. Latex SLIT products are commercialized and used.

The products commonly used for AIT are crude extracts, derived from allergenic sources (eg, grasses, ragweed, and mite) and, therefore, contain allergenic and nonallergenic proteins and carbohydrates or lipids. They can be improved by adding adjuvants, which provide an additional enhancement of the T_H1 response. An organic adjuvant usually stimulates the Toll-like receptors of the innate immunity, which in turn favor the T_H1 -oriented response.⁶³ Monophosphoryl lipid A, derived from the cell wall of *Salmonella minnesota*, is proved safe, effective, and capable of reducing the number of injections and the dose of allergen and is currently commercialized. Many other trials with adjuvants are ongoing.⁶⁴ Also prokaryote-derived oligodeonucleotides (CpG sequences) are good adjuvants, because they stimulate the Toll-like receptor 9, with a consequent increase in the T_H1 response. Early trials using this approach provided encouraging results,^{65,66} but the clinical research remains at the initial stage. Another possible manipulation is to give only allergenic fragments, instead of the whole allergenic proteins, because antigen-presenting cells recognize linear sequences; this is called peptide-based immunotherapy. There are so far some promising studies with mixtures of peptides from cat and mite allergens.⁶⁷

As discussed previously, it is now possible to synthesize (or highly purify) the most relevant single sensitizer proteins. Thus, if identifying for each subject the allergenic components toward which IgE are directed, it would be possible to vaccinate only with those molecules (tailored immunotherapy). Nonetheless, it seems that the use of single genuine sensitizers does not perform better than the raw extracts.⁶⁸ In addition, the sensitization profile, dissected by molecular diagnosis, is largely variable in each subject.⁶⁹ Finally, the regulatory authorities require a registration trial for each single allergen product. All those considerations, despite the intriguing immunologic rationale, make this approach so far unfeasible.

UNMET NEEDS AND CONCLUDING REMARKS

The body of evidence for SCIT, SLIT, and VIT is robust, as a result of an abundance of clinical and mechanistic trials. Nonetheless, some points to be clarified, and debated aspects are still present (Table 3). For instance, there is a large variability in administration schedules, dosages, and duration of SLIT, which is marketed in numerous countries as NPPs. Only a few products represent exceptions—Oralair (Stallergenes, Antony Cedex, France), Grazax or Grastek (ALK-Abelló, Copenhagen, Denmark), and Ragwitek (Merck, Whitehouse Station, New Jersey)—because they are registered and marketed as pharmaceutical products.⁷⁰ Another critical point is the standardization. Almost all AIT vaccines commercialized are standardized either biologically or immunologically, based on in-house references. Thus, extracts are labeled in units that differ from one manufacturer to another, and comparison among trials and products is only rarely possible.

Again, there is no experimental demonstration that the regimens used are the most appropriate and cost effective, that the pre-coseasonal regimen for pollen allergens is better, or that for perennial allergens a continuous treatment is needed. There is no rigorous study on the optimal duration of an AIT treatment; thus, the current suggestions are only empirical or based on sparse clinical data.^{71,72} The same is partly true for the preventative effect, demonstration of which is based on only 3 controlled open trials.⁷³ Finally, there is great heterogeneity in clinical trials, which affects the robustness of meta analyses, and the reporting of trials is unsatisfactory.^{74,75}

AIT is a cornerstone in the management of respiratory allergic diseases because it is allergen-specific and immunomodulating and may affect disease progression. SLIT has represented a significant advance, offering patients an excellent safety and acceptance profile. From a historical viewpoint, in the past 3 decades there has been an impressive development of this form of treatment, which has lasted more

Problem	Comments
Optimal maintenance dose	Currently fixed only for grass, ragweed, and mite (soluble tablets, single products). The optimal maintenance dose remains to be clearly defined for the remaining relevant allergens.
Optimal maintenance regimen	Is it needed to give an all-year treatment of perennial allergens? Is the pre-coseasonal (coseasonal regimen) more convenient than the continuous one?
Use of multiple allergens	Few studies are available. The efficacy of multiple allergens, even mixed, is poorly defined.
Adherence	Data about adherence with AIT differ among controlled and real-life studies.
Standardization of extracts	The use of in-house references and of different units make the clinical studies not comparable. The potency of the extracts is still yet not well defined.
Standardization of studies	Large heterogeneity among clinical trials (design, patients' selection, dose, duration, and analysis). Reporting is still poor.
Duration and long-lasting effect	The optimal duration of an AIT course is not experimentally defined. The demonstration of long-lasting and preventive effects relies on a small number of clinical trials

355 than 100 years. The most promising fields are the use of AIT in food allergy, the pre-
356 ventative effects, and the improvement of the routes of administration and standard-
357 ization of extracts and protocols.
358

359 REFERENCES

- 360 1. Noon L. Prophylactic inoculation against hay fever. *Lancet* 1911;i:1572–3.
- 361 2. Johansson SGO. The History of IgE: from discovery to 2010. *Curr Allergy Asthma*
362 *Rep* 2011;11:173–7.
- 363 3. Frankland AW, Augustin R. Prophylaxis of summer hay fever and asthma:
364 controlled trial comparing crude grass pollen extracts with isolated main protein
365 component. *Lancet* 1954;1:1055–7.
- 366 4. Johnstone DE, Dutton A. The value of hyposensitization therapy for bronchial
367 asthma in children: a 14-year study. *Pediatrics* 1968;42:793–802.
- 368 5. Hunt KJ, Valentine MD, Sobotka AK, et al. A controlled trial of immunotherapy in
369 insect hypersensitivity. *N Engl J Med* 1978;299:157–61.
- 370 6. Lockey RF, Turkeltaub PC, Olive ES, et al. The Hymenoptera venom study. III:
371 safety of venom immunotherapy. *J Allergy Clin Immunol* 1990;86:775–80.
- 372 7. Lockey RF, Benedict LM, Turkeltaub PC, et al. Fatalities associated with immuno-
373 therapy and skin testing. *J Allergy Clin Immunol* 1987;79:660–4.
- 374 8. Committee on the Safety of Medicines. CSM update. Desensitizing vaccines. *Br*
375 *Med J* 1986;293:948.
- 376 9. Aaronson DW, Gandhi TK. Incorrect allergy injections: allergists' experiences
377 and recommendations for prevention. *J Allergy Clin Immunol* 2004;113:
378 1117–21.
- 379 10. Windom HH, Lockey RF. An update on the safety of specific immunotherapy. *Curr*
380 *Opin Allergy Clin Immunol* 2008;8:571–6.
- 381 11. Canonica GW, Passalacqua G. Noninjection routes for immunotherapy. *J Allergy*
382 *Clin Immunol* 2003;111:437–48.
- 383 12. Scadding K, Brostoff J. Low dose sublingual therapy in patients with allergic
384 rhinitis due to dust mite. *Clin Allergy* 1986;16:483–91.
- 385 13. Bousquet J, Lockey R, Malling HJ. World Health Organization Position Paper.
386 Allergen immunotherapy: therapeutical vaccines for allergic diseases. *J Allergy*
387 *Clin Immunol* 1998;102(4 Pt 1):558–62.
- 388 14. Bousquet J, Van Cauwenberge P. Allergic rhinitis and its impact on asthma. 026
389 *J Allergy Clin Immunol* 2001;108(5 Supp):S146–50.
- 390 15. Bousquet J, Khaltaev N, Cruz AA, et al. Allergic Rhinitis and its Impact on
391 Asthma (ARIA) 2008 update (in collaboration with the World Health Organization,
392 GA2LEN and AllerGen). *Allergy* 2008;63(Suppl 86):8–160.
- 393 16. Romagnani S. Human TH1 and TH2 subsets: doubt no more. *Immunol Today*
394 1991;12:256–7.
- 395 17. James LK, Shamji MH, Walker SM, et al. Long-term tolerance after allergen im-
396 muno-therapy is accompanied by selective persistence of blocking antibodies.
397 *J Allergy Clin Immunol* 2011;127:509–16.
- 398 18. Rolland JM, Gardner LM, O'Hehir RE. Functional regulatory T cells and allergen
399 immunotherapy. *Curr Opin Allergy Clin Immunol* 2010;10(6):559–66.
- 400 19. Böhm L, Maxeiner J, Meyer-Martin H, et al. IL-10 and regulatory T cells cooperate
401 in allergen-specific immunotherapy to ameliorate allergic asthma. *J Immunol*
402 2015;194(3):887–97.
- 403 20. Fujita H, Soyka MB, Akdis M, et al. Mechanisms of allergen-specific immuno-
404 therapy. *Clin Transl Allergy* 2012;2(1):2.
405

- 406 21. Möller C, Dreborg S, Ferdousi HA, et al. Pollen immunotherapy reduces the
407 development of asthma in children with seasonal rhinoconjunctivitis (the PAT-
408 study). *J Allergy Clin Immunol* 2002;109:251–6.
- 409 22. Novembre E, Galli E, Landi F, et al. Coseasonal sublingual immunotherapy
410 reduces the development of asthma in children with allergic rhinoconjunctivitis.
411 *J Allergy Clin Immunol* 2004;114:851–7.
- 412 23. Marogna M, Tomassetti D, Bernasconi A, et al. Preventive effects of sublingual
413 immunotherapy in childhood: an open randomized controlled study. *Ann Allergy*
414 *Asthma Immunol* 2008;101:206–11.
- 415 24. Durham SR, Yang WH, Pedersen MR, et al. Sublingual immunotherapy with once-
416 daily grass-allergen tablets: a randomised controlled trial in seasonal allergic rhi-
417 noconjunctivitis. *J Allergy Clin Immunol* 2006;117:802.
- 418 25. Didier A, Malling HJ, Worm M, et al. Optimal dose, efficacy, and safety of once
419 daily sublingual immunotherapy with a 5-grass pollen tablet for seasonal allergic
420 rhinitis. *J Allergy Clin Immunol* 2007;120:1338.
- 421 26. Creticos PS, Maloney J, Bernstein DI, et al. Randomized controlled trial of a
422 ragweed allergy immunotherapy tablet in North American and European adults.
423 *J Allergy Clin Immunol* 2013;131:1342–9.
- 424 27. Nolte H, Hébert J, Berman G, et al. Randomized controlled trial of ragweed
425 allergy immunotherapy tablet efficacy and safety in North American adults. *Ann*
426 *Allergy Asthma Immunol* 2013;110:450–5.
- 427 28. Mosbech H, Deckelmann R, de Blay F, et al. Standardized quality (SQ) house
428 dust mite sublingual immunotherapy tablet (ALK) reduces inhaled corticosteroid
429 use while maintaining asthma control: a randomized, double-blind, placebo-
430 controlled trial. *J Allergy Clin Immunol* 2014;134:568–75.
- 431 29. Bergmann KC, Demoly P, Worm M, et al. Efficacy and safety of sublingual tablets
432 of house dust mite allergen extracts in adults with allergic rhinitis. *J Allergy Clin*
433 *Immunol* 2014;133:1608–14.
- 434 30. Frew A, Powell JL, Corrigan CJ, et al. Efficacy and safety of specific immuno-
435 therapy with SQ allergen extract in treatment-resistant seasonal allergic rhinocon-
436 junctivitis. *J Allergy Clin Immunol* 2006;117:319–25.
- 437 31. Canonica GW, Bousquet J, Casale T, et al. Sub-lingual immunotherapy World Al-
438 lergy Organization Position Paper 2009. *Allergy* 2009;64(Supp 91):1–59.
- 439 32. Canonica GW, Cox L, Pawankar R, et al. Sublingual immunotherapy: World Allergy
440 Organization position paper 2013 update. *World Allergy Organ J* 2014;7(1):6.
- 441 33. Thompson CA. Sublingual immunotherapy approved for grass pollen allergies.
442 *Am J Health Syst Pharm* 2014;71:770.
- 443 34. Passalacqua G, Albano M, Fregonese L, et al. Randomised controlled trial of
444 local allergoid immunotherapy on allergic inflammation in mite-induced rhinocon-
445 junctivitis. *Lancet* 1998;351:629–32.
- 446 35. Lombardi C, Incorvaia C, Braga M, et al. Administration regimens for sublingual
447 immunotherapy to pollen allergens: what do we know? *Allergy* 2009;64:849–54.
- 448 36. Pajno GB, Caminiti L, Crisafulli G, et al. Direct comparison between continuous
449 and coseasonal regimen for sublingual immunotherapy in children with grass al-
450 lergy: a randomized controlled study. *Pediatr Allergy Immunol* 2011;22:803–7.
- 451 37. Stelmach I, Kaluzińska-Parzyszek I, Jerezynska J, et al. Comparative effect of pre-
452 coseasonal and continuous grass sublingual immunotherapy in children. *Allergy*
453 2012;67:312–20.
- 454 38. Zuberbier T, Bachert C, Bousquet PJ, et al. GA² LEN/EAACI pocket guide for
455 allergen-specific immunotherapy for allergic rhinitis and asthma. *Allergy* 2010;
456 65:1525–30.

- 457 39. Cox L, Nelson H, Lockey R, et al. Allergen immunotherapy: a practice parameter
458 third update. *J Allergy Clin Immunol* 2011;127(1 Suppl):S1–55.
- 459 40. Cox L, Jacobsen L. Comparison of allergen immunotherapy practice patterns
460 in the United States and Europe. *Ann Allergy Asthma Immunol* 2009;103:
461 451–9.
- 462 41. Nelson HS. Multiallergen immunotherapy for allergic rhinitis and asthma. *J Allergy
463 Clin Immunol* 2009;123:763–9.
- 464 42. Malling HJ, Montagut A, Melac M, et al. Efficacy and safety of 5-grass pollen sub-
465 lingual immunotherapy tablets in patients with different clinical profiles of allergic
466 rhinoconjunctivitis. *Clin Exp Allergy* 2009;39:387–93.
- 467 43. Hankin CS, Cox L. Allergy immunotherapy: what is the evidence for cost saving?
468 *Curr Opin Allergy Clin Immunol* 2014;14:363–70.
- 469 44. Senna G, Ridolo E, Calderon M, et al. Evidence of adherence to allergen-specific
470 immunotherapy. *Curr Opin Allergy Clin Immunol* 2009;9:544–8.
- 471 45. Senna G, Lombardi C, Canonica GW, et al. How adherent to sublingual immuno-
472 therapy prescriptions are patients? The manufacturers' viewpoint. *J Allergy Clin
473 Immunol* 2010;126:668–9.
- 474 46. Savi E, Peveri S, Senna G, et al. Causes of SLIT discontinuation and strategies to
475 improve the adherence: a pragmatic approach. *Allergy* 2013;68:1193–5.
- 476 47. Sastre J. Molecular diagnosis in allergy. *Clin Exp Allergy* 2010;40:1442–60.
- 477 48. Canonica GW, Ansoategui IJ, Pawankar R, et al. A WAO - ARIA - GA²LEN
478 consensus document on molecular-based allergy diagnostics. *World Allergy Or-
479 gan J* 2013;6(1):17.
- 480 49. Sastre J, Landivar ME, Ruiz-García M, et al. How molecular diagnosis can
481 change allergen-specific immunotherapy prescription in a complex pollen area.
482 *Allergy* 2012;67(5):709–11.
- 483 50. Passalacqua G, Melioli G, Bonifazi F, et al. The additional values of microarray
484 allergen assay in the management of polysensitized patients with respiratory al-
485 lergy. *Allergy* 2013;68:1029–33.
- 486 51. Melioli G, Passalacqua G, Canonica GW. Novel in silico technology in combina-
487 tion with microarrays: a state-of-the-art technology for allergy diagnosis and man-
488 agement? *Expert Rev Clin Immunol* 2014;10:1559–61.
- 489 52. Baena Cagnani CE, Larenas D, Sisul C, et al. Allergy training and immunotherapy
490 in Latin America: results of a regional overview. *Ann Allergy Asthma Immunol*
491 2013;111(5):415–9.
- 492 53. Kaul S, May S, Lüttkopf D, et al. Regulatory environment for allergen-specific
493 immunotherapy. *Allergy* 2011;66:753–64.
- 494 54. Bonini S. Regulatory aspects of allergen-specific immunotherapy: europe sets
495 the scene for a global approach. *World Allergy Organ J* 2012;5:120–3.
- 496 55. Albin S, Nowak-Węgrzyn A. Potential treatments for food allergy. *Immunol Allergy
497 Clin North Am* 2015;35:77–100.
- 498 56. Jones SM, Burks AW, Dupont C. State of the art on food allergen immunotherapy:
499 oral, sublingual, and epicutaneous. *J Allergy Clin Immunol* 2014;133:318–23.
- 500 57. Nettis E, Delle Donne P, Di Leo E, et al. Latex immunotherapy: state of the art. *Ann
501 Allergy Asthma Immunol* 2012;109:160–5.
- 502 58. Gendelman SR, Lang DM. Sublingual immunotherapy in the treatment of atopic
503 dermatitis: a systematic review using the GRADE system. *Curr Allergy Asthma
504 Rep* 2015;15:498–507.
- 505 59. Bae JM, Choi YY, Park CO, et al. Efficacy of allergen-specific immunotherapy for
506 atopic dermatitis: a systematic review and meta-analysis of randomized
507 controlled trials. *J Allergy Clin Immunol* 2013;132:110–7.

- 508 60. Senti G, Prinz Vavricka BM, Erdmann I, et al. Intralymphatic allergen administra-
509 tion renders specific immunotherapy faster and safer: a randomized controlled
510 trial. *Proc Natl Acad Sci U S A* 2008;105:17908–12.
- 511 61. Witten M, Malling HJ, Blom L, et al. Is intralymphatic immunotherapy ready for
512 clinical use in patients with grass pollen allergy? *J Allergy Clin Immunol* 2013;
513 132:1248–52.
- 514 62. Senti G, von Moos S, Tay F, et al. Determinants of efficacy and safety in epicuta-
515 neous allergen immunotherapy: summary of three clinical trials. *Allergy* 2015.
516 <http://dx.doi.org/10.1111/all.12600>.
- 517 63. Aryan Z, Holgate ST, Radzioch D, et al. A new era of targeting the ancient gate-
518 keepers of the immune system: toll-like agonists in the treatment of allergic rhinitis
519 and asthma. *Int Arch Allergy Immunol* 2014;164:46–63.
- 520 64. Pfaar O, Cazan D, Klimek L, et al. Adjuvants for immunotherapy. *Curr Opin*
521 *Allergy Clin Immunol* 2012;12:648–57.
- 522 65. Creticos PS, Schroeder JT, Hamilton RG, et al. Immune Tolerance Network Group
523 Immunotherapy with a ragweed-toll-like receptor 9 agonist vaccine for allergic
524 rhinitis. *N Engl J Med* 2006;355:1445–55.
- 525 66. Creticos PS, Chen YH, Schroeder JT. New approaches in immunotherapy:
526 allergen vaccination with immunostimulatory DNA. *Immunol Allergy Clin North*
527 *Am* 2004;24:569–81.
- 528 67. Creticos PS. Advances in synthetic peptide immuno-regulatory epitopes. *World*
529 *Allergy Organ J* 2014;7(1):30.
- 530 68. Pauli G, Larsen TH, Rak S, et al. Efficacy of recombinant birch pollen vaccine for
531 the treatment of birch-allergic rhinoconjunctivitis. *J Allergy Clin Immunol* 2008;
532 122:951–60.
- 533 69. Tripodi S, Frediani T, Lucarelli S, et al. Molecular profiles of IgE to Phleum pra-
534 tense in children with grass pollen allergy: implications for specific immuno-
535 therapy. *J Allergy Clin Immunol* 2012;129:834–9.
- 536 70. Passalacqua G, Canonica GW. Sublingual immunotherapy: focus on tablets. *Ann*
537 *Allergy Asthma Immunol* 2015;115(1):4–9.
- 538 71. Nakonechna A, Hills J, Moor J, et al. Grazax sublingual immunotherapy in pre-
539 co-seasonal and continuous treatment regimens: is there a difference in clinical
540 efficacy? *Ann Allergy Asthma Immunol* 2015;114:73–4.
- 541 72. Stelmach I, Sobocińska A, Majak P, et al. Comparison of the long-term efficacy of
542 3- and 5-year house dust mite allergen immunotherapy. *Ann Allergy Asthma Im-*
543 *munol* 2012;109:274–8.
- 544 73. Passalacqua G. Specific immunotherapy: beyond the clinical scores. *Ann Allergy*
545 *Asthma Immunol* 2011;107:401–6.
- 546 74. Casale TB, Canonica GW, Bousquet J, et al. Recommendations for appropriate
547 sublingual immunotherapy clinical trials. *J Allergy Clin Immunol* 2009;124:
548 665–70.
- 549 75. Bousquet PJ, Calderon MA, Demoly P, et al. The Consolidated Standards of
550 Reporting Trials (CONSORT) Statement applied to allergen-specific immuno-
551 therapy with inhalant allergens: a Global Allergy and Asthma European Network
552 (GA(2)LEN) article. *J Allergy Clin Immunol* 2011;127:49.
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