

PROOF COVER SHEET

Journal acronym: IERX
Author(s): Laura De Ferrari, Alessandra Chiappori, Diego Bagnasco, Anna Maria Riccio,
Giovanni Passalacqua and Giorgio Walter Canonica
Article title: Molecular phenotyping and biomarker development: are we on our way towards targeted therapy for severe
asthma?
Article no: 1111763
Enclosures: 1) Query sheet
2) Article proofs

Dear Author,

1. Please check these proofs carefully. It is the responsibility of the corresponding author to check these and approve or amend them. A second proof is not normally provided. Taylor & Francis cannot be held responsible for uncorrected errors, even if introduced during the production process. Once your corrections have been added to the article, it will be considered ready for publication.

Please limit changes at this stage to the correction of errors. You should not make trivial changes, improve prose style, add new material, or delete existing material at this stage. You may be charged if your corrections are excessive (we would not expect corrections to exceed 30 changes).

For detailed guidance on how to check your proofs, please paste this address into a new browser window:
<http://journalauthors.tandf.co.uk/production/checkingproofs.asp>

Your PDF proof file has been enabled so that you can comment on the proof directly using Adobe Acrobat. If you wish to do this, please save the file to your hard disk first. For further information on marking corrections using Acrobat, please paste this address into a new browser window: <http://journalauthors.tandf.co.uk/production/acrobat.asp>

2. Please review the table of contributors below and confirm that the first and last names are structured correctly and that the authors are listed in the correct order of contribution. This check is to ensure that your name will appear correctly online and when the article is indexed.

Sequence	Prefix	Given name(s)	Surname	Suffix
1		Laura	De Ferrari	
2		Alessandra	Chiappori	
3		Diego	Bagnasco	
4		Anna Maria	Riccio	
5		Giovanni	Passalacqua	
6		Giorgio Walter	Canonica	

Queries are marked in the margins of the proofs, and you can also click the hyperlinks below.

Content changes made during copy-editing are shown as tracked changes. Inserted text is in **red font** and revisions have a red indicator ▲. Changes can also be viewed using the list comments function. To correct the proofs, you should insert or delete text following the instructions below, but **do not add comments to the existing tracked changes**.

AUTHOR QUERIES

General points:

1. **Permissions:** You have warranted that you have secured the necessary written permission from the appropriate copyright owner for the reproduction of any text, illustration, or other material in your article. Please see <http://journalauthors.tandf.co.uk/permissions/usingThirdPartyMaterial.asp>.
2. **Third-party content:** If there is third-party content in your article, please check that the rights holder details for re-use are shown correctly.
3. **Affiliation:** The corresponding author is responsible for ensuring that address and email details are correct for all the co-authors. Affiliations given in the article should be the affiliation at the time the research was conducted. Please see <http://journalauthors.tandf.co.uk/preparation/writing.asp>.
4. **Funding:** Was your research for this article funded by a funding agency? If so, please insert ‘This work was supported by <insert the name of the funding agency in full>’, followed by the grant number in square brackets ‘[grant number xxxx]’.
5. **Supplemental data and underlying research materials:** Do you wish to include the location of the underlying research materials (e.g. data, samples or models) for your article? If so, please insert this sentence before the reference section: ‘The underlying research materials for this article can be accessed at <full link> / description of location [author to complete]’. If your article includes supplemental data, the link will also be provided in this paragraph. See <http://journalauthors.tandf.co.uk/preparation/multimedia.asp> for further explanation of supplemental data and underlying research materials.
6. The **CrossRef database** (www.crossref.org/) has been used to validate the references. Changes resulting from mismatches are tracked in **red font**.

AQ1 Please check whether the institution name is set correctly.

AQ2 Both quotes and italics have been used for quotations. Please confirm if any one form can be retained or whether both are present in the original quotation.

AQ3 Please provide notes for all annotations.

AQ4 Please check whether the edit is correct in this reference.

AQ5 Please provide cited date for Ref. [1].

AQ6 Please provide missing volume number/page number for the “Cox et al., 2015” references list entry.

AQ7 Please provide missing year of publication for the “Mosmann et al., 0000” references list entry.

AQ8 Please provide missing volume number/page number for the “Raundhal et al., 2015” references list entry.

AQ9 Please provide missing Publisher location for the “, 2008” references list entry.

How to make corrections to your proofs using Adobe Acrobat/Reader

Taylor & Francis offers you a choice of options to help you make corrections to your proofs. Your PDF proof file has been enabled so that you can mark up the proof directly using Adobe Acrobat/Reader. This is the simplest and best way for you to ensure that your corrections will be incorporated. If you wish to do this, please follow these instructions:

1. Save the file to your hard disk.
2. Check which version of Adobe Acrobat/Reader you have on your computer. You can do this by clicking on the “Help” tab, and then “About”.

If Adobe Reader is not installed, you can get the latest version free from <http://get.adobe.com/reader/>.

3. If you have Adobe Acrobat/Reader 10 or a later version, click on the “Comment” link at the right-hand side to view the Comments pane.

4. You can then select any text and mark it up for deletion or replacement, or insert new text as needed. Please note that these will clearly be displayed in the Comments pane and secondary annotation is not needed to draw attention to your corrections. If you need to include new sections of text, it is also possible to add a comment to the proofs. To do this, use the Sticky Note tool in the task bar. Please also see our FAQs here:
<http://journalauthors.tandf.co.uk/production/index.asp>.

5. Make sure that you save the file when you close the document before uploading it to CATS using the “Upload File” button on the online correction form. If you have more than one file, please zip them together and then upload the zip file.

If you prefer, you can make your corrections using the CATS online correction form.

Troubleshooting

Acrobat help: <http://helpx.adobe.com/acrobat.html>

Reader help: <http://helpx.adobe.com/reader.html>

Please note that full user guides for earlier versions of these programs are available from the Adobe Help pages by clicking on the link “Previous versions” under the “Help and tutorials” heading from the relevant link above. Commenting functionality is available from Adobe Reader 8.0 onwards and from Adobe Acrobat 7.0 onwards.

Firefox users: Firefox’s inbuilt PDF Viewer is set to the default; please see the following for instructions on how to use this and download the PDF to your hard drive:

http://support.mozilla.org/en-US/kb/view-pdf-files-firefox-without-downloading-them#w_using-a-pdf-reader-plugin

EXPERT
REVIEWS

Molecular phenotyping and biomarker development: are we on our way towards targeted therapy for severe asthma?

Expert Rev. Respir. Med. 00(00), 1-10 (2015)

Laura De Ferrari[†],
Alessandra Chiappori[‡],
Diego Bagnasco,
Anna Maria Riccio,
Giovanni Passalacqua
and Giorgio Walter
Canonica*

Respiratory Diseases and Allergy
Clinic, DIMI-Department Internal
Medicine, University of Genoa, IRCCS
AOUS. Martino-IST, Genoa, Italy
†L.D.F. and A.C. contributed equally to
this work.

*Author for correspondence:
Tel.: +39 010 554 890
Fax: +390 105 556 307
canonica@unige.it

Although different phenotypes of severe asthma can be identified, all are characterized by common symptoms. Due to their heterogeneity, they exhibit differences in pathogenesis, etiology and clinical responses to therapeutic approaches. The identification of distinct molecular phenotypes to define severe asthmatic patients will allow us to better understand the pathophysiology of the disease and thus to more precisely target the treatment for each patient. To achieve this goal, a systematic search for new, reliable and stable biomarkers specific for each phenotype is essential. This review focuses on the current known molecular phenotypes of severe asthma and highlights the need for biomarkers that could (either alone or in combination) be predictive of the treatment outcome.

KEYWORDS: severe asthma • molecular phenotypes • biomarkers • personalized medicine • target therapy • monoclonal antibodies • T helper 2

Bronchial asthma is a highly prevalent chronic respiratory disease. It may be heterogeneous in etiology, pathogenesis, clinical manifestations and outcomes. Comorbidities, pathogenic aspects, symptom severity, natural history, treatment responsiveness and extrinsic and/or intrinsic factors are primarily responsible for this variability. According to the Global Initiative for Asthma Guidelines, asthma severity is classifiable based on different levels of intensity, with an estimated 5–10% of patients suffering from severe asthma.[1] As recently described by the American Thorax Society/ European Respiratory Society Taskforce, “severe asthma requires treatment with high dose inhaled corticosteroids plus a second controller (and/or systemic corticosteroids) to prevent it from becoming uncontrolled or which remains uncontrolled despite this therapy.”[2] This context demonstrates the true refractory nature of asthma.[3]

Despite the fact that pathogenic mechanisms of severe asthma have been detailed over the

years, only a single clinical approach remains available for most patients. Over the last few decades, technological advances have achieved relevant progress, thereby providing significant improvements in the understanding and management of severe asthma.[4,5] Data from genomics, proteomics, transcriptomics, metabolomics and epigenetics studies demonstrated distinct functional and pathological mechanisms related to specific biomarkers and different treatment responses for the severe asthma phenotypes. This clinical entity was introduced into clinical practice several years ago and became increasingly known, accepted and shared.[6] As mentioned above, key elements are needed to cluster and better characterize patients to identify the phenotypes. Recently, it has been established that a single clinical characteristic is not adequate to define a phenotype if it is not integrated with several other factors. This integration of genetics, biology and clinical aspects should lead to better

AQ1

40

45

50

AQ2

55

60

identified phenotypes matched to biomarkers and provide prognostic and potential therapeutic information, thereby resulting in an improved understanding of asthma pathogenesis and an increased ability to effectively treat complex heterogeneous diseases such as severe asthma. A successful identification of reliable and specific biomarkers for severe asthma should be the basis for the introduction of biomarker-related personalized medicine, where clinicians could be oriented to a specific therapeutic approach and severe asthmatics could receive the most effective drug or biological agent on the basis of their single profile.[7–9] In this review, **the authors** will focus on severe asthma and provide an overview of the evolving biomarkers, molecular phenotypes and target-related therapies, paying special attention to lymphocyte Th2 and non-Th2 pathways.

Biomarkers of severe asthma: an overview

In clinical practice, an ideal biomarker is a physical trait or a laboratory measurement that is reliable and reproducible and can help identify the best therapeutic approach, track changes in disease activity, confirm a diagnosis or predict a response. It should also be easy to collect and evaluate, noninvasive and inexpensive.[10,11] For example, in asthma and airway disease management, biomarkers can be assessed in the sputum,[12–15] bronchoalveolar lavage fluid (BALF),[16,17] bronchial epithelial brushings,[16,18] bronchial biopsies,[19,20] exhaled air via fractional exhaled nitric oxide (FeNO) evaluation,[13,21–24] exhaled breath condensate (EBC) analysis,[25–28] peripheral blood [12,13,29] and urine.[30,31] Diagnostic procedures such as bronchoalveolar lavage, bronchoscopy or bronchial biopsy are sometimes additionally used to evaluate airway inflammation and remodeling and to provide detailed pathological information on the patient's condition. Nonetheless, these are invasive techniques that may be risky and are not easily applicable in severe asthma patients. Compared to bronchoalveolar lavage and endoscopy with bronchial specimen collection, spontaneous or induced sputum cell counts have been the method of choice to investigate the inflammatory pattern of bronchial tissues for a long time.[32] Subjects with a concurrent increase in eosinophils ($\geq 2\%$) and neutrophils ($\geq 40\%$) usually have the clinical characteristics of severe asthma.[33] Several studies have demonstrated the high reproducibility of sputum eosinophil and neutrophil counts and proposed them as an index for the therapeutic adjustment and prediction of exacerbation or steroid responsiveness.[34,35] Blood eosinophilia was recently confirmed to be the best surrogate marker for the identification of sputum eosinophilia in moderate to severe asthma patients; this analysis was more specific and accurate than FeNO levels and serum periostin.[13] Over time, sputum and the other above-mentioned samples have become substrates for the implementation of new clinical research techniques and trials. The aim of previous studies was essentially to investigate cytokines (primarily Th1/Th2), chemokines, growth factors, metabolites, proteins and gene expression with a recognized or hypothetical pathogenic role in severe asthma. To date, the most relevant cells and mediators considered to be promising biomarkers are those belonging to the type 2 immune

response pathway; these biomarkers are known to be inherent to severe allergic asthma, which is the most common form of severe asthma.[2] The existence of two different functional subsets of human CD4⁺ T cells (termed Th1 and Th2) that produce distinct cytokine panels was demonstrated in mice in 1986 by Mossman *et al.* and later in human T cells.[36,37] During allergic sensitization and the immune response to allergic stimuli, the respiratory epithelium produces cytokines and chemokines such as thymic stromal lymphopoietin, IL-25 and IL-33. These molecules enhance the Th2 immune response by activating pulmonary dendritic cells and inducing the maturation of CD4⁺ progenitor cells towards a CD4⁺ Th2 functional cellular profile.[38] Activated Th2 cells produce specific cytokines (i.e., IL-4, IL-5, IL-9 and IL-13) that favor and enhance the Th2 response. IL-5 is the primary specific trigger for eosinophils. It drives the hematopoietic process from their bone marrow progenitors (CD34+IL-5R α +) to their release, activation and tissue recruitment.[39] IL-9, which was previously classified as an exclusive Th2 cytokine, is also secreted by another novel subtype of CD4⁺ T cells termed Th9 cells. IL-9 induces T cell activation and mast cell proliferation and differentiation.[40] IL-4 and IL-13 are crucial for type 2 inflammation. These cytokines share a common receptor sub-chain (IL-4R α) that is present in both type 1 (IL-4 exclusive) and type 2 (IL-4 and IL-13) receptors. IL-4 and IL-13 synergistically promote the allergen-specific synthesis of immunoglobulin E (IgE) from B lymphocytes. The secreted IgE binds to its receptor Fc ϵ R1 on mast cells and basophils, resulting in its opsonization. Allergen-IgE complexes trigger the release of a cascade of pre-formed pro-inflammatory mediators (cytokines, interleukins, prostaglandins and leukotrienes) that contribute to inflammatory cell recruitment.[41] Moreover, IL-4/IL-13 together play a key role in airway remodeling and inflammation, causing increased NO production, airway hyperresponsiveness, goblet cell hyperplasia, mucus hypersecretion, the induction of epithelial inducible nitric oxide synthase (only IL-13) and periostin expression in bronchial epithelial cells and lung fibroblasts.[42] The measurement of FeNO is a simple, quantitative, noninvasive, reproducible and safe method to assess airway Th2 and eosinophilic inflammation and to monitor the responsiveness of inhaled corticosteroid (ICS) therapy.[22,43] It has been standardized for clinical use in adults and children but is not considered to be an effective biomarker for severe asthma management.[2]

Periostin is a secreted matricellular protein that is involved in chronic inflammation in allergic diseases and tissue remodeling.[44,45] Thus, serum periostin is considered to be a promising marker of Th2 inflammation and airway eosinophilia despite the fact that it is not lung specific.[12,18,46] We recently studied airway remodeling in severe allergic asthma patients treated with omalizumab. We evaluated bronchial biopsies collected pre-treatment and 12 months after anti-IgE therapy using morphometric and proteomic approaches.[19,20] **The authors'** data showed that Galectin-3 could serve as a predictive biomarker of airway remodeling improvement.

Th2 cytokines also trigger the production of eotaxins, which are strong chemoattractant proteins. Eotaxin-1 is involved in

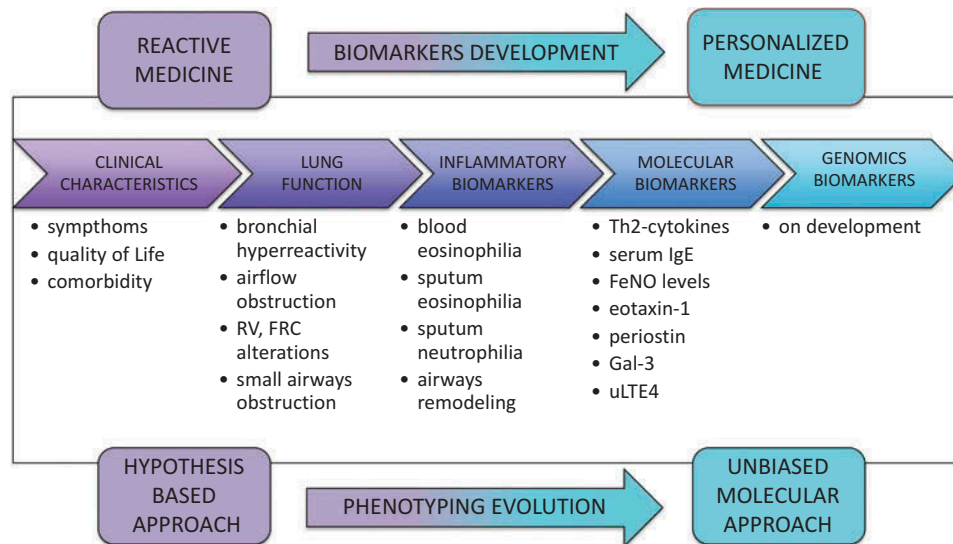


Figure 1. Biomarker discovery and phenotyping in severe asthma: actuality and perspectives. FeNO: Fractional exhaled nitric oxide; FRC: Functional residual capacity; Gal-3: Galectin-3; IgE: Immunoglobulin E; RV: Residual volume; uLTE4: Urinary leukotriene E4.

eosinophil recruitment and activation and has been detected in the blood, sputum, BALF and EBC. Recently, Wu *et al.* proposed eotaxin-1 as a novel potential biomarker for the assessment of asthma severity and control status.[47]

Finally, various markers of airway nitrosative/oxidative stress and inflammation (leukotrienes and lipoxines) have been found to be elevated in EBC from severe asthmatics compared with healthy subjects.[27,48]

Leukotrienes are released from various cells, including eosinophils, neutrophils and mast cells, and can also be detected in the urine. Urinary leukotriene E4 is used as a systemic marker of LT production, and high levels in asthmatic patients seem to be linked to a better response to leukotriene antagonist administration [30] (Figure 1).

Despite ongoing treatment with high doses of ICS, some severe asthmatics present a Th1 inflammatory signature characterized by the production of IFN- γ and reduced Th2 and IL-17 responses.[49] However, Th17 cells (CD4+IL-17+ expressing T cells) have been recognized to play a key role in the promotion of neutrophil selective migration to the site of airway inflammation.[50]

Severe asthma molecular phenotyping

As mentioned above, severe asthma patients are heterogeneous in terms of their severity, natural history and treatment responsiveness; this heterogeneity is partially related to the underlying mechanisms. Nonetheless, they are characterized by common features that allow clinicians to group them into so-called “phenotypes.”[6] In this context, phenotype means “the observable properties of an organism that are produced by the interaction of the genotype and the environment.”[51] Each phenotype exhibits a subtype of a condition that is defined by distinct functional

or pathophysiological mechanisms. These features or pathways are defined as “endotypes.”[6,52] By applying the clinically oriented phenotype definition to traditional medicine, we can say that a “phenotype” is the result of a specific interaction between a patient’s genes and the environment, and many different approaches have been proposed to characterize these subgroups in severe asthma. Clinical, biological and physiological characteristics have been used to classify the different forms of asthma.[53] Severe asthma phenotypes are currently identifiable by two main strategies: a hypothesis-based approach and an unbiased approach. Severe asthma phenotyping began decades ago using a biased approach. Patients were classified into broad categories based upon a single variable, including disease severity, symptomatic triggers, age at onset, patterns of inflammation, exacerbations and airflow obstruction.[54–56] Using this approach, overlaps between different groups were common. In contrast, the statistically unbiased approach attempted to cluster patients to avoid the clinical biases. The Severe Asthma Research Program (SARP) study identified five asthma clusters using an unsupervised hierarchical analysis of the predominant clinical features. Each of the clusters contained subjects who met the ATS definition of severe asthma.[57] Another SARP study was performed on 161 asthmatic children. The authors identified four phenotypic clusters and placed children with severe asthma into each cluster.[58] Recently, Schatz *et al.* performed a *post hoc* cluster analysis on a large number of severe or difficult-to-treat asthmatic children, adolescents and adults belonging to “The Epidemiology and Natural History of Asthma: Outcomes and Treatment Regimens” study cohort. Cluster-defining variables were chosen, including sex, race, atopy, age at onset and others. Five clusters were identified in each age stratum, and the association between clusters and asthma-related health outcomes were

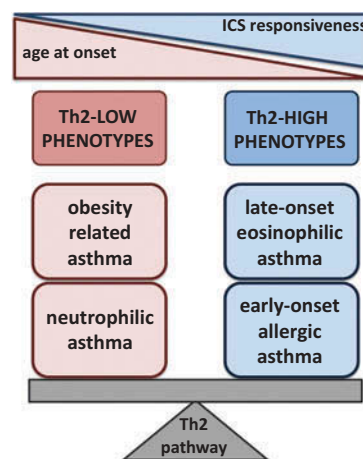


Figure 2. The primary defined Th2-high and Th2-low severe asthma phenotypes. ICS: Inhaled corticosteroid.

235 searched. The results confirmed the existence of distinct phenotypes that were related to outcomes in adults and adolescents but not in children.[59] It became progressively clear that the above-mentioned clinical clusters were not adequate to fully explain and clarify patients' phenotypes. Thus, studies of molecular phenotyping analysis were developed to investigate the molecular pathways underlying the clinical phenotypes. In a randomized control trial, Woodruff *et al.* analyzed airway epithelial brushings from 42 mild-to-moderate nonsmoking asthmatic patients and nonsmoking healthy subjects using microarray and PCR analyses. The authors also investigated airway epithelium gene expression to explore its possible dysfunctional mechanism in asthma and reported that the *chloride channel, calcium-activated, family member 1 (CLCA1)*, *periostin*, and *serine peptidase inhibitor, clade B (ovalbumin)*, and *member 2 (serpinB2)* genes were upregulated in the asthma patients. In *in vitro* studies, the authors found that this enhanced gene expression was stimulated by IL-13.[60] Based on the hypothesis that IL-13 could be used to identify subsets of asthma patients, the gene expression of cells isolated from epithelial brushing were re-evaluated by taking the high or low expression of IL-13-inducible genes into consideration. The experimental results suggested the classification of asthmatic patients into cohorts in two subgroups according to the degree of Th2-driven inflammation. IL-5 and IL-13 expression, airway hyperresponsiveness, ICS responsiveness, serum IgE and blood and airway eosinophilia were significantly increased in the "Th2-high" group versus the "Th2-low" group/healthy controls.[18] The authors analyzed induced sputum cells from 37 asthmatic patients and 15 healthy control subjects to better profile the epithelial cell gene expression following IL-13 exposure. The genes evaluated included periostin, *LCA1* and *SERPINB2*, Th2 genes and other genes involved in the Th2 response. Periostin and *CLCA1* (but not *SerpinB2*) exhibited significantly increased expression in the sputum-obtained cells from asthmatic subjects. Induced sputum proved to be an easy-to-collect biological sample method for molecular evaluation. IL-4, IL-5 and IL-13 transcripts could be easily detected in sputum cells from asthmatic patients, and their expression levels were proposed for use to classify asthma into Th2-high and Th2-low endotypes.[14] Thus, the concept of "severe asthma molecular phenotypes" was defined.

Th2-high phenotypes, their biomarkers and targeted therapy

275 Historically, asthma has been always considered a Th2 disease that is linked to atopy, allergy and eosinophilic inflammation, and therefore, is responsive to corticosteroids (CSs). The majority of patients fit this common context, but asthmatics are a heterogeneous group that range from allergic asthma to aspirin-induced asthma to exercise-induced asthma (EIA) [61] (Figure 2). EIA is usually found in individuals who suffer from bronchoconstriction following strenuous exercise, frequently under cold and dry conditions. In this phenotype, increased levels of mast cells and their mediators can be observed in addition to the Th2 component, although the underlying

inflammatory response is not fully understood and no specific biomarker has been described.[62] These subjects are often responsive to anti-IL-9 therapy and cysteinyl leukotriene modifiers, but they usually have milder asthma and therefore will not be considered in this review.[63]

Overall, as the complexity of the immuno-inflammatory background increases, additional immune pathways are likely to be engaged beyond the original Th2 core, leading to greater severity and the reduction of CS responsiveness.[64] Some biomarkers have been associated with Th2 inflammation, but many of them still need to be understood. Moreover, targeted Th2 pathway inhibition in non-phenotyped patients is less frequently effective, and therefore, new biomarkers need to be found to offer a tailored therapy to the patient (Figure 3).

Early-onset allergic asthma

290 This group accounts for approximately 40–50% of all severe asthmatics and includes subjects who developed their disease during childhood (often school age). These patients are characterized by an atopic condition and Th2 inflammation. Early-onset allergic asthma can present with mild-to-severe forms, and it is not clear whether severe asthma is the result of a progression from a milder form or instead arises as severe during childhood. [58] These patients are polysensitized, have high total IgE levels, a frequent family history of atopy and a clearly identified symptom exposure. Genome-wide association studies have evidenced a genetic component linked to early-onset asthma, and the genes associated with this phenotype are epithelial-related rather than allergy-related.[65] Generally, CS therapy can modulate Th2 cytokines and associated inflammation, but its activity is nonspecific. Thus, treatment with molecules that selectively target components of the Th2 pathway will be more appropriate.[56] High FeNO levels, sputum eosinophils and increased airway periostin have been proposed as biomarkers for Th2-associated asthma and could be used to identify patients responsive to IL-4/IL-13-targeted therapy. Periostin identifies a

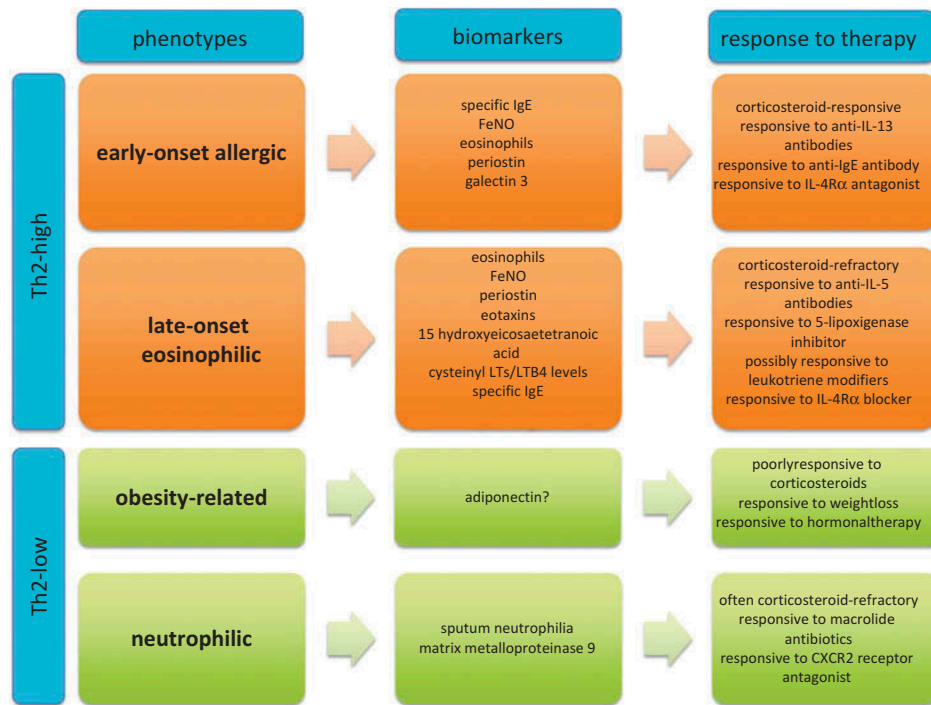


Figure 3. Currently known aspects of asthma phenotypes. CXCR2: CXC chemokine receptor 2; FeNO: Fractional exhaled nitric oxide; Ig: Immunoglobulin; IL4R α : Interleukin 4 receptor alpha; LT: Leukotriene.

subgroup of moderate-to-severe asthmatics with lung eosinophilia who respond to therapy with a monoclonal antibody directed against IL-13 (lebrikizumab), whereas IgE levels, atopy and blood eosinophils do not allow the prediction of a favorable clinical response. Only subjects with a high serum concentration of periostin showed an improvement in lung function parameters, and to a lesser extent exacerbations, following lebrikizumab therapy.[66] Piper *et al.* recently evaluated another anti-IL-13 antibody (tralokinumab). In this study, only patients with sputum IL-13 levels above 10 ng/ml showed a good response, with a significant improvement in the asthma control questionnaire score and forced expiratory volume in 1 s.[67] The presence of high IgE levels makes this group an ideal target for anti-IgE treatment; however, the available data evidenced a modest efficacy of omalizumab in severe asthma patients with high IgE.[68,69] Recently, Hanania *et al.* described a retrospective analysis of a trial with omalizumab using the Th2 biomarkers periostin, FeNO and blood eosinophils. The authors split the asthmatics into two groups: subjects with high and low biomarker values. Subjects with high levels of these biomarkers responded better to omalizumab therapy than subjects with lower levels. Moreover, patients on placebo therapy in the Th2-high group had more frequent exacerbations than patients without Th2-high-related inflammation.[22] Recently, the authors' group demonstrated that the expression of Galectin-3 in bronchial biopsies of allergic severe asthma patients could also be used to predict the modulation of airway remodeling following omalizumab treatment.[20] In a 12-week study, Corren

et al. failed to demonstrate significant efficacy of an antibody to IL-4R α in severe asthma patients, although a modest benefit was seen in the most symptomatic subjects.[70] Recently, the analysis of IL-4R α polymorphisms allowed the identification of participants with a more favorable IL-4R α antagonist therapy response.[71,72] A monoclonal antibody to IL-5 reduced blood eosinophils but did not show significant efficacy in these patients, suggesting that other inflammatory components may have a more important role than eosinophils. Additional data would certainly improve our knowledge concerning the pathological mechanisms involved in this phenotype and enhance our tools for the prediction of the degree of the treatment response.

Late-onset highly eosinophilic asthma

Several studies with patients characterized by late-onset disease have observed persistent blood and lung eosinophilia despite treatment with inhaled and oral CS $_2$. This phenotype represents a more heterogeneous and less well-defined group of subjects and includes approximately 25% of severe asthma patients.[56] Adult eosinophilic asthma is severe at the time of onset and is sometimes associated with chronic sinusitis and nasal polyps without a clear atopy history. Moreover, this group also includes patients with aspirin-exacerbated respiratory disease (AERD). AERD refers to patients with non-IgE-mediated responses to aspirin and other cyclooxygenase-1 inhibitors; the cysteinyl leukotriene pathway in these patients is upregulated by Th2 cytokines. Eosinophilia is not specific for this phenotype, suggesting that other biomarkers may be

more relevant. Many patients are characterized by a strong Th2 skew, with increased FeNO, periostin, eotaxins, 15-lipoxygenase-1 and 15-hydroxyicosatetraenoic acid.[16,73] Moreover, an increase in urinary cysteinyleukotrienes can be observed. Generally, eosinophils undergo apoptosis in the presence of CS_s because Th2 inflammation is usually CS_s sensitive.[74] In this phenotype, a persistent eosinophilia is indicative of CS refractoriness, and high systemic doses are able to overcome this CS inefficacy.[75] The lack of a significant response to CS_s treatment in this phenotype implies that the Th2 process differs from the process associated with the early-onset phenotype and is probably more complex. However, late-onset persistent eosinophilic asthma is associated with a better response to Th2-targeted therapy than early-onset allergic asthma, and the presence of eosinophilic inflammation detected in blood (or preferably sputum) identifies patients responsive to selective anti-eosinophilic therapies, including anti-IL-5.[76] IL-5 is a quite selective cytokine that is involved in the recruitment, maturation and activation of eosinophils, and therefore, is an optimal target for treatment. In these patients, the monoclonal antibody to IL-5 (mepolizumab) decreased the exacerbation rate, allowed systemic CS withdrawal and improved lung function.[77,78] Another anti-IL-5 monoclonal antibody (reslizumab) improved airway function compared with the placebo.[79] Moreover, beneficial effects on lung function in these individuals could also be obtained with 5-lipoxygenase inhibitors and leukotriene modifiers.[80] Finally, Wenzel *et al.* found that the IL-4R α blocker pitrakinra was significantly better at reducing the exacerbation rate compared with the placebo in asthma patients with baseline blood eosinophils >350 mm³. [81] Pitrakinra is a recombinant protein with a double inhibitory effect on IL-4 and IL-13. It acts as a competitive antagonist to prevent cell activation by binding to the α subunit of the IL-4 receptor, which is shared by both cytokines. Moreover, dupilumab (a human monoclonal antibody to IL-4R α) seems to be associated with fewer asthma exacerbations and improved lung function in moderate-to-severe asthma patients with elevated eosinophil levels.[82]

Th2-low phenotypes, their biomarkers and targeted therapy

Th2-low asthma is generally identified by the absence of a clear Th2 aspect because several other factors (i.e., Th1 and Th17 immunity, neutrophil inflammation, mast cell activation, smooth muscle hyperreactivity, infection and oxidative stress) seem to play more important roles. Th2-low asthma probably represents a relevant percentage of all severe asthmatic patients, and the lack of efficacy of Th2-targeted treatments, including CS_s, supports the existence of asthma phenotypes not characterized by Th2 immunity. However, because the phenotypes included in this group are not well defined, detailed studies are needed to enable the definition of new phenotypes, specific biomarkers and clinical/pathological features.[83,84]

Obesity-related asthma

These patients are most commonly women with severe symptoms despite a moderately preserved lung function. The patients typically have a high body mass index, onset of disease in the fifth decade or later and less frequent history of allergic diseases.[85] The obesity condition is associated with a generalized inflammatory state with increased TNF- α , IL-6 and leptin expression, but confirmatory data are needed to identify potentially useful specific biomarkers.[86] Murine models of obesity demonstrated that adiponectin played a protective role and was inversely correlated with lung inflammation; however, in humans these data are discordant.[87–89] Obesity-associated asthma is usually less responsive to inhaled CS_s therapy than allergic asthma.[18] Whether the lack of response to CS_s is due to additional immune pathways is not fully understood. Dixon *et al.* observed improvements in symptoms, quality of life and bronchial hyperresponsiveness in a group of nonallergic, obese, late-onset subjects after weight loss was achieved following bariatric surgery [90]; the same result was reported by Lombardi *et al.*[91]

Neutrophilic asthma

Several authors have detected increased neutrophil counts in the sputum, BALF and biopsy tissues from adult-onset, severely obstructed patients that were associated with a high dose of CS therapy and sometimes with high past/current tobacco use. Neutrophilia can coexist with eosinophilia. However, the definition of the specific role of neutrophils in asthma pathogenesis is difficult because neutrophilic inflammation can also be the result of concomitant diseases or can be derived from the CS inhibitory effect on the apoptosis of these cells.[92] Gene expression analysis of cells from sputum found increased expression of the TNF- α pathway and IL-1 concentrations in patients with neutrophilic asthma, although anti-TNF- α therapy did not show significant clinical efficacy in these subjects.[93,94] Enhanced expression of α -defensins and neutrophil proteases was also detected in the peripheral blood of these patients.[95] Moreover, the presence of neutrophils was associated with increased levels of matrix metalloproteinase 9 in the BALF and tissues, and these high concentrations were not reduced by CS treatment. Whether Th17 lymphocytes are involved in the pathological process is not clear because the majority of the data are derived from experimental models and studies in humans have not associated Th17 pathways with neutrophil asthma inflammation.[96] Currently, there are few neutrophil asthma-targeting drugs. Macrolide antibiotics have an anti-neutrophilic activity, and recently a CXC chemokine receptor 2 antagonist was shown to decrease sputum neutrophils in severe asthmatics.[97,98]

Other possible phenotypes

It is very likely that new distinct phenotypes will be added in the future to those described here based on current knowledge.[99] Further studies are necessary to better characterize severe asthma patients. Many authors have agreed with the existence of a paucigranulocytic phenotype characterized by the absence of an

485 observable inflammatory process and a general lack of both sputum
eosinophils and neutrophils. In this case, the underlying pathological
mechanisms are poorly understood and add further difficulty to the
selection of appropriate therapeutic approaches. Furthermore, smok-
ing-associated asthma may also be a distinct phenotype that overlaps
with chronic obstructive pulmonary disease.[100]

490 Conclusions

The so-called “personalized medicine” is expected to be more
advantageous and possibly more cost-effective than the current
diagnostic and clinical approach. A number of important aspects
are still under investigation, including the identification of biomar-
495 kers that can be used to predict patient progression and therapeutic
responses.[101,102] A more detailed investigation into biomarkers
may help clinicians determine the best treatment based on the
phenotype and the expected response. “Omics” approaches (genom-
ics, proteomics and metabolomics) are expected to be of great
500 help in identifying biomarkers and phenotypes. Currently, several
drugs acting at different levels in the cascade of reactions involved
in Th2 pathogenesis are under investigation, whereas there has
been less progress for the so-called non-Th2 asthma (the obesity-
related, neutrophilic and paucigranulocytic phenotypes).[103]

505 The identification of new asthma phenotypes is essential to make
the best use of the new monoclonal drugs still under study and in the
experimental phases. The possibility of introducing these biologicals
in therapy allows us to integrate high dosage therapy that often fails
to control the symptoms of patients with severe asthma. The coop-
510 eration between academia, research and the pharmaceutical industry
could provide new resources and tools to seek new phenotypes of
severe asthma and related biomarkers for characterization.[104]

Expert commentary

515 The advances in the field of biomedical research are changing the
world and contributing to the expansion and improvement of our
knowledge, with important, crucial consequences for the manage-
ment of complex and difficult to treat diseases such as severe
asthma. Along the pathway of hard sciences, we progressively
abandoned traditional reactive medicine, mass targeted therapy

and descriptive studies. According to the concept of “heterogeneity
of severe asthma,” we focused our resources on phenotyping and
investigating molecular pathways that led to more stratified med-
icine. Many biological drugs for Th2 pathway targets and specific
biomarkers for severe asthma phenotypes and endotypes are now
525 available in clinical practice. However, the state of the art in
biomarker discovery and phenotyping still does not allow us to
plan a personalized therapeutic approach and to predict the patho-
logical evolution of severe asthma patients.

Five-year view

530 The recent advances in immunology, genetics and “omics” sciences
have allowed us to better understand complex diseases such as
severe asthma, which still represents a diagnostic and therapeutic
challenge for clinicians. Research has focused on an improved and
refined definition of the phenotypes and the identification of
535 biomarkers that will allow a more accurate diagnosis and targeted
prescription. According to recent opinions, it is important to move
from generalized and standardized medicine, where every severe
asthma patient is treated only on the basis of his/her symptoms, to
personalized medicine where each subject will receive the best
540 targeted therapy.[105] Severe asthma patients should be referred
to highly specialized Asthma Units whenever possible to optimize
both patient outcomes and the cost-effectiveness of the new bio-
logical agents and to improve our current knowledge of asthma
mechanisms. This integrated approach by experienced clinicians
545 and researchers would surely help to identify the definitive pathway
to personalized medicine for asthma.

Financial & competing interests disclosure

*The authors have no relevant affiliations or financial involvement
with any organization or entity with a financial interest in or
550 financial conflict with the subject matter or materials discussed in
the manuscript. This includes employment, consultancies, honoraria,
stock ownership or options, expert testimony, grants or patents received
or pending, or royalties. Writing assistance was used in the preparation
of this manuscript.* 555

Key issues

- The heterogeneity of severe asthma limits its characterization and makes the selection of appropriate therapeutic approaches difficult.
- The response to an anti-inflammatory drug depends on the presence of a specific type of airway inflammation.
- A biomarker can be indicative of a specific inflammatory process.
- A biomarker can be useful for the prediction of the degree of treatment response.
- New biomarkers need to be identified to allow clinicians to choose the most appropriate targeted therapy for each patient.
- In many cases, the characterization of patients with severe asthma with a single biomarker is not sufficient to identify the specific phenotype.
- Understanding the immune pathways beyond severe asthma inflammation is moving the concept of clinical phenotypes towards the concept of molecular phenotypes.
- Panels of different, noninvasive, stable biomarkers should be studied for application in daily clinical practice.
- Severe asthma patients should be referred to specialized units for optimal disease management.

References

Papers of special note have been highlighted as:

* of interest

** of considerable interest

1. Global Initiative for Asthma (GINA). From the Global Strategy for Asthma Management and Prevention. 2015. Available from: <http://www.ginasthma.org/>.
2. Chung KF, Wenzel SE, Brozek JL, et al. International ERS/ATS guidelines on definition, evaluation and treatment of severe asthma. *Eur Respir J*. 2014;43:343–373.
3. Bousquet J, Mantzouranis E, Cruz AA, et al. Uniform definition of asthma severity, control, and exacerbations: document presented for the World Health Organization Consultation on Severe Asthma. *J Allergy Clin Immunol*. 2010;126:926–938.
4. Hamburg MA, Collins FS. The path to personalized medicine. *N Engl J Med*. 2010;363:301–304. Erratum in: *N Engl J Med*. 2010;363:1092.
5. Cox C, Kjarsgaard M, Surette MG, et al. A multidimensional approach to the management of severe asthma: Inflammometry, molecular microbiology and bronchial thermoplasty. *Can Respir J*. 2015; pii: 16891Epub ahead of print
6. Wenzel S. Severe asthma: from characteristics to phenotypes to endotypes. *Clin Exp Allergy*. 2012;42:650–658.
7. Gustafsson M, Nestor CE, Zhang H, et al. Modules, networks and systems medicine for understanding disease and aiding diagnosis. *Genome Med*. 2014;6:82.
8. Drazen JM. A step toward personalized asthma treatment. *N Engl J Med*. 2011;365:1245–1246.
9. Braido F, Holgate S, Canonica GW. From “blockbusters” to “biosimilars”: an opportunity for patients, medical specialists and health care providers. *Pulm Pharmacol Ther*. 2012;25:483–486.
10. Cazzola M, Novelli G. Biomarkers in COPD. *Pulm Pharmacol Ther*. 2010;23:493–500.
11. Rossi R, De Palma A, Benazzi L, et al. Biomarker discovery in asthma and COPD by proteomic approaches. *Proteomics Clin Appl*. 2014;8:901–915.
12. Jia G, Erickson RW, Choy DF, et al. Periostin is a systemic biomarker of eosinophilic airway inflammation in asthmatic patients. *J Allergy Clin Immunol*. 2012;130:647–654.
13. Wagener AH, De Nijs SB, Lutter R, et al. External validation of blood eosinophils, FE(NO) and serum periostin as surrogates for sputum eosinophils in asthma. *Thorax*. 2015;70:115–120.
14. Peters MC, Mekonnen ZK, Yuan S, et al. Measures of gene expression in sputum cells can identify TH2-high and TH2-low subtypes of asthma. *J Allergy Clin Immunol*. 2014;133:388–394.
15. Oreo KM, Gibson PG, Simpson JL, et al. Sputum ADAM-8 expression in increased in severe asthma and COPD. *Clin Experim Allergy*. 2013;44:342–352.
16. Coleman JM, Naik C, Holguin F, et al. Epithelial eotaxin-2 and eotaxin-3 expression: relation to asthma severity, luminal eosinophilia and age at onset. *Thorax*. 2012;67:1061–1066.
17. Hosoki K, Ying S, Corrigan C, et al. Analysis of a panel of 48 cytokines in BAL fluids specifically identifies IL-8 levels as the only cytokine that distinguishes controlled asthma from uncontrolled asthma, and correlates inversely with FEV1. *PLoS One*. 2015;10: e0126035.
18. Woodruff PG, Modrek B, Choy DF, et al. T-helper type 2-driven inflammation defines major subphenotypes of asthma. *Am J Respir Crit Care Med*. 2009;180:388–395.
19. Riccio AM, Dal Negro RW, Micheletto C, et al. Omalizumab modulates bronchial reticular basement membrane thickness and eosinophil infiltration in severe persistent allergic asthma patients. *Int J Immunopathol Pharmacol*. 2012;25:475–484.
20. Mauri P, Riccio AM, Rossi R, et al. Proteomics of bronchial biopsies: galectin-3 as a predictive biomarker of airway remodelling modulation in omalizumab-treated severe asthma patients. *Immunol Lett*. 2014;162(1 Pt A):2–10.
21. Gemiciglu B, Musellim B, Dogan I, et al. Fractional exhaled nitric oxide (FeNo) in different asthma phenotypes. *Allergy Rhinol*. 2014;5:157–161.
22. Hanania NA, Wenzel S, Rosen K, et al. Exploring the effects of Omalizumab in allergic asthma. an analysis of biomarkers in the EXTRA study. *Am J Respir Crit Care Med*. 2013;187:804–811.
23. Peirsman EJ, Carvelli TJ, Hage PY, et al. Exhaled nitric oxide in childhood allergic asthma management: a randomised controlled trial. *Pediatr Pulmonol*. 2014;49:624–631.
24. Yang S, Park J, Lee YK, et al. Association of longitudinal fractional exhaled nitric oxide measurements with asthma control in atopic children. *Respir Med*. 2015;109:572–579.
25. Di Gangi IM, Pirillo P, Carraro S, et al. Online trapping and enrichment ultra-performance liquid chromatography-tandem mass spectrometry method for sensitive measurement of “arginine-asymmetric dimethylarginine cycle” biomarkers in human exhaled breath condensate. *Anal Chim Acta*. 2012;754:67–74.
26. Carraro S, Giordano G, Reniero F, et al. Asthma severity in childhood and metabolomic profiling of breath condensate. *Allergy*. 2013;68:110–117.
27. Kazani S, Planaguma A, Ono E, et al. Exhaled breath condensate eicosanoid levels associate with asthma and its severity. *J Allergy Clin Immunol*. 2013;132:547–553.
28. Schwarz K, Biller H, Windt H, et al. Characterization of exhaled particles from the human lungs in airway obstruction. *J Aerosol Med Pulm Drug Deliv*. 2015;28:52–58.
29. Katz LE, Gleich GJ, Hartley BF, et al. Blood eosinophil count is a useful biomarker to identify patients with severe eosinophilic asthma. *Ann Am Thorac Soc*. 2014;11:531–536.
30. Cai C, Yang J, Hu S, et al. Relationship between urinary cysteinyl leukotriene E4 levels and clinical response to antileukotriene treatment in patients with asthma. *Lung*. 2007;185:105–112.
31. Mattarucchi E, Baraldi E, Guillou C. Metabolomics applied to urine samples in childhood asthma; differentiation between asthma phenotypes and identification of relevant metabolites. *Biomed Chromatogr*. 2012;26:89–94.
32. Bossley CJ, Fleming L, Gupta A, et al. Pediatric severe asthma is characterized by eosinophilia and remodeling without T (H)2 cytokines. *J Allergy Clin Immunol*. 2012;129:974–82.e13.

33. Moore WC, Hastie AT, Li X, et al. Sputum neutrophil counts are associated with more severe asthma phenotypes using cluster analysis. *J Allergy Clin Immunol*. 2014;133:1557-63.e5.
34. Simpson JL, McElduff P, Gibson PG. Assessment and reproducibility of non-eosinophilic asthma using induced sputum. *Respiration*. 2010;79:147-151.
- AQ8 35. Rossall MR, Cadden PA, Molphy SD, et al. Repeatability of induced sputum measurements in moderate to severe asthma. *Respir Med*. 2014;108:1566-1568.
- AQ7 36. Mosmann TR, Cherwinski HM, Bond MW, et al. *J Immunol*. 136:2348-2357.
37. Del Prete GF, De Carli M, Mastromauro C, et al. *J Clin Invest*. 1991;88:346-350.
- AQ9 38. Dunn RM, Wechsler ME. Anti-interleukin therapy in asthma. *Clin Pharmacol Ther*. 2015;97:55-65.
39. Mory Y, Iwasaki H, Kohno K, et al. Identification of the human eosinophil lineage-committed progenitor: revision of phenotypic definition of the human common myeloid progenitors. *J Exp Med*. 2009;206:183-193.
40. Farahani R, Sherkat R, Hakemi MG, et al. Cytokines (interleukin-9, IL-17, IL-22, IL-25 and IL-33) and asthma. *Adv Biomed Res*. 2014;3:127.
41. Siracusa MC, Comeau MR, Artis D. New insights into basophils biology: initiators, regulators, and effectors of type 2 inflammation. *Ann N Y Acad Sci*. 2011;1217:166-177.
42. Vatrella A, Fabozzi I, Calabrese C, et al. Dupilumab: a novel treatment for asthma. *J Asthma Allergy*. 2014;7:123-130.
43. Dweik RA, Boggs PB, Erzurum SC, et al. An Official ATS Clinical Practice Guideline: Interpretation of Exhaled Nitric Oxide Levels (FENO) for Clinical Applications. *Am J Respir Crit Care Med*. 2011;184:602-615.
44. Conway SJ, Izuhara K, Kudo Y, et al. The role of periostin in tissue remodeling across health and disease. *Cell Mol Life Sci*. 2014;71:1279-1288.
45. Sidhu SS, Yuan S, Innes AL, et al. Roles of epithelial cell-derived periostin in TGF-beta activation, collagen production, and collagen gel elasticity in asthma. *Proc Natl Acad Sci USA*. 2010;107:14170-14175.
46. Matsumoto H. Serum periostin: a novel biomarker for asthma management. *Allergol Int*. 2014;63:153-160.
47. Wu D, Zhou J, Bi H, et al. CCL11 as a potential diagnostic marker for asthma?. *J Asthma*. 2014;51:847-854.
48. Comhair SA, Erzurum SC. Redox control of asthma: molecular mechanisms and therapeutic opportunities. *Antioxid Redox Signal*. 2010;12:93-124.
49. Raundhal M, Morse C, Khare A, et al. High IFN-gamma and slow SLPI mark severe asthma in mice and humans. *J Clin Invest*. 2015; [Epub ahead of print].
50. Roussel L, Houle F, Chan C, et al. IL-17 promotes p38 MAPK-dependent endothelial activation enhancing neutrophil recruitment to sites of inflammation. *J Immunol*. 2010;184:4531-4537.
51. Merriam-Webster's Collegiate Dictionary English dictionary, 12th. Meriam-Webster, Inc.; 2008.
52. Lotvall J, Akdis CA, Bacharier LB, et al. Asthma endotypes: a new approach to classification of disease entities within the asthma syndrome. *The Journal of Allergy and Clinical Immunology*. 2011;127:355-360.
53. Chung KF. Defining phenotypes in asthma: a step towards personalized medicine. *Drugs*. 2014;74:719-728.
54. Wenzel SE, Schwartz LB, Langmack EL, et al. Evidence that severe asthma can be divided pathologically into two inflammatory subtypes with distinct physiologic and clinical characteristics. *Am J Respir Crit Care Med*. 1999;160:1001-1008.
- ** **Authors identified for the first time neutrophilic inflammation in mild asthmatics and found an association between clinical variables and different amounts of eosinophilic inflammation.**
55. Gibson PG, Simpson JL, Hankin R, et al. Relationship between induced sputum eosinophils and the clinical pattern of childhood asthma. *Thorax*. 2003;58:116-121.
56. Miranda C, Busacker A, Balzar S, et al. Distinguishing severe asthma phenotypes: role of age at onset and eosinophilic inflammation. *J Allergy Clin Immunol*. 2004;113:101-108.
- ** **An interesting study on phenotypic differences between early-onset severe asthma as compared with late-onset disease.**
57. Moore WC, Meyers DA, Wenzel SE, et al. Identification of asthma phenotypes using cluster analysis in the Severe Asthma Research Program. *Am J Respir Crit Care Med*. 2010;181:315-323. 37.
58. Fitzpatrick AM, Teague WG, Meyers DA, et al. Heterogeneity of severe asthma in childhood: confirmation by cluster analysis of children in the National Institutes of Health/National Heart, Lung, and Blood Institute Severe Asthma Research Program. *J Allergy Clin Immunol*. 2011;127:382-389.
59. Schatz M, Hsu JW, Zeiger RS, et al. Phenotypes determined by cluster analysis in severe or difficult-to-treat asthma. *J Allergy Clin Immunol*. 2014;133:1549-1556.
60. Woodruff PG, Boushey HA, Dolganov GM, et al. Genome-wide profiling identifies epithelial cell genes associated with asthma and with treatment response to corticosteroids. *Proc Natl Acad Sci U S A*. 2007;104:15858-15863.
- ** **This study began the concept of molecular phenotyping.**
61. Wenzel SE. Asthma: defining of the persistent adult phenotypes. *Lancet*. 2006;368:804-813.
62. Hallstrand TS, Moody MW, Wurfl MM, et al. Inflammatory basis of exercise-induced bronchoconstriction. *Am J Resp Crit Care Med*. 2005;172:679-8.
63. Parker JM, Oh CK, LaForce C, et al. Safety profile and clinical activity of multiple subcutaneous doses of MEDI-528, a humanized anti-interleukin-9 monoclonal antibody, in two randomized phase 2a studies in subjects with asthma. *BMC Pulm Med*. 2011;11:14.
64. Wenzel SE. Complex phenotypes in asthma: current definitions. *Pulm Pharm Ther*. 2013;26:710-715.
65. Moffatt MF, Gut IG, Demenais F, et al. A large-scale, consortium-based genome-wide association study of asthma. *N Engl J Med*. 2010;363:1211-1221.
66. Corren J, Lemanske RF, Hanania NA, et al. Lebrikizumab treatment in adults with asthma. *N Engl J Med*. 2011;365:1088-1098.
- ** **High serum periostin identified as predicting biomarkers of lebrikizumab treatment outcome.**
67. Piper E, Brightling C, Niven R, et al. A phase II placebo-controlled study of tralokinumab in moderate-to-severe asthma. *Eur Resp J*. 2013;41:330-338.
68. Humbert M, Berger W, Rapatz G, et al. Add-on omalizumab improves day-to-day symptoms in inadequately controlled severe persistent allergic asthma. *Allergy*. 2008;63:592-596.
69. Hanania NA, Alpan O, Hamilos DL, et al. Omalizumab in severe allergic asthma inadequately controlled with standard

- therapy: a randomized trial. *Ann Intern Med.* 2011;154:573–582.
70. Corren J, Busse W, Meltzer EO, et al. A randomized, controlled, phase 2 study of AMG 317, an IL-4/IL-13 antagonist, in patients with asthma. *Am J Respir Crit Care Med.* 2010;181:788–796.
 71. Slager RE, Hawkins GA, Ampleford EJ, et al. IL-4 receptor α polymorphisms are predictors of a pharmacogenetic response to a novel IL-4/IL-13 antagonist. *J Allergy Clin Immunol.* 2010;126:875–878.
 72. Slager RE, Otulana BA, Hawkins GA, et al. IL-4 receptor polymorphisms predict reduction in asthma exacerbations during response to an anti-IL-4 receptor α antagonist. *J Allergy Clin Immunol.* 2012;130:516–22. e4.
 73. Chu HW, Balzar S, Wesrcott JY, et al. Expression and activation of 15-lipoxygenase pathway in severe asthma: relationship to eosinophilic phenotype and collagen deposition. *Clin Exp Allergy.* 2002;32:1558–1565.
 74. Woolley KL, Gibson PG, Carty K, et al. Eosinophil apoptosis and the resolution of airway inflammation in asthma. *Am J Respir Crit Care Med.* 1996;154:237–243.
 75. Ten Brinke A, Zwinderman AH, Sterk PJ, et al. “Refractory” eosinophilic airway inflammation in severe asthma: effect of parenteral corticosteroids. *Am J Respir Crit Care Med.* 2004;170:601–605.
 76. Pavord ID, Korn S, Howarth P, et al. Mepolizumab for severe eosinophilic asthma (DREAM): a multicentre, double-blind, placebo-controlled trial. *Lancet.* 2012;380:651–659.
 77. Haldar P, Brightling CE, Hargadon B, et al. Mepolizumab and exacerbations of refractory eosinophilic asthma. *N Engl J Med.* 2009;360:973–984.
 78. Nair P. Anti-interleukin-5 monoclonal antibody to treat severe eosinophilic asthma. *N Engl J Med.* 2014;371:1249–1251.
 79. Castro M, Mathur S, Hargreave F, et al. Reslizumab for poorly controlled, eosinophilic asthma: a randomized, placebo-controlled study. *Am J Respir Crit Care Med.* 2011;184:1125–1132.
 80. Dahlén SE, Malmström K, Nizankowska E, et al. Improvement of aspirin-intolerant asthma by montelukast, a leukotriene antagonist: a randomized, double-blind, placebo-controlled trial. *Am J Respir Crit Care Med.* 2002;165:9–14.
 81. Wenzel S, Wilbraham D, Fuller R, et al. Effect of an interleukin-4 variant on late phase asthmatic response to allergen challenge in asthmatic patients: results of two phase 2a studies. *Lancet.* 2007;370:1422–1431.
 82. Wenzel S, Ford L, Pearlman D, et al. Dupilumab in persistent asthma with elevated eosinophils levels. *N Engl J Med.* 2013;368:2455–2466.
 - * **Effectiveness of anti-IL4R α on Th2 inflammation and pulmonary function.**
 83. Kim HY, DeKruyff RH, Umetsu DT. The many paths to asthma: phenotype shaped by innate and adaptive immunity. *Nat Immunol.* 2010;11:577–584.
 84. Black JL, Roth M. Intrinsic asthma: is it intrinsic to the smooth muscle?. *Clin Exp Allergy.* 2009;39:962–965.
 85. Holguin F, Bleecker ER, Busse WW, et al. Obesity and asthma: an association modified by age of asthma onset. *J Allergy Clin Immunol.* 2011;127:1486–93. e2.
 86. Lugogo NL, Kraft M, Dixon AE. Does obesity produce a distinct asthma phenotype?. *J Appl Physiol.* 2010;108:729–734.
 87. Calixto MC, Lintomen L, Schenka A, et al. Obesity enhances eosinophilic inflammation in a murine model of allergic asthma. *Br J Pharmacol.* 2010;159:617–625.
 88. Holguin F, Rojas M, Brown LA, et al. Airway and plasma leptin and adiponectin in lean and obese asthmatics and controls. *J Asthma.* 2011;48:217–223.
 89. Aydin M, Koca C, Ozol D, et al. Interaction of metabolic syndrome with asthma in post-menopausal women: role of adipokines. *Inflammation.* 2013;36:1232–1238.
 90. Dixon AE, Pradley RE, Forgione PM, et al. Effects of obesity and bariatric surgery on airway hyperresponsiveness, asthma control, and inflammation. *J Allergy Clin Immunol.* 2011;128:508–515. e2.
 91. Lombardi C, Gargioni S, Gardinazzi A, et al. Impact of bariatric surgery on pulmonary function and nitric oxide in asthmatic and non-asthmatic obese patients. *J Asthma.* 2011;48:553–557.
 92. Kato T, Takeda Y, Nakata T, et al. Inhibition by dexamethasone of human neutrophil apoptosis in vitro. *Nat Immun.* 1995;14:198–208.
 93. Baines KJ, Simpson JL, Wood LG, et al. Transcriptional phenotypes of asthma defined by gene expression profiling in induced sputum sample. *J Allergy Clin Immunol.* 2011;127:153–160.
 94. Wenzel SE, Barnes PJ, Bleecker ER, et al. A randomized, double-blind, placebo-controlled study of tumor factor- α blockade in severe persistent asthma. *Am J Respir Crit Care Med.* 2009;179:549–558.
 95. Baines KJ, Simpson JL, Wood LG, et al. Systemic upregulation of neutrophil α -defensins and serine proteases in neutrophilic asthma. *Thorax.* 2011;66:942–947.
 96. Doe C, Bafadhel M, Siddiqui S, et al. Expression of the T helper 17-associated cytokines IL-17A and IL-17F in asthma and COPD. *Chest.* 2010;138:1140–1147.
 97. Simpson JL, Powell H, Boyle MJ, et al. Clarithromycin targets neutrophilic airway inflammation in refractory asthma. *Am J Respir Crit Care Med.* 2008;177:148–155.
 98. Nair P, Gaga M, Zervas E, et al. Safety and efficacy of a CXCR2 antagonist in patients with severe asthma and sputum neutrophils: a randomized, placebo-controlled clinical trial. *Clin Exp Allergy.* 2012;42:1097–1103.
 99. Balzar S, Fajt ML, Comhair SA, et al. Mast cell phenotype, location, and activation in severe asthma. Data from the Severe Asthma Research Program. *Am J Respir Crit Care Med.* 2011;183:299–309.
 100. Bujarski S, Parulekar AD, Sharafkhaneh A, et al. The asthma COPD overlap syndrome (ACOS). *Curr Allergy Asthma Rep.* 2015;15:509.
 101. Elborn JS. The impact of personalised therapies on respiratory medicine. *Eur Respir Rev.* 2013;22:72–74.
 102. Chung KF, Adcock IM. Clinical phenotypes of asthma should link up with disease mechanisms. *Curr Opin Allergy Clin Immunol.* 2015;15:56–62.
 103. Fajt ML, Wenzel SE. Asthma phenotypes and the use of biologic medications in asthma and allergic disease: the next steps toward personalized care2. *J Allergy Clin Immunol.* 2015;135:299–310.
 104. Holgate ST. Stratified approaches to the treatment of asthma. *Br J Clin Pharmacol.* 2013;76:277–291.
 105. Agustí A, Antó JM, Auffray C, et al. Personalized respiratory medicine: exploring the horizon, addressing the issues. *Am J Respir Crit Care Med.* 2015;191:391–401. Summary of a BRN-AJRCCM workshop held in Barcelona on June 12, 2014.