

1 **Basic Cardiovascular Science Highlights of 2021/2022 – from novel discovery**
2 **tools and biomarkers to precision medicine.**
3
4

Paul C Evans^{^*}, Sean M. Davidson^{^*}, Johann Wojta^{^*}, Magnus Bäck^{^*}, Sveva Bollini, Mairi Brittan, Alberico L. Catapano, Bill Chaudhry, Matthijs Cluitmans, Massimiliano Gneccchi, Tomasz J Guzik, Imo Hofer, Rosalinda Madonna, João P Monteiro, Henning Morawietz, Elena Osto, Teresa Padró, Judith C. Sluimer, Carlo Gabriele Tocchetti, Kim Van der Heiden, Gemma Vilahur, Johannes Waltenberger, Christian Weber[^].

5
6 Here we review the highlights of cardiovascular basic science in published in 2021 and early 2022 on
7 behalf of the European Society of Cardiology Council for Basic Cardiovascular Science. We begin
8 with non-coding RNAs which have emerged as central regulators cardiovascular biology, and then
9 discuss how technological developments in single-cell 'omics are providing new insights in
10 cardiovascular development, inflammation and disease. We also review recent discoveries on the
11 biology of extracellular vesicles in driving either protective or pathogenic responses. The Nobel Prize
12 in Physiology or Medicine 2021 recognised the importance of the molecular basis of mechanosensing
13 and here we review breakthroughs in cardiovascular sensing of mechanical force. We also
14 summarise discoveries in the field of atherosclerosis including the role of clonal haematopoiesis of
15 indeterminate potential, and new mechanisms of cross-talk between hyperglycemia, lipid mediators
16 and inflammation. The past 12 months also witnessed major advances in the field of cardiac
17 arrhythmia including new mechanisms of fibrillation. We also focus on inducible pluripotent stem cell
18 (iPSC) technology which has demonstrated disease causality for several genetic polymorphisms in
19 long QT syndrome and aortic valve disease, paving the way for personalized medicine approaches.
20 Finally, the cardiovascular community has continued to better understand COVID-19 with significant
21 advancement in our knowledge of cardiovascular tropism, molecular markers, the mechanism of
22 vaccine-induced thrombotic complications and new anti-viral therapies that protect the cardiovascular
23 system.
24
25

1 **From novel discovery tools and biomarkers to precision medicine - basic**
2 **cardiovascular science highlights of 2021/2022**
3
4

Paul C Evans^{^*}, Sean M. Davidson^{^*}, Johann Wojta^{^*}, Magnus Bäck^{^*}, Sveva Bollini, Mairi Brittan, Alberico L. Catapano, Bill Chaudhry, Matthijs Cluitmans, Massimiliano Gnecci, Tomasz J Guzik, Imo Hofer, Rosalinda Madonna, João P Monteiro, Henning Morawietz, Elena Osto, Teresa Padró, Judith C. Sluimer, Carlo Gabriele Tocchetti, Kim Van der Heiden, Gemma Vilahur, Johannes Waltenberger, Christian Weber[^].

* equal contributions.

[^] Corresponding authors.

PCE: Department of Infection, Immunity and Cardiovascular Disease and Insigneo Institute, University of Sheffield, Sheffield, UK.

SMD: The Hatter Cardiovascular Institute, University College London, 67 Chenies Mews, WC1E 6HX, London, United Kingdom;

JWo: Department of Internal Medicine II, Medical University of Vienna, Vienna, Austria; Ludwig Boltzmann Institute for Cardiovascular Research, Vienna, Austria

MB; JPM: Queens Medical Research Institute, BHF Centre for Cardiovascular Sciences, University of Edinburgh, Scotland

SB: Department of Experimental Medicine (DIMES), University of Genova, L.go R. Benzi 10, 16132 Genova, Italy

MBä: Translational Cardiology, Karolinska Institutet and Karolinska University Hospital, Stockholm, Sweden, and University of Lorraine, INSERM U1116, Nancy University Hospital, Nancy, France

ALC: University of Milano and Multimedica IRCCS Milano Italy

BC: Biosciences Institute, Newcastle University, Newcastle upon Tyne, United Kingdom

MC: Cardiovascular Research Institute Maastricht, Maastricht University, Maastricht, Netherlands; Philips Research, Eindhoven, Netherlands

MG: Department of Molecular Medicine, Unit of Cardiology, University of Pavia;

Division of Cardiology, Unit of Translational Cardiology, Fondazione IRCCS Policlinico San Matteo, Pavia, Italy; Department of Medicine, University of Cape Town, South Africa.

TJG: Department of Internal Medicine, Jagiellonian University Medical College, Krakow, Poland and Institute of Cardiovascular and Medical Sciences, University of Glasgow, Glasgow, UK

IH: Central Diagnostic Laboratory, UMC Utrecht, the Netherlands

RM: Institute of Cardiology, Department of Surgical, Medical, Molecular and Critical Care Area, University of Pisa, Pisa, 56124 Italy; Department of Internal Medicine, Cardiology Division, University of Texas Medical School, Houston, TX, USA,

HM: Division of Vascular Endothelium and Microcirculation, Department of Medicine III, Faculty of Medicine and University Hospital Carl Gustav Carus, Technische Universität Dresden, Germany

EO: Institute of Clinical Chemistry and Department of Cardiology, Heart Center, University Hospital & University of Zurich, Switzerland;

TP; GV: Cardiovascular Program-ICCC, IR-Hospital Santa Creu i Sant Pau, IIB-Sant Pau, and CIBERCV-Instituto de Salud Carlos III, Barcelona, Spain.

JCS: Cardiovascular Research Institute Maastricht, Maastricht University Medical Center, Maastricht, Netherland, and University/BHF Centre for Cardiovascular Sciences, University of Edinburgh, Edinburgh, UK

CGT: Cardio-Oncology Unit, Department of Translational Medical Sciences, Center for Basic and Clinical Immunology (CISI), Interdepartmental Center of Clinical and Translational Sciences (CIRCET), Interdepartmental Hypertension Research Center (CIRIAPA), Federico II University, 80131 Napoli, Italy

KVDH: Biomedical Engineering, Thoraxcenter, Erasmus Medical Center, Rotterdam, The Netherlands

JWa: Cardiovascular Medicine, Medical Faculty, University of Muenster, Muenster, Germany; Diagnostic and Therapeutic Heart Center, Zurich, Switzerland

CW: Institute for Cardiovascular Prevention (IPEK), Ludwig-Maximilian-Universität (LMU) München, German Center for Cardiovascular Research (DZHK), partner site Munich Heart Alliance and Munich Cluster for Systems Neurology (SyNergy), Munich, Germany; Department of Biochemistry, Cardiovascular Research Institute Maastricht (CARIM), Maastricht University, Maastricht, the Netherlands.

1 **ABSTRACT**

2 Here we review the highlights of cardiovascular basic science in published in 2021 and early 2022 on
3 behalf of the European Society of Cardiology Council for Basic Cardiovascular Science. We begin
4 with non-coding RNAs which have emerged as central regulators cardiovascular biology, and then
5 discuss how technological developments in single-cell 'omics are providing new insights in
6 cardiovascular development, inflammation and disease. We also review recent discoveries on the
7 biology of extracellular vesicles in driving either protective or pathogenic responses. The Nobel Prize
8 in Physiology or Medicine 2021 recognised the importance of the molecular basis of mechanosensing
9 and here we review breakthroughs in cardiovascular sensing of mechanical force. We also
10 summarise discoveries in the field of atherosclerosis including the role of clonal haematopoiesis of
11 indeterminate potential, and new mechanisms of cross-talk between hyperglycemia, lipid mediators
12 and inflammation. The past 12 months also witnessed major advances in the field of cardiac
13 arrhythmia including new mechanisms of fibrillation. We also focus on inducible pluripotent stem cell
14 (iPSC) technology which has demonstrated disease causality for several genetic polymorphisms in
15 long QT syndrome and aortic valve disease, paving the way for personalized medicine approaches.
16 Finally, the cardiovascular community has continued to better understand COVID-19 with significant
17 advancement in our knowledge of cardiovascular tropism, molecular markers, the mechanism of
18 vaccine-induced thrombotic complications and new anti-viral therapies that protect the cardiovascular
19 system.

20
21
22
23

1 1. INTRODUCTION

2 The aim of this review from the European Society of Cardiology (ESC) Council for Basic
3 Cardiovascular Science is to highlight the most noteworthy developments over the past year, in the
4 field of cardiovascular basic science. The cited reports were selected as representative examples of
5 studies which provided robust evidence for particularly novel insights. *Cardiovascular Research*
6 previously reviewed the highlights of 2020 divided into vascular and cardiac topics,^{1 2} but here we
7 integrate both areas to generate the Basic Cardiovascular Science Highlights of 2021/2022.

9 2. CARDIOVASCULAR RNA UNIVERSE

10 2.1 Non-coding RNAs (nc RNAs)

11 In addition to the role of messenger RNA (mRNAs) in the 'central dogma' of molecular biology as a
12 template for protein synthesis, the RNA universe also contains multiple constellations of microRNAs
13 (miRNAs; miRs), long non-coding RNAs (lncRNAs) and circular RNAs (circRNAs) that control
14 fundamental processes of life. These RNA species adopt complex structures and interact with
15 nucleotides, proteins and lipids to control multiple functions including chromatin structure,
16 transcription, RNA splicing and stability, intracellular signalling and organelle dynamics. Research
17 reported in 2021 has provided further insight into the role of miRNAs, lncRNAs, and circRNAs in the
18 regulation of vascular remodeling and cardiac disease. Using both single-cell (sc) and bulk RNA-
19 sequencing to investigate transcriptional changes associated with endothelial-to-mesenchymal
20 transition (EndMT), *Monteiro et al* identified for the first time the genomic locus hosting the lncRNA
21 MIR503HG as necessary to maintain endothelial cell (EC) identity and function³. In a series of our
22 loss- and gain-of-function experiments the group demonstrated that loss of lncRNA is a causal event
23 in EndMT observed in pulmonary arterial hypertension (PAH) in association with vascular remodelling
24 (Figure 1). Further, located upstream from the vascular smooth muscle cell (vSMC)-associated miR-
25 143 and -145 cluster, the lncRNA CARMN (Cardiac Mesoderm Enhancer-associated Noncoding
26 RNA) was recently identified as key regulator of vSMC function and the pathophysiology of
27 atherosclerotic disease⁴. Crucially, while crosstalk between lncRNA host genes and coupled miRNAs
28 is often seen, CARMN was found to function independently from miR-143/-145 in regulating vSMC
29 and activating a pro-atherogenic proliferative state (Figure 1).

30
31 *Gong et al* identified in atherosclerotic mouse models a novel circRNA, circEsys2, involved in vascular
32 remodeling through the targeted inhibition of alternative mRNA splicing. By performing loss- and gain-
33 of-function mutation analyses in vascular smooth muscle cells, circEsys2 was shown to enhance cell
34 proliferation and migration and blunt apoptosis and differentiation. Furthermore, silencing of circEsys2

1 prevented neointima formation while circEsys2 overexpression enhanced neointimal hyperplasia in an
2 *in vivo* model of carotid artery injury.⁵ The role of miRNAs in atherosclerosis progression was
3 examined by Liu *et al* by describing the role of the Nuclear Factor of Activated T-cell isoform c3
4 (NFATc3)/miR-204 axis in the regulation of foam cell formation in atherosclerosis. Using genetically
5 modified mice, they showed that NFATc3 prevents macrophage foam cell formation and limits the
6 expression of scavenger receptors SR-A and CD36 by inducing expression of the microRNA miR-
7 204,⁶ suggesting the NFATc3/miR-204 axis as a potential therapeutic target to reduce plaque
8 formation. In a separate study involving macrophages, *Schober et al* illuminated the circadian
9 patterns of myocardial infarction (MI) by evaluating macrophage-related miRNAs. They evidence, in a
10 murine model of atherosclerosis, that macrophage miR-21 drives circadian regulation of macrophage
11 apoptosis by targeting proapoptotic Xaf1 (XIAP-associated factor 1), thereby regulating plaque
12 composition and susceptibility to rupture.⁷ Further studies in a murine model of pressure-overload
13 heart failure have also found a key role for macrophage miR-21 in modulating cardiac fibrosis by
14 regulating macrophage polarization towards a pro-inflammatory (M1) phenotype.⁸ In addition, *Hinkel*
15 *et al* identified a pivotal role of miR-132 in the mediation of pathologic cardiac hypertrophy in a novel
16 porcine model of percutaneous aortic constriction by stent implantation.⁹

17
18 ncRNAs have also continued to attract attention as biomarkers with prognostic and diagnostic
19 potential. A landmark study from *Blanco-Dominguez et al.* identified a novel miRNA with potential
20 diagnostic value in acute myocarditis. The authors performed miRNA microarray analyses in sorted
21 CD4+ T cells and type 17 helper T (Th17) cells after inducing experimental autoimmune myocarditis
22 or MI in mice and identified mmu-miR-72 as a differentially expressed miRNA. They further identified
23 the human homologue hsa-miR-Chr8:96 and demonstrated its potential to distinguish patients with
24 myocarditis from those with MI and healthy controls.¹⁰ Thus, miR-Chr8:96 has translational potential
25 as a novel biomarker to diagnose myocarditis. miR-133a is a well-established, diagnostic circulating
26 biomarker in patients with heart failure.¹¹ *Escate et al.* expanded on the diagnostic potential of this
27 miRNA by demonstrating that elevated plasma levels of miR-133a predict the future occurrence of
28 major adverse cardiovascular events (MACE) in patients with familial hypercholesterolaemia (FH).¹²
29 This observation supports the potential utility of miR-133a in improving risk stratification and
30 prognosis in high-risk patients. More broadly, an international consortium supporting collaboration
31 and research on ncRNAs in cardiovascular disease (CardioRNA Cost Action CA17129) published a
32 Position Paper on the pathophysiologic role of ncRNAs, and to provide recommendations to translate
33 this into clinical practice.¹³

1 Other studies have progressed ncRNA candidates with therapeutic potential towards clinical
2 translation.^{9, 14, 15} *Kay et al* examined the potential of targeting ncRNAs to promote cell-based
3 regenerative strategies for heart disease. Using an integrated approach, they identified CARMA
4 (CARdiomyocyte Maturation-Associated lncRNA), a conserved lncRNA controlling cardiomyocyte
5 differentiation and maturation in human embryonic stem cells. CARMA knockdown promoted
6 cardiogenic commitment and cardiomyocyte differentiation in embryonic stem cells, and is therefore a
7 novel target for improving human ESC-derived cardiomyocyte production in regenerative
8 cardiovascular medicine.¹⁴ On the other hand, *Modica et al* provided evidence for the effectiveness of
9 a novel nanotechnology-based approach for delivering exogenous synthetic miR-133a. The authors
10 demonstrated that intra-tracheal nebulization of miR-133a-nanoconstruct once-a-day on alternate
11 days for 4 consecutive weeks protects against heart failure progression (improved cardiac function
12 parameters and lower fibrosis) in a murine model. This improvement was associated with the
13 restoration of physiological levels of miR-133a in cardiomyocytes without significant accumulation in
14 other myocardial cells or organs.¹⁵

15 16 **2.2 Single cell approaches**

17 Single-cell RNA sequencing (scRNAseq) has emerged as a powerful tool to dissect transcriptional
18 profiles of the complex cardiovascular system at single-cell resolution. scRNAseq has been insightful
19 in our understanding of the earliest stages of cardiac development by identifying the epicardial
20 progenitor field, which is anatomically and transcriptionally distinct from the currently known first and
21 second heart fields.¹⁶ In the formed heart, scRNAseq and spatial transcriptomics were used to show
22 that dysregulation of TBX5, the mutated gene causing septal and conduction defects in patients with
23 Holt-Oram syndrome, leads to transcriptional consequences in specific cardiomyocyte subtypes.¹⁷
24 The study went on to show using cell-based analyses and mice that the stability of many gene
25 regulatory networks, including those that have been shown to be relevant to congenital heart disease,
26 are sensitive to TBX5 dosage.

27
28 At the level of the vasculature, the number of publications of atlas-type human or primate scRNAseq,
29 or Assay for Transposase-Accessible Chromatin (ATAC) datasets has steadily increased, which
30 provides a valuable, yet often descriptive resource.¹⁸⁻²¹ scRNA-seq has been used to identify
31 transcriptional changes upon conditional cell type-specific genetic deletion, otherwise obscured in
32 bulk tissue RNA sequencing.²² As for immune cells in atherosclerosis, the detection of different
33 subsets has culminated in a consensus on cell type markers,²³ yet to be achieved for the many
34 varieties of vSMCs identified using scRNAseq in recent years, i.e. fibromyocytes, proinflammatory or

1 modified vSMCs, SMC-derived intermediate cells.^{21, 24-26} scRNAseq has also progressed our
2 understanding of EC,^{27 28} with *Rodor et al* identifying CD74 as potential target in PAH and showing its
3 capacity to regulate barrier integrity.²⁸

4
5 At a cardiac level, the implementation of scRNAseq allowed the impact of heart failure on circulating
6 immune cells to be determined.²⁹ Furthermore, it demonstrated an exacerbated inflamed
7 transcriptome in circulating monocytes and a signature of T-cell activation in heart failure patients
8 harbouring clonal haematopoiesis-driver mutations in DNA methyltransferase DNMT3A, thereby
9 providing further insights into the potential effect of DNMT3A mutations in heart failure progression.³⁰
10 On the other hand, *Hesse et al.* have defined a high level of heterogeneity of epicardial stromal cells
11 following MI, similar to cardiac fibroblast heterogeneity, with evidence of regenerative capacity and
12 hypoxic signalling.³¹ *Tombor et al* used scRNAseq of endothelial-lineage traced mice to change the
13 dogma on EndMT in MI, showing this is a transient affair, often without a definite mesenchymal
14 endstage.³²

15
16 Moving forward, cardiovascular scientists will benefit greatly from the generation of multi-omics
17 reference atlases, including different layers of information on RNA, protein, spatial anatomy,
18 interactome and cell ontology.³³⁻³⁵ Overall, scientific progress can be expedited by open-access
19 science and data sharing. Thus, the integration of available datasets for mesenchymal cells,³⁶ as
20 previously carried out for immune cells in atherosclerosis,³⁷ and a web-based application by the *Miller*
21 lab (plaqview.com),³⁸ pave the way for new, meaningful discoveries in cardiovascular biology.

22 23 **3. CARDIOVASCULAR DEVELOPMENT**

24 2021 witnessed progress in several important aspects of heart development with implications for our
25 understanding of both congenital and acquired heart conditions. Genomic studies of congenital heart
26 malformations now allow the analysis of variants within the context of gene networks. A good
27 example of this is the recent genomic study on hypoplastic left heart syndrome (HLHS),³⁹ where
28 whole-exome sequencing, coupled to nuclear transcriptomics and scRNAseq identified genetic
29 heterogeneity in HLHS that converges to alter fundamental processes (e.g. autophagy, apoptosis,
30 proliferation) in myogenesis.

31
32 Despite the relative ease in differentiating functional, if immature, cardiomyocytes from iPSC, it has
33 proven remarkably difficult to create organoids resembling the cellular and structural complexity of the
34 vertebrate heart *in vitro*. However, *Lewis-Israeli et al*⁴⁰ described a robust protocol for producing

1 cardiac organoids from iPSC using a three-step Wnt signaling modulation strategy. These organoids
2 develop a broad range of cardiac cell types, including those that are induced through interactions
3 between distinct primary cardiac cell types, and develop cavities that superficially resemble the lumen
4 of the chambers. Moreover, they are vascularised and display regular beating. Importantly, the
5 transcriptome of the organoids more closely resembles foetal hearts than monolayer cardiomyocytes.
6 This method is an important step on the path to developing a robust human-based *in vitro* model of
7 the heart.

8
9 It is increasingly apparent that the majority of valve malformations and dysfunction arise from
10 abnormal development, and yet the mechanisms of valve development are incompletely understood.
11 The study by *Fukui* et al focussed on the role of mechanical factors using zebrafish embryos.⁴¹ They
12 identified a critical role for shear stress by showing that ectopic activation of wall shear stress, using
13 agarose beads implanted into the atrium of the early zebrafish heart, resulted in the formation of
14 valve-like structures that expressed the characteristic molecular signature of primitive valves,
15 including the activation of NFATc and *klf2a*. Downstream of this, they ruled out a number of well-
16 known mechanosensitive pathways, and instead identified adenosine tri-phosphate (ATP) signalling
17 as a mediator of Ca^{2+} oscillations that were essential for specifying valve cell identity. Overall, the
18 convergence of large-scale genomic network analyses, scRNAseq and spatial transcriptomics and
19 experimental developmental biology is coming close to explaining the mechanisms underlying heart
20 malformations presenting at birth and in adulthood.

21 22 **4. VASCULAR DISEASE AND REPAIR**

23 **4.1 Mechanosensing**

24 The Nobel Prize in Physiology or Medicine 2021 was awarded to *David Julius* from the University of
25 California San Francisco and *Ardem Patapoutian* from The Scripps Research Institute La Jolla for
26 explaining the molecular basis for sensing heat, cold and mechanical force.⁴² *Ardem Patapoutian*
27 identified PIEZO 1 and 2 as ion channels activated by mechanical force,⁴³ and they are central
28 responders of arterial responses to flow.⁴⁴ Recently, the protein kinase N2 (PKN2) has been shown to
29 be activated by flow through the mechanosensitive ion channel PIEZO1 and mediate flow-induced
30 endothelial NO synthase activation and vascular tone regulation⁴⁵ (Figure 2). As another important
31 mechanosensor, the glycocalyx modulates the endothelial redox state in response to shear stress
32 and could mediate an atheroprotective synergism between glycocalyx sialic acids and nuclear factor
33 erythroid 2-related factor (NRF2) antioxidant signaling.⁴⁶ The regulation of NRF2 plays also a major
34 role in the reduced endothelial cell viability and wound healing in response to cigarette smoke

1 extracts under atherogenic low flow conditions.⁴⁷ The concept of disturbed flow as an initial stimulus
2 for the development of atherosclerotic plaques has led to exciting new therapies to target
3 mechanosensitive genes like *TWIST1*, *GATA4*, and bone morphogenic proteins (*BMPs*) using siRNA-
4 based technologies in an attempt to slow down the progression of atherosclerosis.^{48, 49}

6 **4.2. Atherosclerosis risk factors**

7 The metabolic syndrome – in concert with inflammation - plays a central role in atherosclerosis. In
8 particular, the causal role low-density lipoprotein (LDL) in atherosclerosis is indisputably supported by
9 multiple lines of evidence such as epidemiological studies, Mendelian randomization and genetic
10 analyses, as well as randomized clinical trials and animal model experimentation.

11
12 Traditional lipid-lowering drugs such as statins aim to reduce lipid uptake and/or cholesterol synthesis
13 and are still widely used. However, the availability of genetic data and the identification of the genetic
14 cause for rare diseases linked to dyslipidaemias has prompted spectacular advances in the
15 identification of pharmacological targets for the treatment of dyslipidaemias (Figure 3). The most
16 recent advances in lipid-lowering relate to the inhibition of proprotein convertase subtilisin kexin 9
17 (PCSK9), angiotensin-like 3 (ANGPTL3) and lipoprotein (a) (Lp(a)). Besides monoclonal antibodies,
18 additional options to inhibit PCSK9 are emerging, including gene silencing with an siRNA or gene
19 editing employing the CRISPR/Cas system. Inclisiran, a siRNA conjugated with *N*-
20 acetylgalactosamine residues ensuring hepatic selectivity, decreases PCSK9 production by
21 promoting the degradation of its mRNA. This approach allows for twice-yearly dosing, with long-term
22 lowering of LDL-C (~50%), potentially enhancing patient compliance compared with other cholesterol-
23 lowering drugs.^{50, 51} Along the same line of RNA interference, Lp(a)-reducing drugs are being
24 investigated in phase 2-3 trials.⁵² At earlier stages of development are gene-editing technologies,
25 which introduce permanent genomic changes to alter gene function. A single treatment with *PCSK9*
26 gene or base editors has been shown to confer durable LDL-C reduction in primates⁵³. Evinacumab is
27 a monoclonal antibody targeting ANGPTL3. It reduces significantly triglycerides (TG) by up to 80% in
28 hypertriglyceridaemic subjects⁵⁴ and it is highly effective in reducing LDL-C levels in patients with
29 homozygous FH carrying null *LDLR* mutations⁵⁵ providing a new pharmacological tool. In a recent
30 study, membrane type 1 matrix metalloproteinase (MT1-MMP), in addition to activating MMP-2, was
31 shown to regulate LDL-receptor (LDLR) shedding, affecting circulating lipid concentrations and
32 atherosclerosis.⁵⁶

1 The past year has further blurred the borders between traditional risk factors and the role of
2 inflammation in atherosclerosis as their connections and interplay become more evident. Diabetes
3 mellitus elevates cardiovascular risk, and hyperglycaemia contributes strongly to metabolic
4 syndrome. Besides these known effects, *Edgar et al* elucidated a pro-inflammatory and pro-
5 atherogenic switch in macrophages from diabetic mice persisting even when cultured under
6 normoglycaemic conditions.⁵⁸ This persevering effect of earlier hyperglycaemia may explain the
7 relatively low degree of risk reduction upon glucose level normalisation in diabetics. The inseparable
8 connection between cholesterol and inflammation and atherosclerosis is further supported by a recent
9 study that showed how sensing of cholesterol crystals by macrophages induces complement
10 component C5aR1 signaling on mitochondrial membranes and results in interleukin (IL)-1 β production
11 and sterile inflammation.⁵⁹ Hence, intracellular C5aR1 targeting may be used to normalize
12 mitochondrial function and reduce IL-1 β release. This has translational relevance since inhibition of
13 IL-1 β production through targeting the inflammasome has been identified as a target in cardiovascular
14 disease previously. Another old acquaintance in cardiovascular disease therapy, rivaroxaban, a direct
15 oral anticoagulant, not only targets factor Xa activity, but may also reduce inflammasome formation.
16 In mice treated with rivaroxaban, macrophage autophagocytic activity increased significantly, which
17 the authors were able to trace back to the Xa-PAR2 axis.⁵⁷

18
19 Recent studies show the complex intertwinement between traditional risk factors, vascular biology
20 and immunology. Cardiovascular risk factors can affect haematopoiesis through defective
21 angiogenesis in the bone marrow towards generation of inflammatory leukocytes, thereby creating a
22 self-energizing circle of cardiovascular risk factors – defective angiogenesis – release of inflammatory
23 cells – cardiovascular disease exacerbation.⁶⁰ Sakic et al emphasised crosstalk between vSMCs and
24 vascular inflammation by demonstrating that S100A4 induces vSMC change towards a
25 proinflammatory phenotype to drive features of plaque instability⁶¹. Together, these studies call for an
26 integrated and unprejudiced approach in atherosclerosis research to link traditional risk factors with
27 novel molecular mechanisms.

28 29 **4.3 Inflammation in Atherosclerosis**

30 The immune response is critical throughout the development of atherosclerotic lesions, during
31 disease initiation, as a trigger for episodic plaque progression, and a contributor to thrombotic
32 complications.⁶² A failure in the resolution of inflammation can prevent healing and repair of the
33 vascular wall.⁶²⁻⁶⁴ This concept was advanced by *Arnardottir et al* who found that lipid-specialized,
34 pro-resolving mediators (SPM) signalling through G-protein coupled receptor (GPR)-32,

1 Is critical for inflammatory resolution and atheroprotection.⁶⁴

2
3 The proposal that macrophage uptake mechanisms are decisive for the turning point that leads either
4 to inflammation resolution or to chronic inflammation and plaque progression has received further
5 support from analysis pro-resolving pathways⁶⁴ or phagocytic immune checkpoints in murine
6 models.⁶⁵ Focussing on the CD47- signal-regulatory protein (SIRP) α immune checkpoint, loss of
7 SIRP α in macrophages stimulated efferocytosis, attenuated oxidized LDL-induced inflammation and
8 induced an M2 macrophage phenotype.⁶⁵ These findings may pave the way for novel interventions to
9 promote inflammatory resolution through macrophage uptake mechanisms and phenotypic transitions
10 to protect the vasculature.

11
12 Adaptive immune responses are critical regulators of atherosclerosis. On a systemic level, pro-
13 inflammatory and cytotoxic T-lymphocytes prevail in atherosclerosis, as demonstrated by a
14 preferential expansion and function of CD28^{null} T lymphocytes after *ex vivo* IL-7 and IL-15 stimulation
15 of high-purity sorted CD4⁺ cells isolated from patients with acute coronary syndrome.⁶⁶ The local
16 recruitment of regulatory T lymphocytes (T_{reg}) is critical for the control of atherosclerotic lesion
17 inflammation and is, in part, regulated by cellular metabolism.⁶⁷ As an approach to use T_{reg}
18 recruitment as a therapeutic strategy to selectively target adaptive immune regulation in the
19 atherosclerotic plaque, adoptive transfer of the fractalkine receptor CX3CR1 overexpressing T_{reg} was
20 shown to increase their recruitment to atherosclerotic lesions and decreased atherosclerosis
21 burden.⁶⁸

22
23 However, inhibition of some immune checkpoints can lead to enhanced atherosclerosis. This is
24 exemplified by *Poels et al.* who found that short-term immune checkpoint inhibitors (ICIs) therapy
25 aggravates T cell-mediated plaque inflammation and drives plaque progression in mice.⁶⁹ Also, ICIs
26 used to treat cancer, such as monoclonal antibodies targeting CTLA-4, PD-1, and PD-L1, have been
27 associated with adverse cardiovascular events.⁷⁰ For example, *Michel et al.* discovered that anti-PD1
28 therapy in a mouse model of melanoma led to impaired left ventricular function and promoted
29 myocardial infiltration with CD4⁺ and CD8⁺ T cells via a TNF-dependent mechanism.^{71, 72} Therefore,
30 the use of ICIs in the treatment of cancer provides exciting new opportunities for therapies but should
31 be pursued with caution.

4.4 Haematopoiesis of Indeterminate Potential

Clonal haematopoiesis of indeterminate potential (CHIP) has recently emerged as an exciting topic in cardiovascular medicine and biology. CHIP is defined as positive selection of specific somatic mutations in haematopoietic stem cells that provide a proliferative advantage and finally result in a clonal population carrying the mutation. Besides being associated with a 0.5 to 1% risk per year to develop leukaemia, CHIP is also associated with ageing, smoking, obesity and type 2 diabetes mellitus, chronic inflammation, infections, sleep deprivation, stress, hyperlipidaemia and atherosclerosis. Most mutations identified in CHIP affect the epigenetic regulators DNA (cytosine-5)-methyltransferase 3A (*DNMT3A*), tet methylcytosine dioxygenase 2 (*TET2*) and ASXL transcriptional regulator 1 (*ASXL1*) and the tyrosine kinase janus kinase 2 (*JAK2*) which result in a pro-inflammatory state that offers a possible explanation for the association of CHIP with a two-fold increase in risk to develop cardiovascular disease.^{73, 74} Using mice that express the *JAK2*^{V617F} variant exclusively in macrophages, *Fidler et al* reported increased proliferation of macrophages in atherosclerotic lesions and greater necrotic cores. These effects were ameliorated when caspase 1 and 11, which are key components of the inflammasome or gasdermin D, which plays a major role in pyroptosis, were deleted. The authors also noted increased lesional expression of absent in melanoma 2 (*AIM2*) and found that atherosclerosis was reduced in mice deficient in *Aim2*. The authors concluded that enhanced proliferative stress caused by *JAK2*^{V617F} leads to DNA damage and to activation of the *AIM2* inflammasome resulting in IL-1 β activation, which then in turn starts a feed forward loop resulting in even more macrophage proliferation thereby aggravating atherosclerosis.⁷⁵

A new perspective to the field added *Heyde et al* who recently showed by mathematical modeling and murine models that increased proliferation of haematopoietic stem cells occurs in individuals suffering from atherosclerosis thereby increasing the risk to develop clonal haematopoiesis by the age of 70 3.5-fold. Based on their findings the authors propose a vicious cycle in which atherosclerosis leads to clonal haematopoiesis, which in turn aggravates atherosclerosis.⁷⁶

5. CARDIAC DISEASE AND REPAIR

5.1 Extracellular vesicles and nanoparticles

2021 was another exciting year in the field of extracellular vesicle (EV) biology for regenerative medicine, including cardiac repair and regeneration (Figure 4). There was increasing interest in understanding the mechanism of EV-based intercellular communication within the myocardium during ventricular remodeling after acute MI. In terms of the role of EVs in cardiac fibrosis after MI, however, findings differ. For example, *Li et al* showed that miR-30d is mainly secreted in EVs by

1 cardiomyocytes and inhibits fibroblast proliferation by acting on integrin $\alpha 5$ via paracrine signaling.⁷⁷
2 Counterbalancing this view, *Wang et al* evidenced, in a mouse model of MI, that EVs released by
3 myocardial M2 macrophages exacerbate migration, proliferation and myofibroblastic transformation of
4 cardiofibroblasts.⁷⁸ By performing mechanistic studies in cocultured primary cardiofibroblasts and M2
5 macrophages, the authors linked these effects to activation of miR-138-5p/RhoC signaling after
6 delivery of the M2 macrophage-derived EVs containing circular RNA *circUbe3a* into the
7 cardiofibroblasts.⁷⁸ These findings may offer an additional therapeutic target to optimize the
8 endogenous mechanism of cardiac repair but suggest that EV function may depend on cell of origin.

9
10 There is great interest in the potential for EVs prepared from stem or progenitor cells to enhance
11 cardiac repair. Increasing evidence suggests the mechanism may involve the resolution of
12 inflammation. For example, *Correa et al* reported that EVs secreted from human iPSC-derived
13 cardiovascular progenitor cells (CPC) can trigger a pro-resolving immune response in preclinical
14 murine models of either chronic or acute heart failure. Similar results were confirmed *in vitro* on
15 human inflammatory cells, suggesting that this EV formulation can instruct the immune cell response
16 towards a pro-resolving phenotype.⁷⁹ *Patil et al* showed a similar pro-resolving effect of mesenchymal
17 stem cell (MSC)-derived small EVs, which they attributed to the EVs both enhancing opsonisation of
18 dead cells and activating phagocytic signaling, thereby augmenting removal of apoptotic cells,
19 resolution of inflammation, and improving cardiac recovery after injury.⁸⁰

20
21 In order to investigate a clinically feasible translational approach, *Katsur et al* assessed whether
22 cardioprotection could be achieved using a reproducible, clinical-grade preparation of small EVs
23 obtained from the CTX0E03 human neural stem cell line. Systemic administration of small EVs from
24 differentiating CTX0E03 reduced infarct size in mice and prevented *in vitro* cardiomyocyte
25 mitochondrial permeability transition pore opening, which is responsible for cardiomyocyte death
26 during reperfusion injury. These findings provide evidence for considering non-cardiovascular, yet
27 stabilised, cell lines as additional candidate source of therapeutic EVs.⁸¹ Interestingly, however, EVs
28 from proliferating CTX0E03 cells were not cardioprotective, which suggests that the status of cells of
29 origin can impact their secreted EV activity.⁸¹ Further evidence of this is provided by a study showing
30 that systemic administration of serum small EVs from young rats into aged ischaemic rats improved
31 functional outcomes after ischemic stroke, in contrast to small EVs from aged rats that worsened
32 outcome.⁸² This provides further evidence that EV function is altered in disease, and further suggests
33 that EV-miR-mediated vascular intercellular communication is altered in patients with chronic kidney
34 disease and coronary artery disease.

1
2 A major goal in cardiac regenerative medicine is to identify novel methods to reinstate cardiomyocyte
3 renewal. In such a scenario, EVs released from cardiac progenitors have been widely investigated,
4 given the role of cardiac stromal cells such as the epicardium-derived progenitor cells play in cardiac
5 muscle growth during embryonic development, and in heart regeneration in zebrafish and in neonatal
6 mice. *Villa del Campo et al* reported that epicardial EVs isolated from the secretome of both mouse
7 and human progenitors enhanced the proliferative activity of neonatal murine cardiomyocytes *in vitro*
8 and promoted cell cycle re-entry when injected into the injured area of infarcted neonatal hearts.
9 These EVs also enhanced regeneration in cryoinjured engineered human myocardium constructs, as
10 a novel model of human myocardial injury. Notably, the epicardial EV cargo was found enriched with
11 specific miRNAs, including miR-30a, miR-100, miR-27a, and miR-30e, which recapitulated the EV
12 regenerative influence on human stem cell-derived cardiomyocytes and cryoinjured cardiac
13 constructs *in vitro*.⁸³
14

15 The relevance of the content of cardiovascular cell-derived EVs was highlighted by publications
16 showing that miRNAs of the miR-106a-363 cluster,⁸⁴ periostin⁸⁵ and mitochondrial cargoes⁸⁶ can act
17 as effectors of cardiac repair. While such encouraging evidence supports the exploitation of
18 stem/progenitor cell-EVs as candidate therapeutics to promote adult cardiomyocyte proliferation, a
19 general consensus has not been reached yet on their mechanism of action. In fact, *Lima Correa et al*
20 recently showed that EVs obtained from human iPSC-derived cardiac progenitor cells failed to trigger
21 the generation of new cardiomyocytes in chronically infarcted hearts in mouse models. Despite this
22 negative result, the authors confirmed that EVs from cardiac progenitor cells remained capable of
23 significantly improving cardiac function by non-regenerative mechanisms.⁸⁷
24

25 These findings suggest that further analyses and accurate lineage tracing are required to better
26 understand the regenerative potential of cardiac EVs. At present, the rapid clearance of EVs from
27 circulation is a limitation for their clinical application. During 2021, a number of studies aimed to
28 overcome this barrier by constructing specific nanoparticles and genetically modifying cells to improve
29 retention time of the cell-derived EVs. Thus, *Wei et al* demonstrated that intravenously-injected EV
30 derived from modified mouse bone marrow MSC overexpressing CD47, a transmembrane protein
31 known to elicit blockade of the mononuclear cell phagocytosis, have prolonged retention in the
32 circulation and accumulate at greater levels in the ischemic heart.⁸⁸
33
34

5.2 Cardiotoxicity and regeneration

A wide range of drugs, including but not limited to antineoplastic chemotherapeutic agents, can cause heart electrophysiology dysfunction, muscle damage and other cardiovascular pathologies. For example, anthracyclines such as doxorubicin (DOX) are a cornerstone for the treatment of many cancers, but their use is complicated by cardiotoxicity, especially left ventricular dysfunction.

An interesting 2021 paper reported that transcutaneous vagal nerve stimulation prevented DOX-induced cardiotoxicity in rats by rebalancing autonomic tone, ameliorating cardiac dysfunction and remodelling. It was hypothesized that the mechanism involved crosstalk between autonomic neuromodulation, innate immune cells such as macrophages and chemokines.⁸⁹ Indeed, there are multiple mechanisms responsible for anthracycline cardiotoxicity.^{70, 90, 91} *Chan et al.* found that two orally available MMP inhibitors ameliorated DOX cardiotoxicity by attenuating intracellular and extracellular matrix remodelling, suggesting that they may be a potential prophylactic strategy to prevent heart injury during chemotherapy.⁹⁰ Remote ischaemic preconditioning can ameliorate DOX-induced cardiotoxicity by preserving mitochondrial integrity⁹² and this is currently the subject of the RESILIENCE clinical trial.⁹³

Other recent studies (discussed in⁹⁴) have identified harmful effects of anticancer therapies on the ability of stem/progenitor cells to repair cardiac damage, through a reduction of stem cell viability and paracrine activity. Thus numerous animal and clinical studies have demonstrated that local or systemic administration of mesenchymal stem cells significantly improve cardiac function, through a reduction in inflammatory responses and myocardial fibrosis.⁹⁵ Antivirals can also induce cardiotoxicity, including the only FDA-approved treatment for hospitalized COVID-19 patients, remdesivir which can induce toxicity in human iPSC-derived cardiomyocytes through mitochondrial fragmentation, electrophysiological alterations and sarcomere disarray.⁹⁶

5.3 Cardiac arrhythmias

Several key insights into fibrillation and re-entrant arrhythmias were obtained in 2021 (Figure 5). *Handa et al* revealed that the degree of gap junction coupling as well as the pattern of fibrosis influences mechanisms sustaining ventricular fibrillation.⁹⁷ Differentiating between these underlying mechanisms of maintenance of fibrillation may help to guide therapy. Re-entrant arrhythmias may also initiate in the absence of structural abnormalities, shown recently in a study on the spatiotemporal interaction between trigger and electrical substrate in the context of unexplained sudden cardiac arrest (SCA).⁹⁸ Analysis of explanted hearts and observations in survivors of

1 unexplained SCA, identified key elements required for re-entry initiation including the occurrence of
2 an early premature beat from an early repolarizing region of the ventricles, which may block against a
3 steep repolarization time (RT) gradient to start re-entry. They also showed that detection of the origin
4 of premature beats and their relation to RT gradients in patients is possible with non-invasive
5 electrocardiographic imaging (ECGI) and may provide targets for therapy. ECGI was also employed
6 by *Leong et al* in survivors of SCA to show that not only repolarization abnormalities, but also
7 underlying conduction abnormalities play a role in the initiation of SCA.⁹⁹ A similar mechanistic
8 reasoning extends to atrial arrhythmias.¹⁰⁰ Bringing these studies together highlights that any cause
9 of steep excitability dispersion – whether resulting from local changes in gap junction coupling,
10 fibrosis, local conduction slowing, or inherent repolarization duration heterogeneity – play a critical
11 role in the initiation and maintenance of re-entry and fibrillation.
12

13 New tools are essential to obtain mechanistic insights and recent reports highlight how the field of
14 atrial fibrillation research should transition from a translational approach to an integrative research
15 approach¹⁰¹ and how personalized computer models may provide more individualised insights in
16 disease and guide therapy.¹⁰² Application of novel therapeutic tools also brings new mechanistic
17 insights. Non-invasive radiation therapy for cardiac arrhythmias was initially thought to induce fibrosis,
18 similar to invasive catheter-based therapy.¹⁰³ However, *Zhang et al* found that transmural fibrosis
19 does not develop in the hearts of patients receiving radiation therapy within the timeframe of its
20 ventricular tachycardia-reducing effects.¹⁰⁴ Interestingly, they showed that irradiating murine hearts
21 results in a persistent supraphysiologic electrical phenotype, mediated by increases in sodium
22 channel function and gap junction function. This functional restoration was confirmed by a shortening
23 of QRS duration in patients receiving radiation therapy, highlighting that radiation-induced
24 reprogramming of cardiac conduction is the potential mechanism beyond the initial success of
25 radiation therapy for refractory ventricular tachycardia. This holds promise for extending the use of
26 non-invasive radiation therapy to other applications, as for example recently demonstrated in heart
27 failure with reduced ejection fraction.¹⁰⁵
28

29 **6. CARDIOVASCULAR PRECISION MEDICINE AND iPSC**

30 Precision medicine aims to improve risk stratification and customize the management and therapy of
31 patients based on their clinical and genetic characteristics, on datasets of large populations and the
32 use of advanced technologies.¹⁰⁶ Genome-wide association studies (GWAS) has progressed through
33 advances in genome-wide genotyping technology and large population and patient datasets to
34 explore the role of common variants on phenotypic traits and disease susceptibility. According to the

1 GWAS catalogue database, there are known to be 1329 polymorphism-cardiovascular trait
2 associations. This growing catalogue of genome-wide and nominally significant variants has also
3 opened the door to creating polygenic risk scores that could identify individuals at risk of developing
4 specific cardiovascular diseases or sub-groups of patients with a more severe prognosis.¹⁰⁷ However,
5 this approach must consider numerous confounding factors such as epigenetic and transcriptomic
6 data that may correlate with genetic variants. *Boix et al* undertook a *tour de force* to create EpiMap, a
7 compendium comprising 10,000 epigenomic maps across 800 samples, which were used to define
8 chromatin states, high-resolution enhancers, enhancer modules, upstream regulators, and
9 downstream target genes.¹⁰⁸ This resource allowed the annotation of 30,000 genetic loci associated
10 with 540 traits, predicting trait-relevant tissues, putative causal nucleotide variants in enriched tissue
11 enhancers and candidate tissue-specific target genes for each of them. These different data
12 integration layers could be essential for understanding the genetic architecture underlying the broad
13 phenotypic traits encountered in common and complex cardiovascular diseases such as coronary
14 artery disease. For instance, while “only” 56 ‘unifactorial’ traits were enriched in the case of long QT
15 syndrome (LQTS), a total of 192 ‘multifactorial’ traits were enriched in an average of five different
16 tissues, and in the case of coronary artery disease, 26 ‘polyfactorial’ traits were enriched in 14
17 tissues. The study by *Boix et al* is at the same time a rich scientific resource, but also a lesson
18 regarding the profound and magnificent complexity of the human genome and the causal basis of
19 common diseases like coronary artery disease.

20
21 The GENMED consortium conducted a large GWAS study focused on dilated cardiomyopathy
22 (DCM), enrolling 2719 cases and 4440 controls.¹⁰⁹ They identified and replicated two new DCM-
23 associated loci on chromosome 3p25.1 and chromosome 22q11.23. *In silico* annotation and
24 functional 4C-sequencing analyses on cardiomyocytes derived from iPSC-derived cardiomyocytes
25 identified SLC6A6, a gene encoding a taurine, as the most likely DCM candidate at the 3p25.1 locus,
26 and SMARCB1 as the candidate culprit gene at the 22q11.23 locus. The consortium also constructed
27 a genetic risk score for DCM.

28
29 In another important study, exome sequencing data from 811 probands with tetralogy of Fallot (TOF)
30 were used to identify rare loss-of-function and other likely pathogenic variants in genes associated
31 with congenital heart disease.¹¹⁰ The role of some likely pathogenic variants was confirmed and
32 multiple loss-of-function variants provided support for 3 emerging congenital heart disease/TOF
33 candidate genes: *KDR*, *IQGAP1*, and *GDF1*. Moreover, using composite genes in a STRING protein

1 interaction enrichment analysis, a biologically relevant network was revealed, with vascular
2 endothelial growth factor receptor 2 (VEGFR2) and NOTCH1 representing central nodes.

3
4 The use of iPSC technology for disease modelling and drug testing is increasingly used for
5 cardiovascular precision medicine. Last year, for the first time, the combination of patient-specific
6 iPSC-derived cardiomyocytes, genetics and genome editing unveiled the mechanisms of action of
7 modifier genes in subsets of patients affected by long QT syndrome (LQTS).^{111, 112} By comparing
8 patient-specific iPSC-CMs derived from symptomatic and asymptomatic LQT1 carriers of the same
9 mutation, it was shown that genetic variants of *MTMR4*, an upstream regulator of neural precursor
10 cell expressed developmentally downregulated gene 4-like (NEDD4L), control potassium channel
11 turnover, thus influencing the clinical manifestations of the disease. iPSC technology has also been
12 used to gain insights into the molecular mechanisms of atrial septum defect (ASD), a form of
13 congenital heart disease, by implicating a mutation in *GATA4* that modifies *FGF16* induction.¹¹³

14
15 Pioneering work from *Srivastava* and collaborators developed a machine-learning approach to
16 identify small molecules that broadly correct gene networks dysregulated in an iPSC model of aortic
17 valve (AV) disease.¹¹⁴ Correction of the gene network by the most effective therapeutic candidate,
18 XCT790, was sufficient to prevent and treat AV disease *in vivo* in a mouse model. This strategy,
19 made possible by combining iPSC technology, network analytics and machine learning, may can
20 represent an effective path to discovering new therapies.

21 22 **7. COVID-19**

23 **7.1 Cardiovascular tropism and molecular markers**

24 The aetiology of myocarditis caused by cardiotropic viruses has become a major topic of interest
25 during the COVID-19 pandemic.^{115, 116} A comparative study revealed that while myocardial injury
26 occurred with a similar frequency in infection with influenza and SARS-CoV-2, the mortality was
27 almost 4-fold higher in COVID-19 compared with influenza.¹¹⁷ Evidence of viral infection was seen
28 mainly in endothelium and rarely in cardiomyocytes,¹¹⁸ however, evidence for stromal cells infection
29 by SARS-CoV-2 has been found.¹¹⁹ Endothelial-dependent dilation in human arterioles is impaired for
30 months after SARS-CoV-2 exposure, and could contribute to long-lasting symptoms of post-COVID-
31 19 infection.¹²⁰ Consistently, *Bräuninger et al* performed massive analysis of cDNA ends–RNAseq in
32 myocardial tissue from fatal COVID-19 cases with and without cardiac infection to reveal potential
33 SARS-CoV-2-related pro-inflammatory transcriptomic alterations in EC, while no differences were
34 detected in immune cell infiltrations.¹²¹ Interestingly, the levels of several known cardiometabolic

1 biomarkers are associated with COVID-19 severity and mortality, particularly myocyte-derived miR-
2 133a and liver-derived miR-122.¹²² The potential for the use of cardiovascular RNA markers and
3 artificial intelligence in the setting of COVID-19 has been reviewed in.¹²³ In a study of 95 SARS-CoV-
4 2-positive autopsy tissue, cardiac SARS-CoV-2 infection was shown to increase transcription of
5 interferon pathways, originating predominantly from EC.¹¹⁸ The ESC has provided guidance for the
6 diagnosis and management of cardiovascular disease during the COVID-19 pandemic^{124, 125} and
7 recommendations for future research.¹²⁶

9 **7.2 Virus- and vaccine-induced thrombotic complications and COVID-19**

10 Accumulating evidence suggests that patients suffering from COVID-19 have an increased risk to
11 experience thrombotic events such as microthrombosis, venous thromboembolism and ischaemic
12 stroke (for a review see¹²⁷). Two recent studies have found microthrombi in the hearts of patients who
13 succumbed to SARS-CoV-2 infections. *Pellegrini et al* identified microthrombi as a cause of myocyte
14 necrosis. Interestingly these microthrombi contained more fibrin and more of the complement
15 components C5b-9 than thrombi isolated from the myocardium of patients of COVID-19 negative
16 patients and coronary thrombi aspirated from COVID-19 negative and positive patients with ST-
17 *elevation* MI.¹²⁸ *Bois et al* found nonocclusive microthrombi in myocardial arterioles in 12 out of 15
18 patients who died from SARS-CoV-2 infections. However, no evidence of acute ischaemic injury of
19 the heart was detected in this study.¹²⁹ When tissue factor (TF)-bearing microvesicles isolated from
20 the plasma of 100 patients with moderate and severe COVID-19 and from the plasma of 28 healthy
21 subjects were studied, the authors found that TF-activity on such microvesicles, which is indicative of
22 a procoagulatory state, was increased in patients suffering from COVID-19 and is significantly linked
23 to disease severity and mortality.¹³⁰

24
25 Thrombotic complications have been reported in 1 per 100 000 adenoviral COVID-19 vaccinated
26 irrespective of age, rising to 1 in 50 000 above 50 years vaccinated with ChAdOx1.¹³¹ This is referred
27 to as vaccine-induced immune thrombotic thrombocytopenia (VITT).^{131, 132} Fibrinogen, Age, Platelet
28 count, and the presence of Intracranial haemorrhage, and Cerebral venous sinus thrombosis (the
29 *FAPIC* score) are significantly associated with mortality in cases of VITT.¹³³ Increased levels of anti-
30 PF4 antibodies post-vaccination unrelated to previous heparin exposure implicates an augmentation
31 of the antibody response by unknown PF4 co-factors.¹³² The antigenic component with PF4 may be
32 vaccine constituents but remains an unsolved critical question in VITT pathophysiology.¹³² The
33 immune complexes transduce platelet activation through the Fcγ receptor IIA (FcγRIIA) resulting in
34 thrombosis with concomitant thrombocytopenia accompanied by a fulminant immune activation.¹³⁴

1 Among novel therapeutic options for VITT, inhibitors of Bruton tyrosine kinase (Btk), which is used for
2 B-cell malignancies, have been explored for their ability to block FcγRIIA for preventing the
3 downstream platelet activation and aggregation. The Btk inhibitors ibrutinib and fenebrutinib
4 prevented platelet aggregation induced by serum obtained from patients with VITT.¹³⁵ Additional
5 possibly favourable effects of Btk inhibition in VITT are blocking of neutrophil-platelet complexes and
6 reduced NET release,¹³⁶ which are part of the massive immune activation during VITT.¹³⁴

7 8 **7.3 Cardiovascular drugs and COVID-19**

9 In the beginning of the COVID-19 pandemic, the interactions with cardiovascular drugs were focused
10 on ACE-inhibition and anti-thrombotic treatments¹³⁷ and more recently extended to lipid-modulating
11 agents.¹³⁸ In the latter context, omega-3 fatty acids may provide beneficial cardiovascular effects
12 through immunomodulation, anti-thrombosis and improved endothelial function.¹³⁹ Specific cytokine
13 antibodies to dampen the inflammatory storm in COVID-19 exhibit anti-inflammatory strategies
14 explored for cardiovascular prevention and have shown some success in improving survival and
15 clinical outcomes.¹⁴⁰ The RECOVERY trial tested multiple different therapeutic approaches including
16 antiviral, immunomodulatory and antithrombotic treatments, in a multi-arm factorial design inspired by
17 the International Study of Infarct Survival (ISIS) trials of the 1980s, and demonstrated benefit with
18 tocilizumab and dexamethasone, but not hydroxychloroquine, convalescent plasma or other tested
19 approaches.¹⁴¹ In a separate study, anticoagulation with low-molecular-weight heparin (LMWH) may
20 curtail viral persistence and reduce mortality.¹⁴²

21 22 **Perspectives**

23 The substantial progress of basic cardiovascular science during the past year has revealed a plethora
24 of novel therapeutic and diagnostic possibilities. Non-coding RNA, scRNAseq, and iPSC are
25 examples of discovery tools to widen the understanding of cardiac and vascular pathophysiology.
26 Through the integration cardiovascular risk factors, genetics, and biomarkers, the basic
27 cardiovascular science field is expanding towards applications in precision medicine. The year was
28 still marked by the COVID-19 pandemic and several important contributions have increased our
29 knowledge of the cardiac and thrombotic effects of SARS-CoV-2, and the underlying pathways
30 behind reported vaccinal complications. Finally, the mechanistic insights from *in vitro* and *in vivo*
31 basic science models have deepened our understanding of inflammation, CHIP, EVs, regeneration,
32 and mechanosensing in cardiovascular disease.

1
2
3 **Conflicts of interest**

4 CGT has received funding from Amgen, and personal fees from VivaLyfe, and is listed as an inventor
5 on 2 heart failure patents.
6

7
8 **FIGURE LEGENDS**
9

10 **Figure 1. Novel insights into the role of ncRNAs.**

11 Several complex loci composed of lncRNA and miRNA clusters have been identified throughout the
12 genome. Nonetheless, despite their genomic and often transcriptional overlap, they have been found
13 to have distinct functional and regulatory targets. The X-linked lncRNA MIR503HG maintains
14 endothelial cell (EC) identity by interacting with the RNA splicing regulatory protein PTBP1, with
15 decreased expression leading to broad changes associated with EndMT. Importantly, these
16 phenotypic changes seem to be independent of miR-424 and miR-503 expression, which overlap the
17 lncRNA locus³. Similarly, loss of the Cardiac Mesoderm Enhancer-associated Non-coding RNA
18 (CARMN) primes vascular smooth muscle cells (vSMCs) into a pro-atherogenic proliferative state,
19 while migration or dedifferentiation are regulated through the modulation of the overlapping miR-143
20 and miR-145⁴.
21

22 **Figure 2. Recent findings on cardiovascular mechanosensing.**

23 Newly discovered flow-stimulated mechanosensitive signalling pathways. Flow-activated PIEZO1 was
24 shown to activate the protein kinase N2 (PKN2) via PKD1, resulting in phosphorylation of Akt and
25 eNOS, with subsequent vascular tone regulation via NO.⁴⁵ The glycocalyx component sialic acid, was
26 shown to activate NRF2 antioxidant signalling, via phosphorylation of AKT⁴⁶, whereby modulating the
27 endothelial redox state in response to shear stress. The pathways are likely to be interconnected as
28 both result in phosphorylation of AKT and eNOS and as NRF2-induced antioxidant signalling is likely
29 to affect NO bioavailability.
30

31
32 **Figure 3. New insights and interventions in lipid biology.**

33 Gene silencing with small interfering RNA (siRNA) like inclisiran or gene editing are becoming
34 additional options to monoclonal antibodies for the inhibition of proprotein convertase subtilisin kexin
35 9 (PCSK9) leading to long-lasting circulating LDL-Cholesterol (LDL-C) decrease. Lipoprotein(a)

1 (Lp(a))-reducing drugs by RNA interference, via antisense oligonucleotide (ASO), like Pelacarsen or
2 siRNA, like Olpasiran are holding promising results in clinical trials. The inhibition of angiotensin-like
3 3 (ANGPTL3) via evinacumab, a monoclonal antibody or Vupanorsen, a GalNAc-conjugated ASO
4 markedly reduces circulating triglyceride-rich lipoprotein (TGRL) levels. N-acetylgalactosamine
5 (GalNAc) ligands conjugated with siRNAs or ASOs allow its hepatocyte-targeted delivery lowering
6 incidence and severity of off-target effects, commonly observed with the first generation RNA
7 interference.

9 **Figure 4. Source of EVs affects their function**

10 Several thought-provoking studies published in 2021/2022 demonstrated that the cardiovascular
11 effects of extracellular vesicles (EVs) can depend upon their origin. For example, EVs originating from
12 different cell types (cardiomyocytes vs M2 macrophages), different cellular states (proliferating vs
13 differentiated), different ages (young vs old serum) or different health states (chronic kidney disease
14 and coronary artery disease [CKD+CAD] vs healthy) can have opposite effects.

16 **Figure 5. Novel mechanisms of arrhythmia.**

17 Recent publications (top left) and accepted concepts (top right) on the mechanisms leading to re-
18 entry may be combined to arrive at a generalized theory of the spatiotemporal interaction between
19 triggers and substrate leading to re-entry arrhythmias (bottom). The generalized hypothesis highlights
20 that re-entry can only initiate when there is a local dispersion of excitability, with some tissue excitable
21 whereas other tissue is (still, or always) refractory at the time when the trigger occurs. The trigger
22 should originate from the excitable tissue, may block and travel around (relatively large) refractory
23 tissue before it arrives at the previously excited tissue again.

REFERENCES

1. Davidson SM, Padro T, Bollini S, Vilahur G, Duncker DJ, Evans PC, Guzik T, Hoefler IE, Waltenberger J, Wojta J, Weber C. Progress in cardiac research: from rebooting cardiac regeneration to a complete cell atlas of the heart. *Cardiovascular Research* 2021;**117**:2161-2174.
2. Evans P, Wojta J, Hoefler IE, Waltenberger J, Guzik T, Badimon L, Weber C. The year in basic vascular biology research: from mechanoreceptors and neutrophil extracellular traps to smartphone data and omics. *Cardiovascular research* 2021;**117**:1814-1822.
3. Monteiro JP, Rodor J, Caudrillier A, Scanlon JP, Spiroski AM, Dudnakova T, Pflüger-Müller B, Shmakova A, von Kriegsheim A, Deng L, Taylor RS, Wilson-Kanamori JR, Chen SH, Stewart K, Thomson A, Mitić T, McClure JD, Iynikkel J, Hadoke PWF, Denby L, Bradshaw AC, Caruso P, Morrell NW, Kovacic JC, Ulitsky I, Henderson NC, Caporali A, Leisegang MS, Brandes RP, Baker AH. MIR503HG Loss Promotes Endothelial-to-Mesenchymal Transition in Vascular Disease. *Circ Res* 2021;**128**:1173-1190.
4. Vacante F, Rodor J, Lalwani MK, Mahmoud AD, Bennett M, De Pace AL, Miller E, Van Kuijk K, de Bruijn J, Gijbels M, Williams TC, Clark MB, Scanlon JP, Doran AC, Montgomery R, Newby DE, Giacca M, O'Carroll D, Hadoke PWF, Denby L, Sluimer JC, Baker AH. CARMN Loss Regulates Smooth Muscle Cells and Accelerates Atherosclerosis in Mice. *Circ Res* 2021;**128**:1258-1275.
5. Gong X, Tian M, Cao N, Yang P, Xu Z, Zheng S, Liao Q, Chen C, Zeng C, Jose PA, Wang DZ, Jian Z, Xiao Y, Jiang DS, Wei X, Zhang B, Wang Y, Chen K, Wu G, Zeng C. Circular RNA circEysyt2 regulates vascular smooth muscle cell remodeling via splicing regulation. *J Clin Invest* 2021;**131**.
6. Liu X, Guo JW, Lin XC, Tuo YH, Peng WL, He SY, Li ZQ, Ye YC, Yu J, Zhang FR, Ma MM, Shang JY, Lv XF, Zhou AD, Ouyang Y, Wang C, Pang RP, Sun JX, Ou JS, Zhou JG, Liang SJ. Macrophage NFATc3 prevents foam cell formation and atherosclerosis: evidence and mechanisms. *Eur Heart J* 2021;**42**:4847-4861.
7. Schober A, Blay RM, Saboor Maleki S, Zahedi F, Winklmaier AE, Kakar MY, Baatsch IM, Zhu M, Geissler C, Fusco AE, Eberlein A, Li N, Megens RTA, Banafsche R, Kumbrink J, Weber C, Nazari-Jahantigh M. MicroRNA-21 Controls Circadian Regulation of Apoptosis in Atherosclerotic Lesions. *Circulation* 2021;**144**:1059-1073.
8. Ramanujam D, Schon AP, Beck C, Vaccarello P, Felician G, Dueck A, Esfandyari D, Meister G, Meitinger T, Schulz C, Engelhardt S. MicroRNA-21-Dependent Macrophage-to-Fibroblast Signaling Determines the Cardiac Response to Pressure Overload. *Circulation* 2021;**143**:1513-1525.
9. Hinkel R, Batkai S, Bahr A, Bozoglu T, Straub S, Borchert T, Viereck J, Howe A, Hornaschewitz N, Oberberger L, Jurisch V, Kozlik-Feldmann R, Freudenthal F, Ziegler T, Weber C, Sperandio M, Engelhardt S, Laugwitz KL, Moretti A, Klymiuk N, Thum T, Kupatt C. AntimiR-132 Attenuates Myocardial Hypertrophy in an Animal Model of Percutaneous Aortic Constriction. *J Am Coll Cardiol* 2021;**77**:2923-2935.
10. Blanco-Dominguez R, Sanchez-Diaz R, de la Fuente H, Jimenez-Borreguero LJ, Matesanz-Marin A, Relano M, Jimenez-Alejandro R, Linillos-Pradillo B, Tsilingiri K, Martin-Mariscal ML, Alonso-Herranz L, Moreno G, Martin-Asenjo R, Garcia-Guimaraes MM, Bruno KA, Dauden E, Gonzalez-Alvaro I, Villar-Guimerans LM, Martinez-Leon A, Salvador-Garicano AM, Michelhaugh SA, Ibrahim NE, Januzzi JL, Kottwitz J, Iliceto S, Plebani M, Basso C, Baritussio A, Seguso M, Marcolongo R, Ricote M, Fairweather D, Bueno H, Fernandez-Friera L, Alfonso F, Caforio ALP, Pascual-Figal DA, Heidecker B, Luscher TF, Das S, Fuster V, Ibanez B, Sanchez-Madrid F, Martin P. A Novel Circulating MicroRNA for the Detection of Acute Myocarditis. *N Engl J Med* 2021;**384**:2014-2027.
11. Eitel I, Adams V, Dieterich P, Fuernau G, de Waha S, Desch S, Schuler G, Thiele H. Relation of circulating MicroRNA-133a concentrations with myocardial damage and clinical prognosis in ST-elevation myocardial infarction. *Am Heart J* 2012;**164**:706-714.

12. Escate R, Padro T, Suades R, Camino S, Muniz O, Diaz-Diaz JL, Sionis A, Mata P, Badimon L. High miR-133a levels in the circulation anticipates presentation of clinical events in familial hypercholesterolaemia patients. *Cardiovasc Res* 2021;**117**:109-122.
13. Vanhaverbeke M, Attard R, Bartekova M, Ben-Aicha S, Brandenburger T, de Gonzalo-Calvo D, Emanuelli C, Farrugia R, Grillari J, Hackl M, Kalocayova B, Martelli F, Scholz M, Wettinger SB, Devaux Y, CA EU-CCA. Peripheral blood RNA biomarkers for cardiovascular disease from bench to bedside: A Position Paper from the EU-CardioRNA COST Action CA17129. *Cardiovasc Res* 2021.
14. Kay M, Soltani BM, Nemir M, Aghagolzadeh P, Pezzuto I, Chouvardas P, Ruberto F, Movahedi F, Ansari H, Baharvand H, Pedrazzini T. The conserved long noncoding RNA CARMA regulates cardiomyocyte differentiation. *Cardiovasc Res* 2021.
15. Modica J, Di Mauro V, Barandalla-Sobrados M, Chavez SEP, Carullo P, Nemska S, Anselmo A, Condorelli G, Iafisco M, Miragoli M, Catalucci D. Nano-miR-133a Replacement Therapy Blunts Pressure Overload-Induced Heart Failure. *Circulation* 2021;**144**:1973-1976.
16. Tyser RCV, Mahammadov E, Nakanoh S, Vallier L, Scialdone A, Srinivas S. Single-cell transcriptomic characterization of a gastrulating human embryo. *Nature* 2021;**600**:285-289.
17. Kathiriya IS, Rao KS, Iacono G, Devine WP, Blair AP, Hota SK, Lai MH, Garay BI, Thomas R, Gong HZ, Wasson LK, Goyal P, Sukonnik T, Hu KM, Akgun GA, Bernard LD, Akerberg BN, Gu F, Li K, Speir ML, Haeussler M, Pu WT, Stuart JM, Seidman CE, Seidman JG, Heyn H, Bruneau BG. Modeling Human TBX5 Haploinsufficiency Predicts Regulatory Networks for Congenital Heart Disease. *Dev Cell* 2021;**56**:292-309 e299.
18. Hu Z, Liu W, Hua X, Chen X, Chang Y, Hu Y, Xu Z, Song J. Single-Cell Transcriptomic Atlas of Different Human Cardiac Arteries Identifies Cell Types Associated With Vascular Physiology. *Arterioscler Thromb Vasc Biol* 2021;**41**:1408-1427.
19. Ma S, Sun S, Li J, Fan Y, Qu J, Sun L, Wang S, Zhang Y, Yang S, Liu Z, Wu Z, Zhang S, Wang Q, Zheng A, Duo S, Yu Y, Belmonte JCI, Chan P, Zhou Q, Song M, Zhang W, Liu GH. Single-cell transcriptomic atlas of primate cardiopulmonary aging. *Cell Res* 2021;**31**:415-432.
20. Dawson A, Li Y, Li Y, Ren P, Vasquez HG, Zhang C, Rebello KR, Ageedi W, Azares AR, Mattar AB, Sheppard MB, Lu HS, Coselli JS, Cassis LA, Daugherty A, Shen YH, LeMaire SA. Single-Cell Analysis of Aneurysmal Aortic Tissue in Patients with Marfan Syndrome Reveals Dysfunctional TGF-beta Signaling. *Genes (Basel)* 2021;**13**.
21. Wang Y, Gao H, Wang F, Ye Z, Mokry M, Turner AW, Ye J, Koplev S, Luo L, Alsaigh T, Adkar SS, Elishaev M, Gao X, Maegdefessel L, Björkegren JLM, Pasterkamp G, Miller CL, Ross EG, Leeper NJ. Dynamic changes in chromatin accessibility are associated with the atherogenic transitioning of vascular smooth muscle cells. *Cardiovasc Res* 2021.
22. van Kuijk K, Demandt JAF, Perales-Paton J, Theelen TL, Kuppe C, Marsch E, de Bruijn J, Jin H, Gijbels MJ, Matic L, Mees BME, Reutelingsperger CPM, Hedin U, Biessen EAL, Carmeliet P, Baker AH, Kramann RK, Schurgers LJ, Saez-Rodriguez J, Sluimer JC. Deficiency of myeloid phd proteins aggravates atherogenesis via macrophage apoptosis and paracrine fibrotic signaling: Atherogenic effects of myeloid PHD knockdown. *Cardiovasc Res* 2021.
23. Vallejo J, Cochain C, Zerneck A, Ley K. Heterogeneity of immune cells in human atherosclerosis revealed by scRNA-Seq. *Cardiovasc Res* 2021;**117**:2537-2543.
24. Wirka RC, Wagh D, Paik DT, Pjanic M, Nguyen T, Miller CL, Kundu R, Nagao M, Collier J, Koyano TK, Fong R, Woo YJ, Liu B, Montgomery SB, Wu JC, Zhu K, Chang R, Alamprese M, Tallquist MD, Kim JB, Quertermous T. Atheroprotective roles of smooth muscle cell phenotypic modulation and the TCF21 disease gene as revealed by single-cell analysis. *Nat Med* 2019;**25**:1280-1289.
25. Alencar GF, Owsiany KM, Karnewar S, Sukhvasi K, Mocci G, Nguyen AT, Williams CM, Shamsuzzaman S, Mokry M, Henderson CA, Haskins R, Baylis RA, Finn AV, McNamara CA, Zunder ER, Venkata V, Pasterkamp G, Björkegren J, Bekiranov S, Owens GK. Stem Cell Pluripotency Genes Klf4 and Oct4

- 1 Regulate Complex SMC Phenotypic Changes Critical in Late-Stage Atherosclerotic Lesion Pathogenesis.
2 *Circulation* 2020;**142**:2045-2059.
- 3 26. Pan H, Xue C, Auerbach BJ, Fan J, Bashore AC, Cui J, Yang DY, Trignano SB, Liu W, Shi J, Ihuegbu CO,
4 Bush EC, Worley J, Vlahos L, Laise P, Solomon RA, Connolly ES, Califano A, Sims PA, Zhang H, Li M, Reilly
5 MP. Single-Cell Genomics Reveals a Novel Cell State During Smooth Muscle Cell Phenotypic Switching
6 and Potential Therapeutic Targets for Atherosclerosis in Mouse and Human. *Circulation*
7 2020;**142**:2060-2075.
- 8 27. Chen P-Y, Qin L, Li G, Wang Z, Dahlman JE, Malagon-Lopez J, Gujja S, Cilfone NA, Kauffman KJ, Sun L,
9 Sun H, Zhang X, Aryal B, Canfran-Duque A, Liu R, Kusters P, Sehgal A, Jiao Y, Anderson DG, Gulcher J,
10 Fernandez-Hernando C, Lutgens E, Schwartz MA, Pober JS, Chittenden TW, Tellides G, Simons M.
11 Endothelial TGF- β signalling drives vascular inflammation and atherosclerosis. *Nature Metabolism*
12 2019.
- 13 28. Rodor J, Chen SH, Scanlon JP, Monteiro JP, Caudrillier A, Sweta S, Stewart KR, Shmakova A, Dobie R,
14 Henderson BEP, Stewart K, Hadoke PWF, Southwood M, Moore SD, Upton PD, Morrell NW, Li Z, Chan
15 SY, Handen A, Lafyatis R, de Rooij L, Henderson NC, Carmeliet P, Spiroski AM, Brittan M, Baker AH.
16 Single-cell RNA-seq profiling of mouse endothelial cells in response to pulmonary arterial hypertension.
17 *Cardiovasc Res* 2021.
- 18 29. Abplanalp WT, John D, Cremer S, Assmus B, Dorsheimer L, Hoffmann J, Becker-Pergola G, Rieger MA,
19 Zeiher AM, Vasa-Nicotera M, Dimmeler S. Single-cell RNA-sequencing reveals profound changes in
20 circulating immune cells in patients with heart failure. *Cardiovasc Res* 2021;**117**:484-494.
- 21 30. Abplanalp WT, Cremer S, John D, Hoffmann J, Schuhmacher B, Merten M, Rieger MA, Vasa-Nicotera M,
22 Zeiher AM, Dimmeler S. Clonal Hematopoiesis-Driver DNMT3A Mutations Alter Immune Cells in Heart
23 Failure. *Circ Res* 2021;**128**:216-228.
- 24 31. Hesse J, Owenier C, Lautwein T, Zalfen R, Weber JF, Ding Z, Alter C, Lang A, Grandoch M, Gerdes N,
25 Fischer JW, Klau GW, Dieterich C, Kohrer K, Schrader J. Single-cell transcriptomics defines
26 heterogeneity of epicardial cells and fibroblasts within the infarcted murine heart. *Elife* 2021;**10**.
- 27 32. Tombor LS, John D, Glaser SF, Luxan G, Forte E, Furtado M, Rosenthal N, Baumgarten N, Schulz MH,
28 Wittig J, Rogg EM, Manavski Y, Fischer A, Muhly-Reinholz M, Klee K, Looso M, Selnick C, Acker T, Bibli
29 SI, Fleming I, Patrick R, Harvey RP, Abplanalp WT, Dimmeler S. Single cell sequencing reveals
30 endothelial plasticity with transient mesenchymal activation after myocardial infarction. *Nat Commun*
31 2021;**12**:681.
- 32 33. Borner K, Teichmann SA, Quardokus EM, Gee JC, Browne K, Osumi-Sutherland D, Herr BW, 2nd,
33 Bueckle A, Paul H, Haniffa M, Jardine L, Bernard A, Ding SL, Miller JA, Lin S, Halushka MK, Boppana A,
34 Longacre TA, Hickey J, Lin Y, Valerius MT, He Y, Pryhuber G, Sun X, Jorgensen M, Radtke AJ, Wasserfall
35 C, Ginty F, Ho J, Sunshine J, Beuschel RT, Brusko M, Lee S, Malhotra R, Jain S, Weber G. Anatomical
36 structures, cell types and biomarkers of the Human Reference Atlas. *Nat Cell Biol* 2021;**23**:1117-1128.
- 37 34. Osumi-Sutherland D, Xu C, Keays M, Levine AP, Kharchenko PV, Regev A, Lein E, Teichmann SA. Cell
38 type ontologies of the Human Cell Atlas. *Nat Cell Biol* 2021;**23**:1129-1135.
- 39 35. Maron BA, Wang RS, Shevtsov S, Drakos SG, Arons E, Wever-Pinzon O, Huggins GS, Samokhin AO,
40 Oldham WM, Aguib Y, Yacoub MH, Rowin EJ, Maron BJ, Maron MS, Loscalzo J. Individualized
41 interactomes for network-based precision medicine in hypertrophic cardiomyopathy with implications
42 for other clinical pathophenotypes. *Nature Communications* 2021;**12**.
- 43 36. Conklin AC, Nishi H, Schlamp F, Ord T, Ounap K, Kaikkonen MU, Fisher EA, Romanoski CE. Meta-
44 Analysis of Smooth Muscle Lineage Transcriptomes in Atherosclerosis and Their Relationships to In
45 Vitro Models. *Immunometabolism* 2021;**3**.
- 46 37. Zerneck A, Winkels H, Cochain C, Williams JW, Wolf D, Soehnlein O, Robbins CS, Monaco C, Park I,
47 McNamara CA, Binder CJ, Cybulsky MI, Scipione CA, Hedrick CC, Galkina EV, Kyaw T, Ghosheh Y, Dinh

- 1 HQ, Ley K. Meta-Analysis of Leukocyte Diversity in Atherosclerotic Mouse Aortas. *Circ Res*
2 2020;**127**:402-426.
- 3 38. Ma WF, Hodonsky CJ, Turner AW, Wong D, Song Y, Mosquera JV, Ligay AV, Slenders L, Gancayco C, Pan
4 H, Barrientos NB, Mai D, Alencar GF, Owsiany K, Owens GK, Reilly MP, Li M, Pasterkamp G, Mokry M,
5 van der Laan SW, Khomtchouk BB, Miller CL. Enhanced single-cell RNA-seq workflow reveals coronary
6 artery disease cellular cross-talk and candidate drug targets. *Atherosclerosis* 2022;**340**:12-22.
- 7 39. Krane M, Dreßen M, Santamaria G, My I, Schneider CM, Dorn T, Laue S, Mastantuono E, Berutti R,
8 Rawat H, Gilsbach R, Schneider P, Lahm H, Schwarz S, Doppler SA, Paige S, Puluca N, Doll S, Neb I,
9 Brade T, Zhang Z, Abou-Ajram C, Northoff B, Holdt LM, Sudhop S, Sahara M, Goedel A, Dendorfer A,
10 Tjong FVY, Rijlaarsdam ME, Cleuziou J, Lang N, Kupatt C, Bezzina C, Lange R, Bowles NE, Mann M, Gelb
11 BD, Crotti L, Hein L, Meitinger T, Wu S, Sinnecker D, Gruber PJ, Laugwitz KL, Moretti A. Sequential
12 Defects in Cardiac Lineage Commitment and Maturation Cause Hypoplastic Left Heart Syndrome.
13 *Circulation* 2021;**144**:1409-1428.
- 14 40. Lewis-Israeli YR, Wasserman AH, Gabalski MA, Volmert BD, Ming Y, Ball KA, Yang W, Zou J, Ni G, Pajares
15 N, Chatzistavrou X, Li W, Zhou C, Aguirre A. Self-assembling human heart organoids for the modeling of
16 cardiac development and congenital heart disease. *Nat Commun* 2021;**12**:5142.
- 17 41. Fukui H, Chow RW, Xie J, Foo YY, Yap CH, Minc N, Mochizuki N, Vermot J. Bioelectric signaling and the
18 control of cardiac cell identity in response to mechanical forces. *Science (New York, NY)* 2021;**374**:351-
19 354.
- 20 42. Gracheva EO, Bagriantsev SN. Sensational channels. *Cell* 2021;**184**:6213-6216.
- 21 43. Kefauver JM, Ward AB, Patapoutian A. Discoveries in structure and physiology of mechanically
22 activated ion channels. *Nature* 2020;**587**:567-576.
- 23 44. Li J, Hou B, Tumova S, Muraki K, Bruns A, Ludlow MJ, Sedo A, Hyman AJ, McKeown L, Young RS,
24 Yuldasheva NY, Majeed Y, Wilson LA, Rode B, Bailey MA, Kim HR, Fu Z, Carter DA, Bilton J, Imrie H, Ajuh
25 P, Dear T, Cubbon RM, Kearney MT, Prasad K, Evans PC, Ainscough JF, X, Beech DJ. Piezol integration of
26 vascular architecture with physiological force. *Nature* 2014;**515**:279-U308.
- 27 45. Jin YJ, Chennupati R, Li R, Liang G, Wang S, Iring A, Graumann J, Wettschureck N, Offermanns S. Protein
28 kinase N2 mediates flow-induced endothelial NOS activation and vascular tone regulation. *J Clin Invest*
29 2021;**131**.
- 30 46. Psefteli PM, Kitscha P, Vizcay G, Fleck R, Chapple SJ, Mann GE, Fowler M, Siow RC. Glycocalyx sialic
31 acids regulate Nrf2-mediated signaling by fluid shear stress in human endothelial cells. *Redox Biol*
32 2021;**38**:101816.
- 33 47. Giebe S, Hofmann A, Brux M, Lowe F, Breheny D, Morawietz H, Brunssen C. Comparative study of the
34 effects of cigarette smoke versus next generation tobacco and nicotine product extracts on endothelial
35 function. *Redox Biol* 2021;**47**:102150.
- 36 48. Chiva-Blanch G, Evans PC. Scientists on the Spot: A matter of blood flow. *Cardiovasc Res*
37 2021;**117**:e162-e163.
- 38 49. Evans PC, Fragiadaki M, Morris PD, Serbanovic-Canic J. Shear stress: the dark energy of atherosclerotic
39 plaques. *Cardiovasc Res* 2021;**117**:1811-1813.
- 40 50. Ray KK, Landmesser U, Leiter LA, Kallend D, Dufour R, Karakas M, Hall T, Troquay RP, Turner T, Visseren
41 FL, Wijngaard P, Wright RS, Kastelein JJ. Inclisiran in patients at high cardiovascular risk with elevated
42 LDL cholesterol. *N Engl J Med* 2017;**376**:1430-1440.
- 43 51. Raal FJ, Kallend D, Ray KK, Turner T, Koenig W, Wright RS, Wijngaard PLJ, Curcio D, Jaros MJ, Leiter LA,
44 Kastelein JJP, Investigators O-. Inclisiran for the treatment of heterozygous familial
45 hypercholesterolemia. *N Engl J Med* 2020;**382**:1520-1530.
- 46 52. Tsimikas S, Karwatowska-Prokopczuk E, Gouni-Berthold I, Tardif JC, Baum SJ, Steinhagen-Thiessen E,
47 Shapiro MD, Stroes ES, Moriarty PM, Nordestgaard BG, Xia S, Guerriero J, Viney NJ, O'Dea L, Witztum

- 1 JL, Investigators AK-A-LS. Lipoprotein(a) Reduction in Persons with Cardiovascular Disease. *N Engl J*
2 *Med* 2020;**382**:244-255.
- 3 53. Katzmann JL, Cupido AJ, Laufs U. Gene Therapy Targeting PCSK9. *Metabolites* 2022;**12**.
- 4 54. Ahmad Z, Pordy R, Rader DJ, Gaudet D, Ali S, Gonzaga-Jauregui C, Ponda MP, Shumel B, Banerjee P,
5 Dunbar RL. Inhibition of Angiopoietin-Like Protein 3 With Evinacumab in Subjects With High and Severe
6 Hypertriglyceridemia. *Journal of the American College of Cardiology* 2021;**78**:193-195.
- 7 55. Raal FJ, Rosenson RS, Reeskamp LF, Hovingh GK, Kastelein JJP, Rubba P, Ali S, Banerjee P, Chan KC, Gipe
8 DA, Khilla N, Pordy R, Weinreich DM, Yancopoulos GD, Zhang Y, Gaudet D, Investigators EH.
9 Evinacumab for homozygous familial hypercholesterolemia. *N Engl J Med* 2020;**383**:711-720.
- 10 56. Alabi A, Xia XD, Gu HM, Wang F, Deng SJ, Yang N, Adijiang A, Douglas DN, Kneteman NM, Xue Y, Chen L,
11 Qin S, Wang G, Zhang DW. Membrane type 1 matrix metalloproteinase promotes LDL receptor
12 shedding and accelerates the development of atherosclerosis. *Nature Communications* 2021;**12**.
- 13 57. Ito Y, Maejima Y, Nakagama S, Shiheido-Watanabe Y, Tamura N, Sasano T. Rivaroxaban, a Direct Oral
14 Factor Xa Inhibitor, Attenuates Atherosclerosis by Alleviating Factor Xa–PAR2–Mediated Autophagy
15 Suppression. *JACC: Basic to Translational Science* 2021;**6**:964-980.
- 16 58. Edgar L, Akbar N, Braithwaite AT, Krausgruber T, Gallart-Ayala H, Bailey J, Corbin AL, Khojraty TE, Chai
17 JT, Alkhalil M, Rendeiro AF, Ziberna K, Arya R, Cahill TJ, Bock C, Laurencikiene J, Crabtree MJ, Lemieux
18 ME, Riksen NP, Netea MG, Wheelock CE, Channon KM, Rydén M, Udalova IA, Carnicer R, Choudhury
19 RP. Hyperglycemia Induces Trained Immunity in Macrophages and Their Precursors and Promotes
20 Atherosclerosis. *Circulation* 2021:961-982.
- 21 59. Niyonzima N, Rahman J, Kunz N, West EE, Freiwald T, Desai JV, Merle NS, Gidon A, Sporsheim B,
22 Lionakis MS, Evensen K, Lindberg B, Skagen K, Skjelland M, Singh P, Haug M, Ruseva MM, Kolev M,
23 Bibby J, Marshall O, O'Brien B, Deeks N, Afzali B, Clark RJ, Woodruff TM, Pryor M, Yang ZH, Remaley AT,
24 Mollnes TE, Hewitt SM, Yan B, Kazemian M, Kiss MG, Binder CJ, Halvorsen B, Espevik T, Kemper C.
25 Mitochondrial C5aR1 activity in macrophages controls IL-1 β production underlying sterile
26 inflammation. *Science Immunology* 2021;**6**:1-20.
- 27 60. Rohde D, Vandoorne K, Lee I-H, Grune J, Zhang S, McAlpine CS, Schloss MJ, Nayar R, Courties G,
28 Frodermann V, Wojtkiewicz G, Honold L, Chen Q, Schmidt S, Iwamoto Y, Sun Y, Cremer S, Hoyer FF,
29 Iborra-Egea O, Muñoz-Guijosa C, Ji F, Zhou B, Adams RH, Wythe JD, Hidalgo J, Watanabe H, Jung Y, van
30 der Laan AM, Piek JJ, Kfoury Y, Désogère PA, Vinegoni C, Dutta P, Sadreyev RI, Caravan P, Bayes-Genis
31 A, Libby P, Scadden DT, Lin CP, Naxerova K, Swirski FK, Nahrendorf M. Bone marrow endothelial
32 dysfunction promotes myeloid cell expansion in cardiovascular disease. *Nature Cardiovascular*
33 *Research* 2022;**1**:28-44.
- 34 61. Sakic A, Chaabane C, Ambartsumian N, Klingelhöfer J, Lemeille S, Kwak BR, Grigorian M, Bochaton-
35 Piallat ML. Neutralization of S100A4 induces stabilization of atherosclerotic plaques: role of smooth
36 muscle cells. *Cardiovascular research* 2022;**118**:141-155.
- 37 62. Libby P. Inflammation during the life cycle of the atherosclerotic plaque. *Cardiovasc Res*
38 2021;**117**:2525-2536.
- 39 63. Fredman G, MacNamara KC. Atherosclerosis is a major human killer and non-resolving inflammation is
40 a prime suspect. *Cardiovasc Res* 2021;**117**:2563-2574.
- 41 64. Arnardottir H, Thul S, Pawelzik SC, Karadimou G, Artiach G, Gallina AL, Mysdotter V, Carracedo M,
42 Tarnawski L, Caravaca AS, Baumgartner R, Ketelhuth DF, Olofsson PS, Paulsson-Berne G, Hansson GK,
43 Bäck M. The resolvin D1 receptor GPR32 transduces inflammation resolution and atheroprotection. *J*
44 *Clin Invest* 2021;**131**.
- 45 65. Singla B, Lin HP, Ahn W, Xu J, Ma Q, Sghayyer M, Dong K, Cherian-Shaw M, Zhou J, Huo Y, White J,
46 Csanyi G. Loss of myeloid cell-specific SIRPalpha, but not CD47, attenuates inflammation and
47 suppresses atherosclerosis. *Cardiovasc Res* 2021.

- 1 66. Bullenkamp J, Mengoni V, Kaur S, Chhetri I, Dimou P, Astroulakis ZMJ, Kaski JC, Dumitriu IE. Interleukin-
2 7 and interleukin-15 drive CD4+CD28null T lymphocyte expansion and function in patients with acute
3 coronary syndrome. *Cardiovasc Res* 2021;**117**:1935-1948.
- 4 67. Amersfoort J, Schaftenaar FH, Douna H, van Santbrink PJ, van Puijvelde GHM, Slutter B, Foks AC, Harms
5 A, Moreno-Gordaliza E, Wang Y, Hankemeier T, Bot I, Chi H, Kuiper J. Diet-induced dyslipidemia induces
6 metabolic and migratory adaptations in regulatory T cells. *Cardiovasc Res* 2021;**117**:1309-1324.
- 7 68. Bonacina F, Martini E, Svecla M, Nour J, Cremonesi M, Beretta G, Moregola A, Pellegatta F, Zampoleri
8 V, Catapano AL, Kallikourdis M, Norata GD. Adoptive transfer of CX3CR1 transduced-T regulatory cells
9 improves homing to the atherosclerotic plaques and dampens atherosclerosis progression. *Cardiovasc*
10 *Res* 2021;**117**:2069-2082.
- 11 69. Poels K, van Leent MMT, Boutros C, Tissot H, Roy S, Meerwaldt AE, Toner YCA, Reiche ME, Kusters PJH,
12 Malinova T, Huveneers S, Kaufman AE, Mani V, Fayad ZA, de Winther MPJ, Marabelle A, Mulder WJM,
13 Robert C, Seijkens TTP, Lutgens E. Immune Checkpoint Inhibitor Therapy Aggravates T Cell-Driven
14 Plaque Inflammation in Atherosclerosis. *JACC CardioOncology* 2020;**2**:599-610.
- 15 70. Tocchetti CG, Ameri P, de Boer RA, D'Alessandra Y, Russo M, Sorriento D, Ciccarelli M, Kiss B, Bertrand
16 L, Dawson D, Falcao-Pires I, Giacca M, Hamdani N, Linke WA, Mayr M, van der Velden J, Zacchigna S,
17 Ghigo A, Hirsch E, Lyon AR, Gorbe A, Ferdinandy P, Madonna R, Heymans S, Thum T. Cardiac
18 dysfunction in cancer patients: beyond direct cardiomyocyte damage of anticancer drugs. Novel cardio-
19 oncology insights from the joint 2019 meeting of the ESC Working Groups of Myocardial Function and
20 Cellular Biology of the Heart. *Cardiovascular research* 2020.
- 21 71. Michel L, Helfrich I, Hendgen-Cotta UB, Mincu RI, Korste S, Mrotzek SM, Spomer A, Odersky A,
22 Rischpler C, Herrmann K, Umutlu L, Coman C, Ahrends R, Sickmann A, Loffek S, Livingstone E, Ugurel S,
23 Zimmer L, Gunzer M, Schadendorf D, Totzeck M, Rassaf T. Targeting early stages of cardiotoxicity from
24 anti-PD1 immune checkpoint inhibitor therapy. *European heart journal* 2021.
- 25 72. Varricchi G, Galdiero MR, Tocchetti CG. Novel actors on the stage of cardiac dysfunction induced by
26 anti-PD1 oncological treatments. *European heart journal* 2021.
- 27 73. Marnell CS, Bick A, Natarajan P. Clonal hematopoiesis of indeterminate potential (CHIP): Linking
28 somatic mutations, hematopoiesis, chronic inflammation and cardiovascular disease. *J Mol Cell Cardiol*
29 **2021**;**161**:98-105.
- 30 74. Wang Y, Sano S, Ogawa H, Horitani K, Evans MA, Yura Y, Miura-Yura E, Doviak H, Walsh K. Murine
31 models of clonal hematopoiesis to assess mechanisms of cardiovascular disease. *Cardiovasc Res* 2021.
- 32 75. Fidler TP, Xue C, Yalcinkaya M, Hardaway B, Abramowicz S, Xiao T, Liu W, Thomas DG, Hajebrahimi MA,
33 Pircher J, Silvestre-Roig C, Kotini AG, Luchsinger LL, Wei Y, Westerterp M, Snoeck HW, Papapetrou EP,
34 Schulz C, Massberg S, Soehnlein O, Ebert B, Levine RL, Reilly MP, Libby P, Wang N, Tall AR. The AIM2
35 inflammasome exacerbates atherosclerosis in clonal haematopoiesis. *Nature* 2021;**592**:296-301.
- 36 76. Heyde A, Rohde D, McAlpine CS, Zhang S, Hoyer FF, Gerold JM, Cheek D, Iwamoto Y, Schloss MJ,
37 Vandoorne K, Iborra-Egea O, Munoz-Guijosa C, Bayes-Genis A, Reiter JG, Craig M, Swirski FK,
38 Nahrendorf M, Nowak MA, Naxerova K. Increased stem cell proliferation in atherosclerosis accelerates
39 clonal hematopoiesis. *Cell* 2021;**184**:1348-1361 e1322.
- 40 77. Li J, Salvador AM, Li G, Valkov N, Ziegler O, Yeri A, Yang Xiao C, Meechoovet B, Alsop E, Rodosthenous
41 RS, Kundu P, Huan T, Levy D, Tigges J, Pico AR, Ghiran I, Silverman MG, Meng X, Kitchen R, Xu J, Van
42 Keuren-Jensen K, Shah R, Xiao J, Das S. Mir-30d Regulates Cardiac Remodeling by Intracellular and
43 Paracrine Signaling. *Circ Res* 2021;**128**:e1-e23.
- 44 78. Wang Y, Li C, Zhao R, Qiu Z, Shen C, Wang Z, Liu W, Zhang W, Ge J, Shi B. CircUbe3a from M2
45 macrophage-derived small extracellular vesicles mediates myocardial fibrosis after acute myocardial
46 infarction. *Theranostics* 2021;**11**:6315-6333.
- 47 79. Lima Correa B, El Harane N, Gomez I, Rachid Hocine H, Vilar J, Desgres M, Bellamy V, Keirthishana K,
48 Guillas C, Perotto M, Pidial L, Alayrac P, Tran T, Tan S, Hamada T, Charron D, Brisson A, Renault NK, Al-

- 1 Daccak R, Menasche P, Silvestre JS. Extracellular vesicles from human cardiovascular progenitors
2 trigger a reparative immune response in infarcted hearts. *Cardiovasc Res* 2021;**117**:292-307.
- 3 80. Patil M, Saheera S, Dubey PK, Kahn-Krell A, Kumar Govindappa P, Singh S, Tousif S, Zhang Q, Lal H,
4 Zhang J, Qin G, Krishnamurthy P. Novel Mechanisms of Exosome-Mediated Phagocytosis of Dead Cells
5 in Injured Heart. *Circ Res* 2021;**129**:1006-1020.
- 6 81. Katsur M, He Z, Vinokur V, Corteling R, Yellon DM, Davidson SM. Exosomes from neuronal stem cells
7 may protect the heart from ischaemia/reperfusion injury via JAK1/2 and gp130. *J Cell Mol Med*
8 2021;**25**:4455-4465.
- 9 82. Zhang H, Lin S, McElroy CL, Wang B, Jin D, Uteshev VV, Jin K. Circulating Pro-Inflammatory Exosomes
10 Worsen Stroke Outcomes in Aging. *Circ Res* 2021;**129**:e121-e140.
- 11 83. Villa Del Campo C, Liaw NY, Gunadasa-Rohling M, Matthaei M, Braga L, Kennedy T, Salinas G, Voigt N,
12 Giacca M, Zimmermann WH, Riley PR. Regenerative potential of epicardium-derived extracellular
13 vesicles mediated by conserved miRNA transfer. *Cardiovasc Res* 2021.
- 14 84. Jung JH, Ikeda G, Tada Y, von Bornstadt D, Santoso MR, Wahlquist C, Rhee S, Jeon YJ, Yu AC, O'Brien C
15 G, Red-Horse K, Appel EA, Mercola M, Woo J, Yang PC. miR-106a-363 cluster in extracellular vesicles
16 promotes endogenous myocardial repair via Notch3 pathway in ischemic heart injury. *Basic Res Cardiol*
17 2021;**116**:19.
- 18 85. Balbi C, Milano G, Fertig TE, Lazzarini E, Bolis S, Taniyama Y, Sanada F, Di Silvestre D, Mauri P,
19 Gherghiceanu M, Luscher TF, Barile L, Vassalli G. An exosomal-carried short periostin isoform induces
20 cardiomyocyte proliferation. *Theranostics* 2021;**11**:5634-5649.
- 21 86. Ikeda G, Santoso MR, Tada Y, Li AM, Vaskova E, Jung JH, O'Brien C, Egan E, Ye J, Yang PC. Mitochondria-
22 Rich Extracellular Vesicles From Autologous Stem Cell-Derived Cardiomyocytes Restore Energetics of
23 Ischemic Myocardium. *J Am Coll Cardiol* 2021;**77**:1073-1088.
- 24 87. Lima Correa B, El Harane N, Desgres M, Perotto M, Alayrac P, Guillas C, Pidial L, Bellamy V, Baron E,
25 Autret G, Kamaleswaran K, Pezzana C, Perier MC, Vilar J, Alberdi A, Brisson A, Renault N, Gnechi M,
26 Silvestre JS, Menasche P. Extracellular vesicles fail to trigger the generation of new cardiomyocytes in
27 chronically infarcted hearts. *Theranostics* 2021;**11**:10114-10124.
- 28 88. Wei Z, Chen Z, Zhao Y, Fan F, Xiong W, Song S, Yin Y, Hu J, Yang K, Yang L, Xu B, Ge J. Mononuclear
29 phagocyte system blockade using extracellular vesicles modified with CD47 on membrane surface for
30 myocardial infarction reperfusion injury treatment. *Biomaterials* 2021;**275**:121000.
- 31 89. Lai Y, Zhou X, Guo F, Jin X, Meng G, Zhou L, Chen H, Liu Z, Yu L, Jiang H. Non-invasive transcutaneous
32 vagal nerve stimulation improves myocardial performance in doxorubicin-induced cardiotoxicity.
33 *Cardiovascular research* 2021.
- 34 90. Chan BYH, Roczkowsky A, Cho WJ, Poirier M, Sergi C, Keschrumrus V, Churko JM, Granzier H, Schulz R.
35 MMP inhibitors attenuate doxorubicin cardiotoxicity by preventing intracellular and extracellular
36 matrix remodelling. *Cardiovascular research* 2021;**117**:188-200.
- 37 91. Galan-Arriola C, Vilchez-Tschischke JP, Lobo M, Lopez GJ, de Molina-Iracheta A, Perez-Martinez C,
38 Villena-Gutierrez R, Macias A, Diaz-Rengifo IA, Oliver E, Fuster V, Sanchez-Gonzalez J, Ibanez B.
39 Coronary microcirculation damage in anthracycline cardiotoxicity. *Cardiovascular research*
40 2022;**118**:531-541.
- 41 92. Galan-Arriola C, Villena-Gutierrez R, Higuero-Verdejo MI, Diaz-Rengifo IA, Pizarro G, Lopez GJ, Molina-
42 Iracheta A, Perez-Martinez C, Garcia RD, Gonzalez-Calle D, Lobo M, Sanchez PL, Oliver E, Cordoba R,
43 Fuster V, Sanchez-Gonzalez J, Ibanez B. Remote ischaemic preconditioning ameliorates anthracycline-
44 induced cardiotoxicity and preserves mitochondrial integrity. *Cardiovascular research* 2021;**117**:1132-
45 1143.
- 46 93. Heusch G, Rassaf T. Protection from cardiotoxicity of cancer chemotherapy: a novel target for remote
47 ischaemic conditioning? *Cardiovascular research* 2021;**117**:985-986.

- 1 94. Smith AJ. Effects of Cardiotoxins on Cardiac Stem and Progenitor Cell Populations. *Frontiers in*
2 *Cardiovascular Medicine* 2021;**8**.
- 3 95. Lan HR, Xue Q, Liu YY, Jin KT, Fang XL, Shao H. The emerging therapeutic role of mesenchymal stem
4 cells in anthracycline-induced cardiotoxicity. *Cell and Tissue Research* 2021;**384**:1-12.
- 5 96. Kwok M, Lee C, Li HS, Deng R, Tsoi C, Ding Q, Tsang SY, Leung KT, Yan BP, Poon EN. Remdesivir induces
6 persistent mitochondrial and structural damage in human induced pluripotent stem cell derived
7 cardiomyocytes. *Cardiovascular research* 2021.
- 8 97. Handa BS, Li X, Baxan N, Roney CH, Shchendrygina A, Mansfield CA, Jabbour RJ, Pitcher DS, Chowdhury
9 RA, Peters NS, Ng FS. Ventricular fibrillation mechanism and global fibrillatory organization are
10 determined by gap junction coupling and fibrosis pattern. *Cardiovasc Res* 2021;**117**:1078-1090.
- 11 98. Cluitmans MJM, Bear LR, Nguyen UC, van Rees B, Stoks J, Ter Bekke RMA, Muhl C, Heijman J, Lau KD,
12 Vigmond E, Bayer J, Belterman CNW, Abell E, Labrousse L, Rogier J, Bernus O, Haissaguerre M, Hassink
13 RJ, Dubois R, Coronel R, Volders PGA. Noninvasive detection of spatiotemporal activation-
14 repolarization interactions that prime idiopathic ventricular fibrillation. *Sci Transl Med*
15 2021;**13**:eabi9317.
- 16 99. Leong KMW, Ng FS, Shun-Shin MJ, Koa-Wing M, Qureshi N, Whinnett ZI, Linton NF, Lefroy D, Francis
17 DP, Harding SE, Davies DW, Peter NS, Lim PB, Behr E, Lambiase PD, Varnava A, Kanagaratnam P. Non-
18 invasive detection of exercise-induced cardiac conduction abnormalities in sudden cardiac death
19 survivors in the inherited cardiac conditions. *Europace* 2021;**23**:305-312.
- 20 100. Quintanilla JG, Shpun S, Jalife J, Filgueiras-Rama D. Novel approaches to mechanism-based atrial
21 fibrillation ablation. *Cardiovasc Res* 2021;**117**:1662-1681.
- 22 101. Schotten U. From translation to integration: how to approach the complexity of atrial fibrillation
23 mechanisms. *Cardiovasc Res* 2021;**117**:e88-e90.
- 24 102. Heijman J, Sutanto H, Crijns H, Nattel S, Trayanova NA. Computational models of atrial fibrillation:
25 achievements, challenges, and perspectives for improving clinical care. *Cardiovasc Res* 2021;**117**:1682-
26 1699.
- 27 103. Cuculich PS, Schill MR, Kashani R, Mutic S, Lang A, Cooper D, Faddis M, Gleva M, Noheria A, Smith TW,
28 Hallahan D, Rudy Y, Robinson CG. Noninvasive Cardiac Radiation for Ablation of Ventricular
29 Tachycardia. *N Engl J Med* 2017;**377**:2325-2336.
- 30 104. Zhang DM, Navara R, Yin T, Szymanski J, Goldsztejn U, Kenkel C, Lang A, Mpoy C, Lipovsky CE, Qiao Y,
31 Hicks S, Li G, Moore KMS, Bergom C, Rogers BE, Robinson CG, Cuculich PS, Schwarz JK, Rentschler SL.
32 Cardiac radiotherapy induces electrical conduction reprogramming in the absence of transmural
33 fibrosis. *Nat Commun* 2021;**12**:5558.
- 34 105. Dusi V, Vitolo V, Frigerio L, Totaro R, Valentini A, Barcellini A, Mirandola A, Perego GB, Coccia M, Greco
35 A, Ghio S, Valvo F, De Ferrari GM, Gneccchi M, Oltrona Visconti L, Rordorf R. First-in-man case of non-
36 invasive proton radiotherapy for the treatment of refractory ventricular tachycardia in advanced heart
37 failure. *Eur J Heart Fail* 2021;**23**:195-196.
- 38 106. Gneccchi M, Sala L, Schwartz PJ. Precision Medicine and cardiac channelopathies: when dreams meet
39 reality. *European Heart Journal* 2021;**42**:1661-1675.
- 40 107. Barc J, Kovacic JC. From polygenic risk scores to integrative epigenomics: the dawn of a new era for
41 cardiovascular precisionmedicine. *Cardiovascular Research* 2021;**117**:E73-E75.
- 42 108. Boix CA, James BT, Park YP, Meuleman W, Kellis M. Regulatory genomic circuitry of human disease loci
43 by integrative epigenomics. *Nature* 2021;**590**:300-307.
- 44 109. Garnier S, Harakalova M, Weiss S, Mokry M, Regitz-Zagrosek V, Hengstenberg C, Cappola TP, Isnard R,
45 Arbustini E, Cook SA, van Setten J, Calis JJA, Hakonarson H, Morley MP, Stark K, Prasad SK, Li J, O'Regan
46 DP, Grasso M, Muller-Nurasyid M, Meitinger T, Empana JP, Strauch K, Waldenberger M, Marguiles KB,
47 Seidman CE, Kararigas G, Meder B, Haas J, Boutouyrie P, Lacolley P, Jouven X, Erdmann J, Blankenberg
48 S, Wichter T, Ruppert V, Tavazzi L, Dubourg O, Roizes G, Dorent R, de Groote P, Fauchier L, Trochu JN,

- 1 Aupetit JF, Bilinska ZT, Germain M, Volker U, Hemerich D, Raji I, Bacq-Daian D, Proust C, Remior P,
2 Gomez-Bueno M, Lehnert K, Maas R, Olaso R, Saripella GV, Felix SB, McGinn S, Duboscq-Bidot L, van
3 Mil A, Besse C, Fontaine V, Blanche H, Ader F, Keating B, Curjol A, Boland A, Komajda M, Cambien F,
4 Deleuze JF, Dorr M, Asselbergs FW, Villard E, Tregouet DA, Charron P, Consortium G. Genome-wide
5 association analysis in dilated cardiomyopathy reveals two new players in systolic heart failure on
6 chromosomes 3p25.1 and 22q11.23. *European Heart Journal* 2021;**42**:2000-2011.
- 7 110. Reuter MS, Chaturvedi RR, Jobling RK, Pellicchia G, Hamdan O, Sung WWL, Nalpathamkalam T, Attaluri
8 P, Silversides CK, Wald RM, Marshall CR, Williams SG, Keavney BD, Thiruvahindrapuram B, Scherer SW,
9 Bassett AS. Clinical Genetic Risk Variants Inform a Functional Protein Interaction Network for Tetralogy
10 of Fallot. *Circulation-Genomic and Precision Medicine* 2021;**14**:476-484.
- 11 111. Lee YK, Sala L, Mura M, Rocchetti M, Pedrazzini M, Ran XR, Mak TSH, Crotti L, Sham PC, Torre E, Zaza A,
12 Schwartz PJ, Tse HF, Gneccchi M. MTMR4 SNVs modulate ion channel degradation and clinical severity
13 in congenital long QT syndrome: insights in the mechanism of action of protective modifier genes.
14 *Cardiovascular Research* 2021;**117**:767-779.
- 15 112. Ronchi C, Bernardi J, Mura M, Stefanello M, Badone B, Rocchetti M, Crotti L, Brink P, Schwartz PJ,
16 Gneccchi M, Zaza A. NOS1AP polymorphisms reduce NOS1 activity and interact with prolonged
17 repolarization in arrhythmogenesis. *Cardiovascular Research* 2021;**117**:472-483.
- 18 113. Ye L, Yu Y, Zhao ZA, Zhao D, Ni X, Wang Y, Fang X, Yu M, Wang Y, Tang JM, Chen Y, Shen Z, Lei W, Hu S.
19 Patient-specific iPSC-derived cardiomyocytes reveal abnormal regulation of FGF16 in a familial atrial
20 septal defect. *Cardiovascular research* 2021.
- 21 114. Theodoris CV, Zhou P, Liu L, Zhang Y, Nishino T, Huang Y, Kostina A, Ranade SS, Gifford CA, Uspenskiy V,
22 Malashicheva A, Ding S, Srivastava D. Network-based screen in iPSC-derived cells reveals therapeutic
23 candidate for heart valve disease. *Science* 2021;**371**:693-+.
- 24 115. Schultheiss HP, Baumeier C, Pietsch H, Bock CT, Poller W, Escher F. Cardiovascular consequences of
25 viral infections: from COVID to other viral diseases. *Cardiovascular Research* 2021;**117**:2610-2623.
- 26 116. Raman B, Bluemke DA, Lüscher TF, Neubauer S. Long COVID: post-acute sequelae of COVID-19 with a
27 cardiovascular focus. *European heart journal* 2022.
- 28 117. Biasco L, Klersy C, Beretta GS, Valgimigli M, Valotta A, Gabutti L, Bruna RD, Pagnamenta A, Tersalvi G,
29 Ruinelli L, Artero A, Senatore G, Jüni P, Pedrazzini GB. Comparative frequency and prognostic impact of
30 myocardial injury in hospitalized patients with COVID-19 and Influenza. *European Heart Journal Open*
31 2021;**1**.
- 32 118. Bräuninger H, Stoffers B, Fitzek ADE, Meißner K, Aleshcheva G, Schweizer M, Weimann J, Rotter B,
33 Warnke S, Edler C, Braun F, Roedl K, Scherschel K, Escher F, Kluge S, Huber TB, Ondruschka B,
34 Schultheiss HP, Kirchhof P, Blankenberg S, Püschel K, Westermann D, Lindner D. Cardiac SARS-CoV-2
35 infection is associated with pro-inflammatory transcriptomic alterations within the heart.
36 *Cardiovascular research* 2022;**118**:542-555.
- 37 119. Amendola A, Garoffolo G, Songia P, Nardacci R, Ferrari S, Bernava G, Canzano P, Myasoedova V,
38 Colavita F, Castilletti C, Sberna G, Capobianchi MR, Piacentini M, Agrifoglio M, Colombo GI, Poggio P,
39 Pesce M. Human cardiosphere-derived stromal cells exposed to SARS-CoV-2 evolve into hyper-
40 inflammatory/pro-fibrotic phenotype and produce infective viral particles depending on the levels of
41 ACE2 receptor expression. *Cardiovascular Research* 2021;**117**:1557-1566.
- 42 120. Nishijima Y, Hader SN, Hanson AJ, Zhang DX, Sparapani R, Gutterman DD, Beyer AM. Prolonged
43 endothelial-dysfunction in human arterioles following infection with SARS-CoV-2. *Cardiovascular*
44 *research* 2022;**118**:18-19.
- 45 121. Brauninger H, Stoffers B, Fitzek ADE, Meissner K, Aleshcheva G, Schweizer M, Weimann J, Rotter B,
46 Warnke S, Edler C, Braun F, Roedl K, Scherschel K, Escher F, Kluge S, Huber TB, Ondruschka B,
47 Schultheiss HP, Kirchhof P, Blankenberg S, Puschel K, Westermann D, Lindner D. Cardiac SARS-CoV-2

- infection is associated with pro-inflammatory transcriptomic alterations within the heart. *Cardiovasc Res* 2021.
122. Gutmann C, Khamina K, Theofilatos K, Diendorfer AB, Burnap SA, Nabeebaccus A, Fish M, McPhail MJW, O'Gallagher K, Schmidt LE, Cassel C, Auzinger G, Napoli S, Mujib SF, Trovato F, Sanderson B, Merrick B, Roy R, Edgeworth JD, Shah AM, Hayday AC, Traby L, Hackl M, Eichinger S, Shankar-Hari M, Mayr M. Association of cardiometabolic microRNAs with COVID-19 severity and mortality. *Cardiovascular research* 2022;**118**:461-474.
123. Badimon L, Robinson EL, Jusic A, Carpusca I, DeWindt LJ, Emanuelli C, Ferdinandy P, Gu W, Gyongyosi M, Hackl M, Karaduzovic-Hadziabdic K, Lustrek M, Martelli F, Nham E, Potocnjak I, Satagopam V, Schneider R, Thum T, Devaux Y. Cardiovascular RNA markers and artificial intelligence may improve COVID-19 outcome: a position paper from the EU-CardioRNA COST Action CA17129. *Cardiovascular Research* 2021;**117**:1823-1840.
124. European Society of Cardiology guidance for the diagnosis and management of cardiovascular disease during the COVID-19 pandemic: part 1-epidemiology, pathophysiology, and diagnosis. *Cardiovascular research* 2021.
125. ESC guidance for the diagnosis and management of cardiovascular disease during the COVID-19 pandemic: part 2-care pathways, treatment, and follow-up. *Cardiovascular research* 2021.
126. Pesce M, Agostoni P, Botker HE, Brundel B, Davidson SM, De Caterina R, Ferdinandy P, Girao H, Gyongyosi M, Hulot JS, Lecour S, Perrino C, Schulz R, Sluijter JP, Steffens S, Tancevski I, Gollmann-Tepekoylu C, Tschope C, van Linthout S, Madonna R. COVID-19-related cardiac complications from clinical evidences to basic mechanisms: opinion paper of the ESC Working Group on Cellular Biology of the Heart. *Cardiovascular Research* 2021;**117**:2148-2160.
127. McFadyen JD, Stevens H, Peter K. The Emerging Threat of (Micro)Thrombosis in COVID-19 and Its Therapeutic Implications. *Circ Res* 2020;**127**:571-587.
128. Pellegrini D, Kawakami R, Guagliumi G, Sakamoto A, Kawai K, Gianatti A, Nasr A, Kutys R, Guo L, Cornelissen A, Faggi L, Mori M, Sato Y, Pescetelli I, Brivio M, Romero M, Virmani R, Finn AV. Microthrombi as a Major Cause of Cardiac Injury in COVID-19: A Pathologic Study. *Circulation* 2021;**143**:1031-1042.
129. Bois MC, Boire NA, Layman AJ, Aubry MC, Alexander MP, Roden AC, Hagen CE, Quinton RA, Larsen C, Erben Y, Majumdar R, Jenkins SM, Kipp BR, Lin PT, Maleszewski JJ. COVID-19-Associated Nonocclusive Fibrin Microthrombi in the Heart. *Circulation* 2021;**143**:230-243.
130. Rosell A, Havervall S, von Meijenfeldt F, Hisada Y, Aguilera K, Grover SP, Lisman T, Mackman N, Thålin C. Patients With COVID-19 Have Elevated Levels of Circulating Extracellular Vesicle Tissue Factor Activity That Is Associated With Severity and Mortality-Brief Report. *Arteriosclerosis, thrombosis, and vascular biology* 2021;**41**:878-882.
131. Marchandot B, Curtiaud A, Trimaille A, Sattler L, Grunebaum L, Morel O. Vaccine-induced immune thrombotic thrombocytopenia: current evidence, potential mechanisms, clinical implications, and future directions. *Eur Heart J Open* 2021;**1**.
132. McFadyen JD, Peter K. The known knowns and known unknowns of vaccine-induced thrombotic thrombocytopenia. *Cardiovasc Res* 2021;**117**:e147-e150.
133. Hwang J, Park SH, Lee SW, Lee SB, Lee MH, Jeong GH, Kim MS, Kim JY, Koyanagi A, Jacob L, Jung SY, Song J, Yon DK, Shin JI, Smith L. Predictors of mortality in thrombotic thrombocytopenia after adenoviral COVID-19 vaccination: the FAPIC score. *Eur Heart J* 2021;**42**:4053-4063.
134. Holm S, Kared H, Michelsen AE, Kong XY, Dahl TB, Schultz NH, Nyman TA, Fladeby C, Seljeflot I, Ueland T, Stensland M, Mjaaland S, Goll GL, Nissen-Meyer LS, Aukrust P, Skagen K, Gregersen I, Skjelland M, Holme PA, Munthe LA, Halvorsen B. Immune complexes, innate immunity, and NETosis in ChAdOx1 vaccine-induced thrombocytopenia. *Eur Heart J* 2021;**42**:4064-4072.

- 1 135. Weber C, von Hundelshausen P, Siess W. VITT after ChAdOx1 nCoV-19 Vaccination. *N Engl J Med*
2 2021;**385**:2203-2204.
- 3 136. von Hundelshausen P, Lorenz R, Siess W, Weber C. Vaccine-Induced Immune Thrombotic
4 Thrombocytopenia (VITT): Targeting Pathomechanisms with Bruton Tyrosine Kinase Inhibitors. *Thromb*
5 *Haemost* 2021;**121**:1395-1399.
- 6 137. Cenko E, Badimon L, Bugiardini R, Claeys MJ, De Luca G, de Wit C, Derumeaux G, Dorobantu M,
7 Duncker DJ, Eringa EC, Gorog DA, Hassager C, Heinzl FR, Huber K, Manfrini O, Milicic D, Oikonomou E,
8 Padro T, Trifunovic-Zamaklar D, Vasiljevic-Pokrajcic Z, Vavlukis M, Vilahur G, Tousoulis D.
9 Cardiovascular disease and COVID-19: a consensus paper from the ESC Working Group on Coronary
10 Pathophysiology & Microcirculation, ESC Working Group on Thrombosis and the Association for Acute
11 Cardiovascular Care (ACVC), in collaboration with the European Heart Rhythm Association (EHRA).
12 *Cardiovascular research* 2021;**117**:2705-2729.
- 13 138. Talasaz AH, Sadeghipour P, Aghakouchakzadeh M, Dreyfus I, Kakavand H, Ariannejad H, Gupta A,
14 Madhavan MV, Van Tassell BW, Jimenez D, Monreal M, Vaduganathan M, Fanikos J, Dixon DL, Piazza G,
15 Parikh SA, Bhatt DL, Lip GYH, Stone GW, Krumholz HM, Libby P, Goldhaber SZ, Bikdeli B. Investigating
16 Lipid-Modulating Agents for Prevention or Treatment of COVID-19 JACC State-of-the-Art Review.
17 *Journal of the American College of Cardiology* 2021;**78**:1635-1654.
- 18 139. Zhu YF, Wen L, Wang S, Zhang K, Cui Y, Zhang C, Feng L, Yu F, Chen YQ, Wang RX, Ma X. Omega-3 fatty
19 acids improve flow-induced vasodilation by enhancing TRPV4 in arteries from diet-induced obese mice.
20 *Cardiovascular Research* 2021;**117**:2450-2458.
- 21 140. Ridker PM. Targeting cytokine storm in COVID-19: what have we learned? *European Heart Journal*
22 *Open* 2021;**1**.
- 23 141. Pessoa-Amorim G, Mafham MM. The RECOVERY trial: cardiovascular implications of a large, simple
24 randomized trial in COVID-19. *Cardiovascular Research* 2021;**117**:E110-E113.
- 25 142. Pereyra D, Heber S, Schrottmaier WC, Santol J, Pirabe A, Schmuckenschlager A, Kammerer K, Ammon
26 D, Sorz T, Fritsch F, Hayden H, Pawelka E, Krüger P, Rumpf B, Traugott MT, Glaser P, Firbas C,
27 Schörghofer C, Seitz T, Karolyi M, Pabinger I, Brostjan C, Starlinger P, Weiss G, Bellmann-Weiler R,
28 Salzer HJF, Jilma B, Zoufaly A, Assinger A. Low-molecular-weight heparin use in coronavirus disease
29 2019 is associated with curtailed viral persistence: a retrospective multicentre observational study.
30 *Cardiovascular research* 2021;**117**:2807-2820.
- 31
32

Figure 1. Novel insights into the role of ncRNAs.

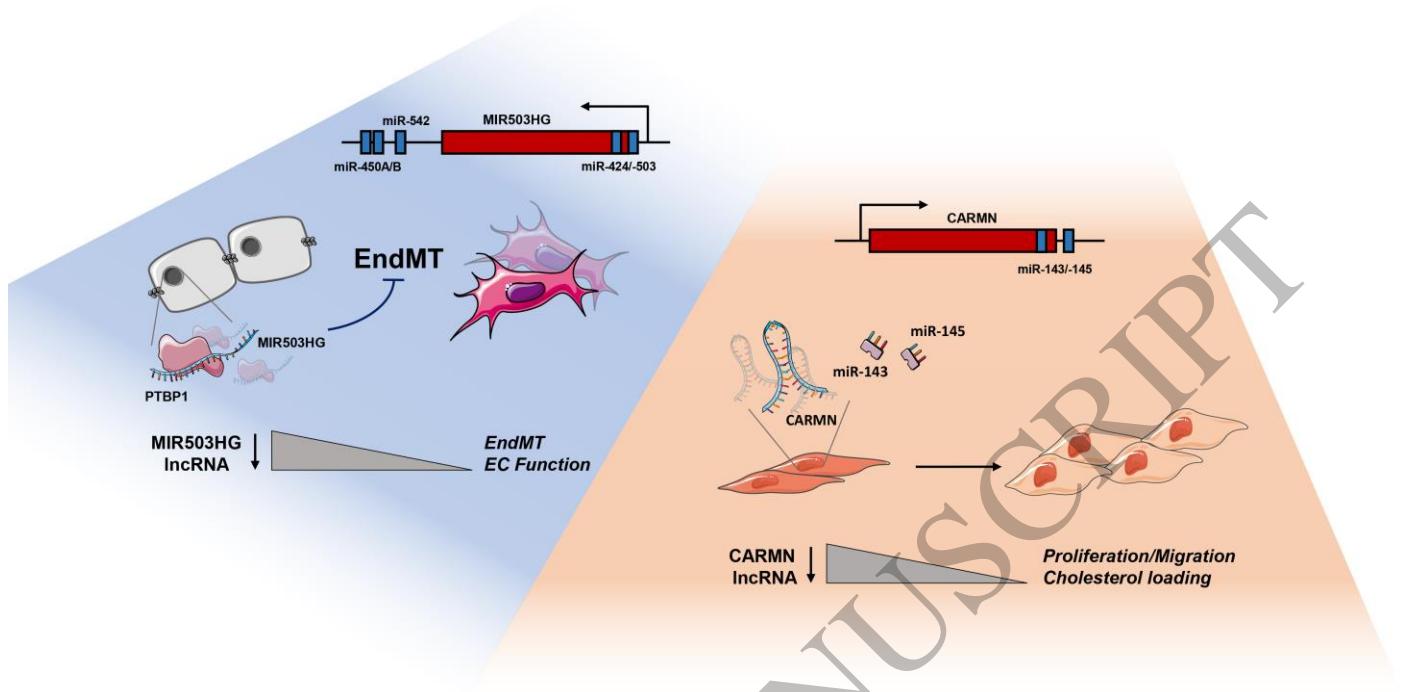


Figure 1
185x103 mm (x DPI)

1
2
3
4

Figure 2. Progress in mechanosensing.

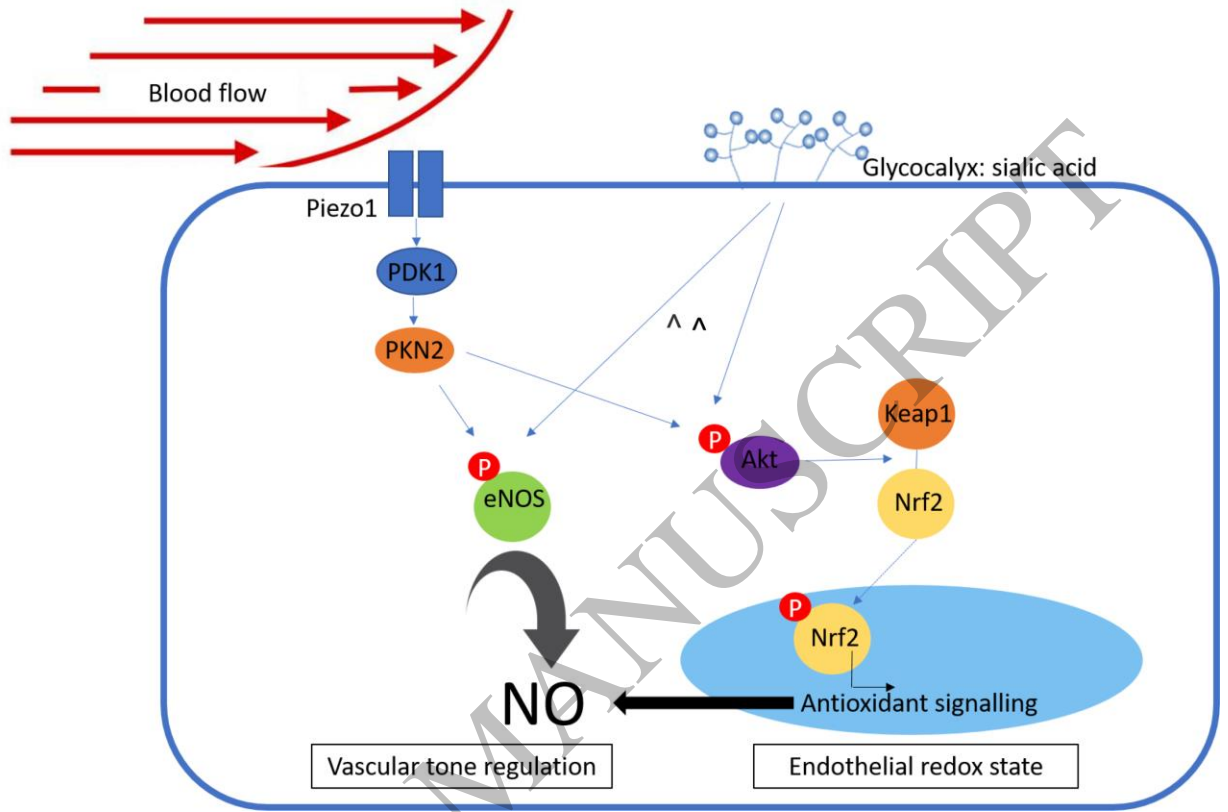


Figure 2
185x129 mm (x DPI)

1
2
3
4

Figure 3. New insights in lipid biology and cross-talk with inflammation

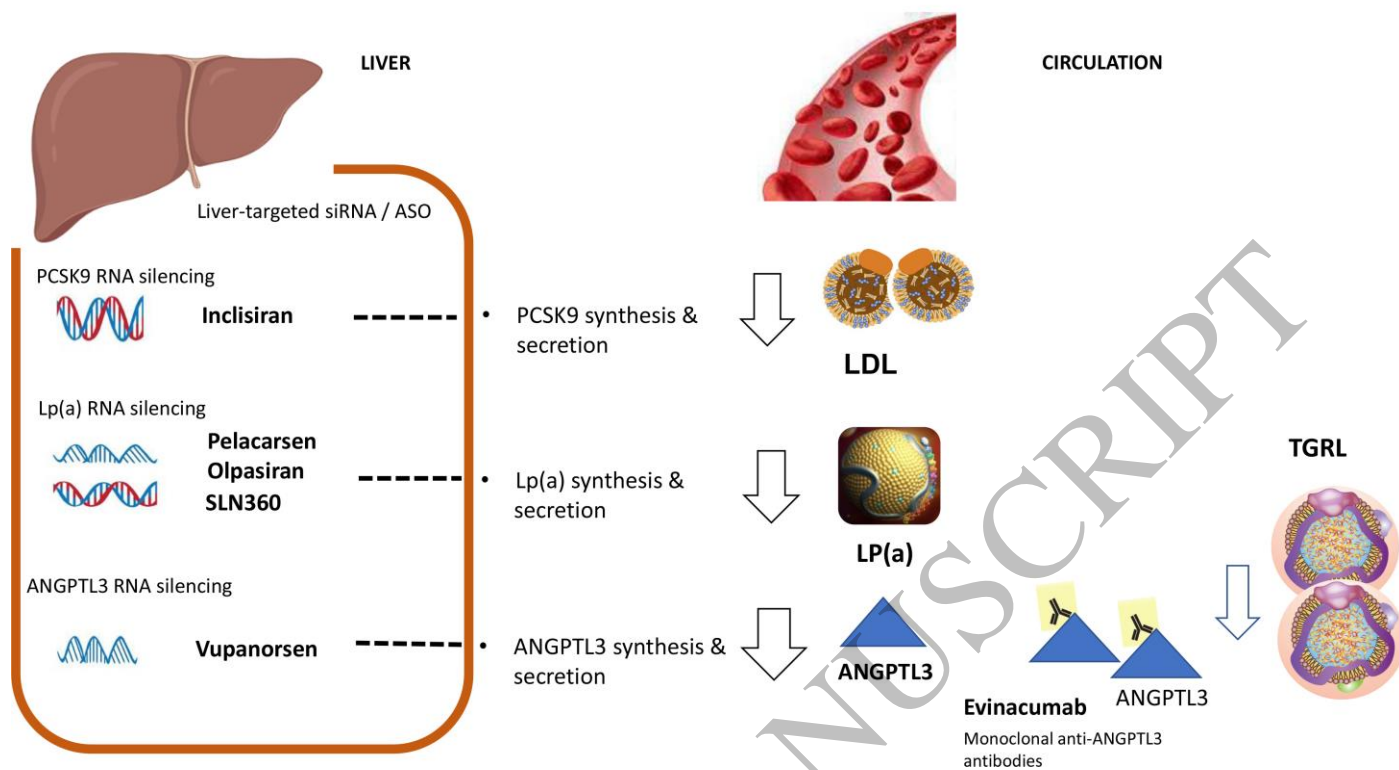


Figure 3
185x107 mm (x DPI)

1
2
3
4

Figure 4. Source of EVs affects their function.

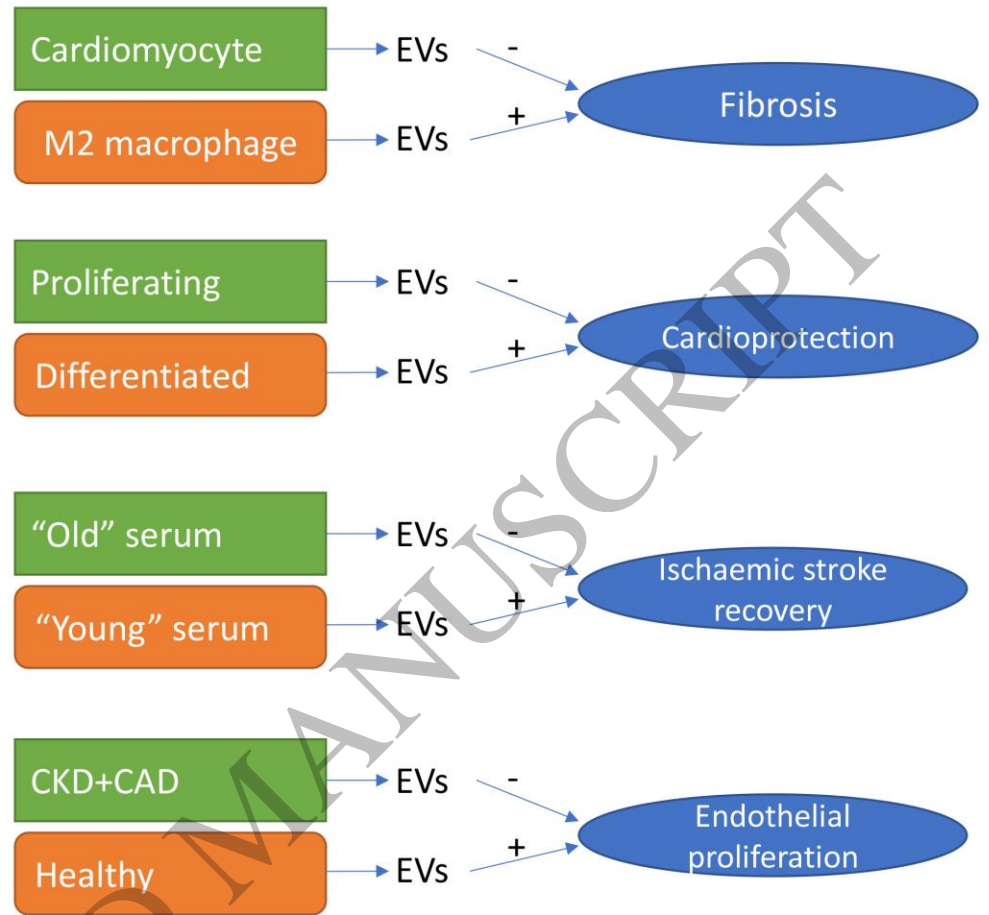


Figure 4
185x135 mm (x DPI)

1
2
3
4

Figure 5. Novel mechanisms of arrhythmia.

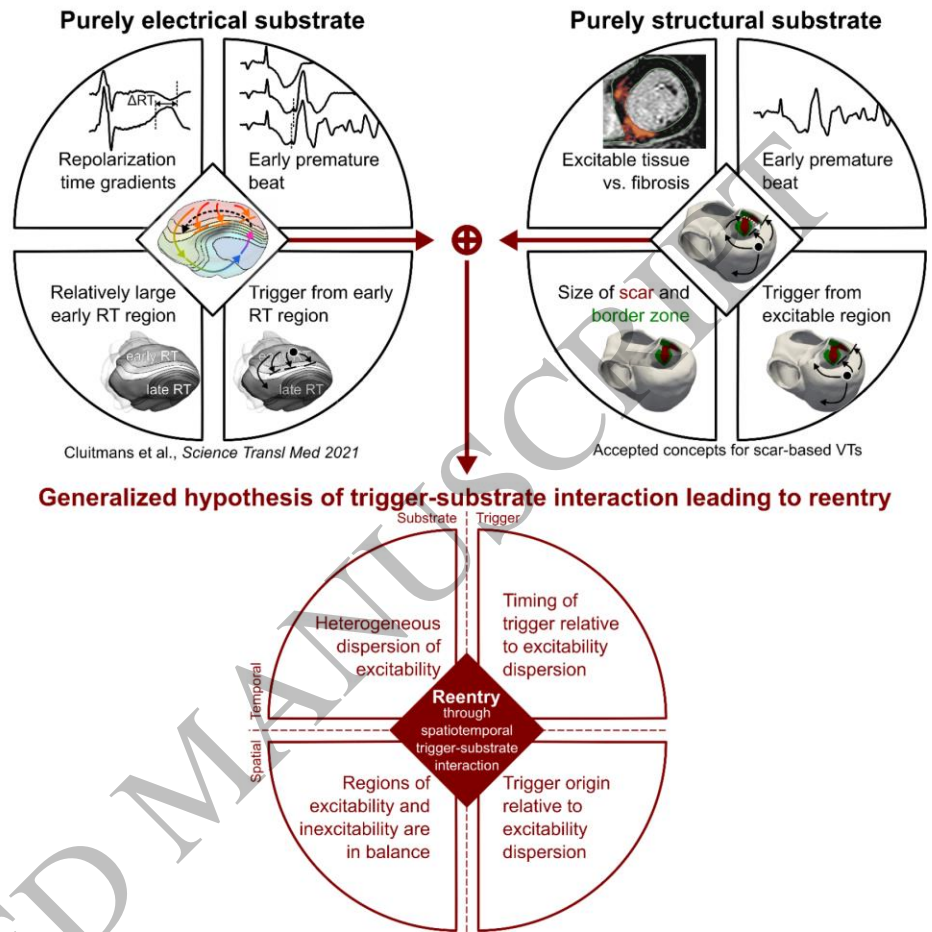


Figure 5
185x143 mm (x DPI)

1
2
3