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Sustainable Biodiesel Production by Transesterification of Waste Cooking Oil and Recycling of Wastewater Rich in Glycerol as a Feed to Microalgae

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Abstract: The amount of solid and liquid organic waste and wastewater is continuously increasing all over the world. The necessity of their reuse and recycling is, therefore, becoming more and more pressing. Furthermore, the limited fossil fuel resources, in conjunction with the need to reduce greenhouse gas emissions, advocate the production of renewable fuels. In this work, we analyze a sustainable second-generation process to produce biodiesel by transesterification of waste cooking oil, coupled with a third-generation process in cascade for recycling the incoming wastewater. Since this latter is rich in glycerol, it is used as a feed for microalgae, from which oil can be extracted and added to the waste cooking oil to further produce biodiesel and close the cycle. We studied the influence of different factors like temperature, catalyst load, and reactants ratio on the kinetics of transesterification of the waste oil and estimated the kinetic parameters by different kinetic schemes. The obtained values of activation energies and pre-exponential factors at chosen conditions of $T = 60\text{ }^{\circ}\text{C}$ and catalyst load of 0.6% w/w in methanol are: $E_{a,\text{direct}} = 35,661\text{ J mol}^{-1}$, $E_{a,\text{reverse}} = 72,989\text{ J mol}^{-1}$, $k_{0,\text{direct}} = 9.7708\text{ [dm}^3\text{ mol}^{-1}]^3\text{ min}^{-1}$, and $k_{0,\text{reverse}} = 24,810\text{ [dm}^3\text{ mol}^{-1}]^3\text{ min}^{-1}$ for the global fourth-order reversible reaction scheme and $E_a = 67,348\text{ J mol}^{-1}$ and $k_0 = 2.157 \times 10^9\text{ min}^{-1}$ for the simplified pseudo-first-order irreversible reaction scheme; both in strong agreement with literature data. Furthermore, we designed very efficient conditions for discontinuous and continuous operating mode, both at lab-scale and pilot-scale. The quality of the biodiesel produced from waste cooking sunflower oil is compared with that of biodiesel produced by different kinds of virgin vegetable oils, showing that the former possesses acceptable quality standards (Cetane number = 48 and LHV = 36,600 kJ kg⁻¹). Finally, the recycling of wastewater rich in glycerol as a nutrient for mixotrophic microalgae nurturing is discussed, and microalgae growing kinetics are evaluated (k_1 about 0.5 day⁻¹), endorsing the possibility of algae extraction each 4–5 days in a semi-continuous operating mode. The experimental results at the pilot scale finally confirm the quality of biodiesel, and the obtained yields for a two-stage process prove the competitiveness of this sustainable process on the global market.



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1. Introduction

Fossil fuels are non-renewable resources, and consequently, they will last for a limited period of time. The bounded fossil fuel resources, along with the need to reduce Green House Gas (GHG) emissions, were in the past a major impulse to the development of alternative fuels, that can be partially or totally derived by organic waste. Historically, political turmoil in the Middle East or natural disasters (such as Hurricane Katrina) have resulted in shortages of petroleum products and accompanying spikes in prices. Even if political and production issues could be remedied, recent estimates suggest that the current rates of consumption will result in the depletion of oil and natural gas reserves in roughly

50 years and less than a century and a quarter for the coal ones [1]. As suggested by the demise of the peak oil theory by Campbell and Laherrère [2], such estimates have to be used only as static references, since the capacity to extract fossil fuels varies continuously over time and the consumption levels may rise and fall. Encouraging signs are provided by the plateauing of the world coal production in 2013, while crude oil should have reached peak demand in 2019 [3].

In 2015, the involved Parties pledged to restrict global warming, committing to put in the efforts for limiting the temperature increase to 1.5 °C and maximum 2 °C with respect to pre-industrial times [3]. As carbon dioxide (CO₂) is the main contributor to GHGs in terms of quantity, the focus is now on both power generation and transport sector. Since 1990, the emissions linked to the transport sector have increased by more than 50%, earning the primacy for the fastest growing CO₂ source and weighing for 25% of the total emissions of carbon dioxide from the combustion of fuels [4]. As a result, increased attention has been given to biofuels, such as biodiesel, that can be used as an alternative fuel in compression–ignition engines. The use of biofuels, along with the more widespread adoption of electric vehicles, embody the pathway for the necessary paradigm shift needed to bring sustainability to the transport industry; furthermore, in sectors like marine transport, aviation, and heavy freight, biofuels are currently seen as the only realistic and practical alternative to fossil fuels [5].

The production of biodiesel from renewable resources, such as vegetable oils and animal fats, makes it biodegradable and non-toxic. Moreover, it contributes to the reduction of CO₂ emissions, because it comprises a closed carbon cycle [6]. Vegetable oils can be considered a good alternative fuel, because of their renewable nature and environmental benefits [7].

Despite all the advantages, the use of vegetable oils as a fuel inevitably has some drawbacks. The direct use in internal combustion engines is problematic because vegetable oils have a higher viscosity than diesel fuel and lower volatility, so they do not burn completely and form deposits in the fuel injectors of diesel engines. According to the literature, there are five ways to reduce these problems: the blending of vegetable oil and diesel, thermal cracking (pyrolysis), micro-emulsions, esterification, and transesterification [8]. Esterification and transesterification reactions are the most favored reaction pathways to produce biodiesel [9].

Biodiesel is a highly biodegradable fuel in water as well as in soil and a great part of it, under aerobic or anaerobic conditions, is mineralized in 28 days at most [10]. It is also a carbon-free fuel, as the plants that serve as raw material for its production absorb more carbon than that which is released during the burning of this biofuel [11]. Moreover, when biodiesel is burned in diesel engines, the emissions of hydrocarbons, carbon monoxide, particulate matter, and sulfur dioxide are reduced except for nitrogen oxides, the emission of which increases due to the oxygen content of biodiesel [12].

Studies of diesel engines powered by biodiesel published by the Health and Safety Institute shows that fine particulate (PM₁₀) is reduced by 58% with a decrease of 76% of the carbonaceous moiety (soot), which is its most harmful one since not only it is the more absorbable during breathing but it also cannot be traced by catalytic filters. Carbon monoxide is reduced by 58% as well at high loads and aromatic compounds undergo a decrease of 68%, thus reducing the risk of cancer, whilst there were no significant changes as regards the other unregulated pollutants.

The production and trade of biofuels have increased in recent years, to meet global demand for renewable fuels. The major participants in the liquid biofuels trade (ethanol more than biodiesel) are Brazil, China, the USA, India, and the European Union (EU) with France and Germany in the lead [5,13].

To increase the biodiesel trade, the governments promulgated different promoting policies and incentives. Among the EU's policies, the Renewable Energy Directive (RED), which caused a high demand for biodiesel imports in the EU, its 2018 evolution, the RED II and the EU's Fuel Quality Directive, stating the requirements and quality standards for

renewable fuel use, can be cited [14]. RED requirements also define the sources of biomass that are considered sustainable (direct/indirect land use in producing biofuels). Current legislation in the EU also proposes limits on transportation energy that can originate from crop-based biofuels and restrictions on these limits are increasing: the RED II introduces a cap on conventional biofuels gradually diminishing to 3.8% in 2030 from the 2021 limit of 7% and a minimum share of 6.8% by 2030 of advanced biofuels increasing from the 1.5% specified for 2021 [15].

For all these reasons it is necessary to move to other sustainable biomass for biofuels production, not in competition with other resources, and a possible solution is to use organic waste. Since biodiesel is made by simple alkyl monoesters of long-chain fatty acids, it can be produced from waste vegetable oils.

Vegetable oils, including waste oils from cooking, generally referred to as Straight Vegetable Oils (SVO) can be burned directly as a fossil diesel substitute for energy production but due to their high viscosity, their use leads to poor efficiency in combustion, incomplete combustion, carbonization on rings and injectors, and poor atomization of the biofuel. For all these reasons the transesterification process is used, allowing the transformation of the initial triglyceride (oil) into esters (biodiesel) and glycerol (byproduct), using alcohol as reactant plus a catalyst, usually a strong alkaline like potassium or sodium hydroxide. The quality of the produced biodiesel depends on the quality of the initial vegetable oil, i.e., on the fatty acids attached to the glycerol and composing the triglyceride, and on the impurities generated during cooking, if waste frying oils are used.

As second-generation processes are used for the production of biofuels, new issues related to their sustainability hide when any by-products or waste generated cannot easily be reused or recycled. Currently, glycerol occupies an almost saturated market, a fact resulting in a continued decline of its price and thus directly affecting the biodiesel production cost. This trend will not cease as more biodiesel production facilities begin production [16] unless other ways of reusing this by-product are found. Among them, pyrolysis, gasification, reforming, photocatalysis, and bioconversion are the most investigated ones.

Wastewater coming out of the biodiesel washing process still contains glycerol and methanol; one possibility of recycling them consists in using them as feed to photobioreactors for the cultivation of microalgae, from which oil can be extracted and exploited yet again for transesterification.

Different strains of microalgae, including *Chlorella vulgaris*, have shown high tolerance to wastewater with high nutrient concentration [17]. Wastewater containing glycerol was used in heterotrophic nurturing of *Chlorella vulgaris* [18,19], resulting in high yields [20]. Paladino and Neviani [21] conducted experimental activity on *Chlorella v.* using synthetic wastewater with the addition of glycerol at concentrations of 4–15 mg L⁻¹ and flue-gas, obtaining interesting growing kinetics, such as to suggest the integration with the biodiesel production process.

In this work, we propose a sustainable process to produce biodiesel by transesterification of waste cooking oil (second generation), in which wastewater in exit from the transesterification stage is completely recycled and used as a feed for microalgae, from which oil is extracted and added to the waste cooking oil to produce again biodiesel (third generation), closing the cycle. We compared the quality of the biodiesel produced from waste cooking oil with that of biodiesel produced by different kinds of virgin vegetable oils, showing that the former can possess high-quality standards, comparable to those of biodiesel produced by virgin oils. We also estimated the kinetics of our production process and the best operating conditions in discontinuous and continuous operating modes. Finally, the recycling of wastewater rich in glycerol as a nutrient for mixotrophic microalgae nurturing is discussed and the estimated microalgae growing conditions are reported.

These represent key data, necessary to gauge the competitiveness of this biodiesel production process on the global market.

2. Materials and Methods

2.1. Transesterification of Vegetable Oils

Transesterification of vegetable oils consists of a set of organic reactions where a triglyceride (ester, TG) is transformed into another ester by reacting with alcohol (alcoholysis of carboxylic esters [22]) and in presence of a catalyst, usually a strong acid or base. The final product is a mixture of fatty acids mono alkyl esters (FAME) plus glycerol (G) [23]. Transesterification is an equilibrium reaction and the transformation occurs essentially by mixing the reactants and the catalyst. The overall process can be considered as a sequence of three consecutive and reversible reactions, in which mono-glycerides (MG) and di-glycerides (DG) are formed as intermediates [24]. A general reaction scheme using methanol (M) is reported in Figure 1. The global reaction scheme (Figure 2) requires a 1:3 TG:M molar ratio; however, an excess of alcohol is required to increase yields.

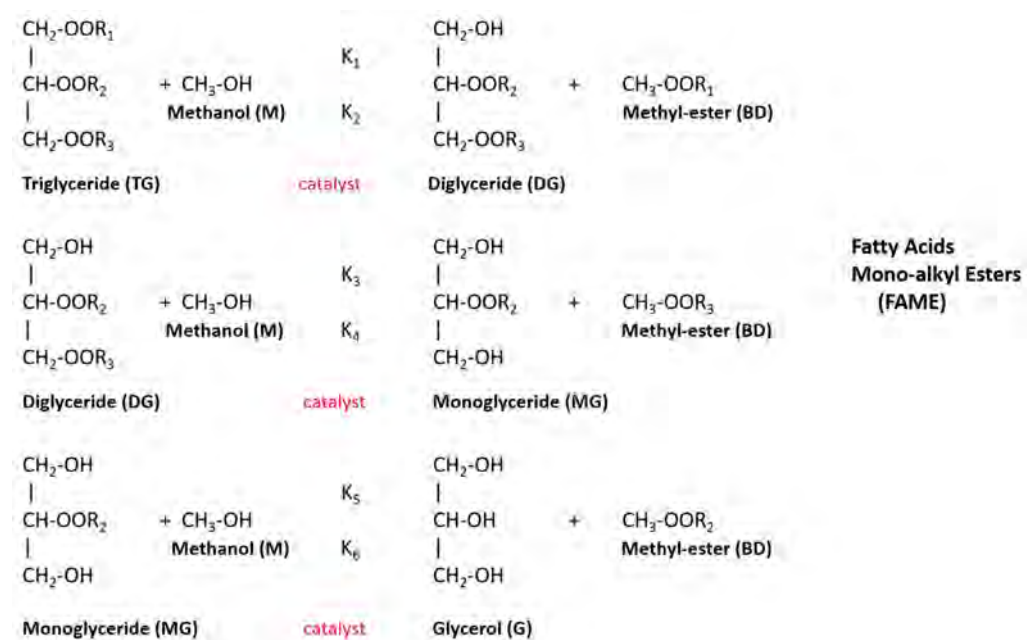


Figure 1. Transesterification of vegetable oils: Series reaction scheme.

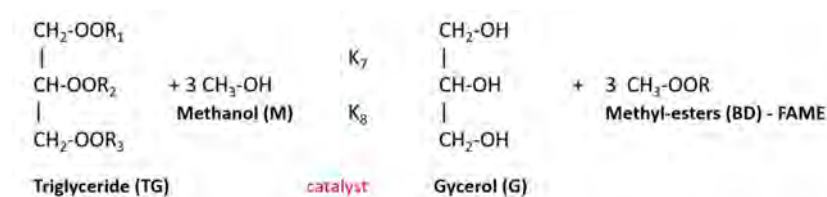


Figure 2. Transesterification of vegetable oils: Global reaction scheme.

The type of catalyst (acid or alkaline), waste oil/alcohol molar ratio, temperature, purity of the reactants (mainly water content), and initial free fatty acid content are the main factors influencing the transesterification process.

Acid-catalyzed transesterification of vegetable oils uses Brønsted acids, preferably sulfonic and sulfuric acids [25]. Even if yields can reach high values, reaction time is still very high also at a temperature greater than 373 K [26].

Even if excess alcohol increases biodiesel formation, on the other hand, it makes it difficult to separate glycerol from the desired product. Consequently, the TG to alcohol ratio must be optimized, depending on the other factors. Acid-catalyzed transesterification should be carried out in the relative absence of water, to avoid the competitive formation of carboxylic acids which reduce the yields in FAME.

Alkali-catalyzed transesterification of vegetable oils uses alkaline metal alkoxides [27] such as CH_3ONa , with yields reaching 98% and half an hour reaction time, and alkaline metal hydroxides [28] such as KOH and NaOH, as well as sodium or potassium carbonates [29]. The kinetics are faster than the corresponding acid-catalyzed ones and alkaline catalysts are usually less corrosive than acid-catalysts. In addition, alkali-catalyzed transesterification requires a relative absence of water in the reactants. Anyway, water is produced by the reaction of the hydroxide with alcohol and its presence gives rise to hydrolysis of some of the produced FAME, with consequent soap formation. This undesirable saponification reaction reduces the ester yields and makes considerably more difficult the recovery of the glycerol due to the formation of emulsions [26]. Soap formation is reduced by the use of potassium carbonate at a molar fraction of 2–3% [29], due to the formation of bicarbonate instead of water, which does not hydrolyze esters.

Untreated biodiesel contains impurities such as free glycerol, soaps, metals, free fatty acids, catalysts, water, and glycerides. The high impurities level can reduce the engine life. Table 1 shows each impurity effect.

Table 1. Effects of impurities on biodiesel and engines.

Impurity	Effect
Water	Oxidation, corrosion, and bacteriological growth (filter blockage).
Glycerol	Deposits in the injectors (carbon residue), high viscosity, crystallization, settling problems.
Methanol	Low values of density and viscosity, low flash point (transport, storage, and usage problems), corrosion of Al and Zn.
Soap	Deposits in the injectors (carbon residue), filter blockage (sulfated ashes), engine weakening.

There are generally two accepted methods for purifying biodiesel: wet and dry. The most traditional one is wet cleaning and it is widely used to remove contaminants as alcohol in excess, catalyst, and soap [30]. However, the water-washing process has several disadvantages, such as yield loss in the effluent, and high soap levels that cause emulsification [31]. The emulsion can occur when the reaction is not complete and residual triglycerides remain inside the mixture allowing the bonding of glycerine molecules to the esters portion. These tend to be not very soluble in water and thicken up, thus creating a creamy-like textured emulsion.

After washing, biodiesel is dried by vacuum flash drying or it can dry when left into settling tanks [32].

2.2. Transesterification of Waste-Cooking Oils

The proposed process at the pilot scale is a two-stage sodium hydroxide-catalyzed transesterification. The waste cooking oil (fried sunflower oil) was collected by the Campus canteen. The other three uncooked oils were used for comparison to produce biodiesel with the same process: sunflower oil, olive oil, and rapeseed oil.

Previous experimental tests using both methanol and ethanol suggested the former as the most promising reactant in terms of yields in biodiesel, so the proposed study was carried out with methanol. Experimental tests were performed by following the classical scale-up steps for studying catalyzed reactions. They consist in:

1. Study of the reaction rate at lab scale in discontinuous operating mode, to find the main factors influencing kinetics, compute conversion and yields, estimate kinetic parameters, and choose the optimal values of the main factors (reactants ratio, catalyst load, temperature) influencing the process;
2. Verification of the quality of products and by-products, analysis of the produced waste and wastewater, identification and test of possible processes to recycle and/or reuse by-products and waste;

3. Experimental tests at pilot-scale in discontinuous operating mode, by considering all the other phenomena influencing kinetics, as heat and mass transfer;
4. Experimental tests and simulation at pilot-scale in continuous operating mode, to optimize residence times and yields.

Regarding step 4, only simulations were performed and results will be presented elsewhere.

2.2.1. Lab-Scale

The mixing and pre-heating of the chosen quantities of alcohol and granular NaOH catalyst were carried out in a flat bottom flask connected to a bulb (Allihn) condenser, and mixed by magnetic stirring until a perfectly homogeneous blend in the liquid phase was obtained. A conical flask (Erlenmeyer Pyrex®) containing oil was posed on the hotplate of another magnetic stirrer and heated until the temperature, constantly measured with a calorimetric sensor, reached the chosen process value; then the mixture of methanol plus catalyst was slowly added to the heated oil and mixed for the chosen reaction time (maximum time equal to 1 h). Samples were collected at different step-times. At the chosen final time the mixture was sent to a separation funnel (decanter, with residence time equal to 5 h) (see Figure 3).



Figure 3. Lab-scale biodiesel production. Waste oil (**left**), catalyst and alcohol (**center**), decanter (**right**).

Purification of the biodiesel was carried out by a bubble washing technique. At first, the pH of biodiesel extracted from the decanter was measured. Then the procedure of bubble washing started by slowly adding dilute acetic acid to neutralize the pH of the ester so as to prevent emulsion formation. The apparatus used for the purification process at lab-scale was an aquarium air pump, responsible for creating bubbles at a rate of about 10 bubbles per second through a sponge-diffuser, plunged in the flat-bottomed conical flask (or separation funnel) containing biodiesel and water/acetic acid solution.

Since the water solution density is higher than that of biodiesel, the water stays at the bottom of the washing tank, where the bubbles sprout out from the diffuser. Bubbles rise, first crossing water and then biodiesel. Every bubble remains covered by a thin film of water and raises passing through biodiesel, washing it on the way. When the bubble comes to the surface it explodes and leaves a small drop of water that returns to the bottom, crossing again the biodiesel and washing it again [33].

2.2.2. Pilot-Scale

Waste oil was pre-treated by a full-flow oil filter. Even if oil pretreatment is very important in base catalyzed transesterification (being this process highly sensitive to initial FFAs composition), expensive pretreatment processes such as distillation or acid pre-esterification are not suitable in this context. Moreover, the possibility of adding the oil extracted from the microalgae to the waste oil will produce a blend of high FFA feedstock with a lower one, so reducing the possibility of initial saponification reactions.

We adopted two stainless steel stirred reactors (AISI 304) of volume $V = 15$ L at low mixing velocity (60–120 rpm) for the first- and second-stage transesterification process; one stainless steel mixer (AISI 304) of $V = 5$ L at high stirring velocity (1200 rpm) to prepare the catalyst/methanol blend and two stainless steel decanters of $V = 20$ L (see Figure 4). Velp Scientifica PW overhead stirrers were used. The heating system of the transesterification reactors was made by resistive wires, electrically insulated from external tubular sheaths (mineral insulation by magnesium oxide) surrounded by insulating material wrapped around the reactor. The tubular sheaths had excellent thermal conductivity, in order to assure high heat transfer to the reactor wall.



Figure 4. Pilot-scale biodiesel production. Mixer (left), transesterification reactor (center), decanter (right).

The products in exit from the first TE reactor were sent to the first decanter while the final product in exit from the second stage was purified by bubble washing inside the second decanter, operated as washing/separation unit. pH was adjusted by manipulating a Gilson Minipuls 3 peristaltic pump (through the PXI station) to dose dilute acetic acid.

2.3. Wastewater Treatment by Microalgae

At lab-scale, microalgae of the *Chlorella vulgaris* species were nurtured in glass stirred tank reactors of $V = 2.5$ L, and internal diameter of 0.120 m, mixed by Velp Scientifica PW overhead stirrers, at 100 rpm (radial impeller with a diameter equal to 0.080 m). The reactors had clamped tops with three nozzles for probes introduction: pH, temperature

(T), conductivity (C), and dissolved oxygen (DO) were online acquired. The operation could be managed in fed-batch and wastewater in exit from transesterification was inserted by controlled peristaltic pumps (Gilson Minipuls 3) so as microalgae extraction could be cyclically operated.

At the pilot scale, we used external loop airlift reactors made by polymethyl methacrylate (PMMA). Each reactor had a total volume of 10.5 L, with the downcomer diameter equal to 0.110 m and the riser diameter equal to 0.050 m. Collectors (diameter = 0.050 m) were made of poly-vinyl-chloride. Probes for pH, T , C , and DO measurements were inserted in the midsection of the lower collector and online measures were managed by MEMO HQ Scada system [34]. Air was sparged at the bottom of the riser by a stainless steel diffuser. Inlet wastewater flowrate and extracted microalgae flowrate were controlled and it was possible to operate in both fed-batch and semi-continuous mode.

Details about photobioreactors scale-up can be found in Paladino and Neviani [35].

In the present paper, we only report results on the capability of the microalgae to exploit wastewater in exit from the transesterification stage to grow in mixotrophic conditions and on their growing kinetics. Other insights about the optimal operating conditions inside the photobioreactors can be found in [21].

2.4. Instrumentation

2.4.1. Transesterification of Waste Oil

At the lab scale, samples were collected every five minutes during the batch transesterification reaction and analyzed to estimate kinetics. At the pilot scale, only the final product in exit from the first and the second stages at the designed residence time was analyzed.

Samples were analyzed by Trace-Ultra GC (Agilent 19091S-433 column) and TSQ 8000 MS by Thermo Fischer Scientific following EU EN 14214 (test methods EN 14103, EN 14105 for FAMES determination, EN ISO 5165 for cetane number; specifics and conditions can be found in the Supplementary Material, Table S1). Before proceeding with the analysis, samples were prepared by operating cold methylation, weighing 400 mg sample + 6 mL hexane + 0.25 mL of methanolic KOH. This operation guarantees that all the peaks are methyl esters.

Low Heating Value was measured by a semi-automatic bomb calorimeter (mod C200 Basic, IKA).

The research laboratory is quality certified UNI EN ISO 9001: 2008 and Environmental quality certified UNI EN ISO 14001: 2015. The analytical chemistry laboratory where samples are analyzed is certified UNI CEI EN ISO/IEC 17025:2018 and instrumentation is managed under UNI EN ISO 10012:2004.

2.4.2. Wastewater Recycling in Microalgae Cultivation

Online measures of the main parameters inside the reactors were acquired by B&C Electronics series 7685 Microcontrollers and related probes. Flowrates were manipulated by the PXI National Instrument system plus Labview. Gilson Minipuls 3 and Welko WP1100 peristaltic pumps were respectively used at the lab and pilot scale.

The microalgae concentration was offline daily acquired by measuring the absorbance of samples at different wavelengths (430, 480, 650, and 680 nm). Details about calibration curves can be found in [35]. A Hach DR-6000 UV-VIS spectrophotometer was used, while the wastewater composition was measured by both GC and UV-VIS spectrophotometers.

3. Results

3.1. Experiments at the Lab Scale: Kinetics

The experimental campaign at the lab scale was designed by considering four factors at different levels.

- Factor N.1: type of oils, at four levels (waste sunflower oil, uncooked sunflower, olive, and rapeseed oil);
- Factor N.2: molar reactants ratio, at three levels (oil/alcohol = 1:3, 1:5, 1:7);
- Factor N.3: temperature, at four levels (20 °C, 50 °C, 60 °C, 70 °C);

- Factor N.4: catalyst load at three levels (0.3%, 0.6%, 1.2% *w/w* in methanol).

Some runs were eliminated due to expected lack of information on kinetics and a total of 78 runs were carried out.

Mixing is another important factor. For this reaction a slow reaction rate at initial times was observed, followed by a sudden increase for about 40–60 min, followed again by a rate decrease as the reaction approaches equilibrium. At the initial time, a two-phase L-L system is present, with a diffusion-controlled mechanism; as time passes, the formed methyl esters act as solvents for the remaining reactants and the system can be considered as a homogeneous liquid phase under a kinetic-controlled mechanism. A good mixing reduces the period of mass transfer control to less than one minute. All the experiments were conducted at Reynolds number *Re* greater than 8000, and this value was kept during the scale-up.

The concentration of biodiesel was measured every 5 min in order to estimate the kinetic constants. Tables 2 and 3 show some of the collected data.

Table 2. Methyl-esters (BD) concentration (mol dm⁻³) versus time at different molar reactants ratio, fixed temperature (*T* = 60 °C), and fixed catalyst load (0.6% *w/w*).

TG:M	5 min	10 min	15 min	20 min	30 min	40 min	60 min
1:3	0.35	0.45	0.51	0.52	0.55	0.58	0.64
1:5	1.50	1.72	1.85	1.92	1.97	1.98	2.01
1:7	1.80	1.95	2.00	2.10	2.15	2.15	2.16

Table 3. Methyl-esters (BD) concentration (mol dm⁻³) versus time at different temperatures, fixed molar reactants ratio (TG:M = 1:5), and fixed catalyst load (0.6% *w/w*).

<i>T</i>	5 min	10 min	15 min	20 min	30 min	40 min	60 min
20 °C	0.15	0.21	0.35	0.50	0.65	0.80	1.15
50 °C	1.35	1.60	1.80	1.88	1.92	1.98	1.98
70 °C	1.90	2.08	2.10	2.15	2.15	2.16	2.16

By adopting the reaction scheme of Figure 1, and defining the generation/consumption term for each component *i* inside the reacting system as *r_i* (mol dm⁻³ s⁻¹), the following molar balances can be written for the discontinuous reactor at the lab scale:

$$\frac{d[\text{TG}]}{dt} = r_{\text{TG}} = -k_1[\text{TG}] \cdot [\text{M}] + k_2[\text{DG}] \cdot [\text{BD}] \quad (1)$$

$$\frac{d[\text{M}]}{dt} = r_{\text{M}} = -k_1[\text{TG}] \cdot [\text{M}] + k_2[\text{DG}] \cdot [\text{BD}] - k_3[\text{DG}] \cdot [\text{M}] + k_4[\text{MG}] \cdot [\text{BD}] - k_5[\text{MG}] \cdot [\text{M}] + k_6[\text{G}] \cdot [\text{BD}] \quad (2)$$

$$\frac{d[\text{DG}]}{dt} = r_{\text{DG}} = k_1[\text{TG}] \cdot [\text{M}] - k_2[\text{DG}] \cdot [\text{BD}] - k_3[\text{DG}] \cdot [\text{M}] + k_4[\text{MG}] \cdot [\text{BD}] \quad (3)$$

$$\frac{d[\text{MG}]}{dt} = r_{\text{MG}} = k_3[\text{DG}] \cdot [\text{M}] - k_4[\text{MG}] \cdot [\text{BD}] - k_5[\text{MG}] \cdot [\text{M}] + k_6[\text{G}] \cdot [\text{BD}] \quad (4)$$

$$\frac{d[\text{G}]}{dt} = r_{\text{G}} = k_5[\text{MG}] \cdot [\text{M}] - k_6[\text{G}] \cdot [\text{BD}] \quad (5)$$

$$\frac{d[\text{BD}]}{dt} = r_{\text{BD}} = k_1[\text{TG}] \cdot [\text{M}] - k_2[\text{DG}] \cdot [\text{BD}] + k_3[\text{DG}] \cdot [\text{M}] - k_4[\text{MG}] \cdot [\text{BD}] + k_5[\text{MG}] \cdot [\text{M}] - k_6[\text{G}] \cdot [\text{BD}] \quad (6)$$

A first simulation was carried out by using the kinetic data reported in [36] for transesterification of soybean oil at operating temperature equal to 50 °C, and *Re* = 6200. The six unknown pre-exponential factors were computed by using the Arrhenius law (see Table 4) as

$$k_{0,i} = k_i \left(\exp \left(-\frac{E_{A,i}}{R T} \right) \right)^{-1} \quad (7)$$

Table 4. Kinetic values used for simulation.

Kinetic Constant [dm ³ mol ⁻¹ min ⁻¹]		Activation Energy [J mol ⁻¹]		Pre-Exponential Factor [dm ³ mol ⁻¹ min ⁻¹]	
k_1	0.050	$E_{a,1}$	55,000	$k_{0,1}$	1.9×10^7
k_2	0.110	$E_{a,2}$	41,556	$k_{0,2}$	3.35×10^5
k_3	0.215	$E_{a,3}$	83,094	$k_{0,3}$	1.99×10^{12}
k_4	1.228	$E_{a,4}$	61,250	$k_{0,4}$	4.43×10^9
k_5	0.242	$E_{a,5}$	26,866	$k_{0,5}$	3.77×10^3
k_6	0.007	$E_{a,6}$	44,116	$k_{0,6}$	5.36×10^4

By looking at Table 4, since $E_{a,1} > E_{a,2}$ and $E_{a,3} > E_{a,4}$, the first two reactions of the scheme reported in Figure 1 should be favored by high temperatures. By considering the pre-exponential factors $k_{0,i}$ constant with temperature, the kinetic constants at every level of the factor temperature in our experiments are calculated.

Other literature data were taken into consideration, as those reported in [37] for the transesterification of sunflower oil with ethanol and NaOH as the catalyst and those presented in [38] for sunflower methanolysis catalyzed by potassium hydroxide but parameters of [30] showed the best fit with our experimental data.

Simulations were carried out by recalling that vegetable oils are composed of triglycerides for about 96–98%, and, since seed oils are mainly composed of oleic and linoleic acid, we supposed that the waste oil is only made by the ester between glycerol and oleic acid (glyceryl trioleate). Under this hypothesis (and supposing no degradation of waste oil during transesterification), it was possible to determine the initial number of moles of TG into the reactor, by considering the TG molar weight equal to 884 g mol⁻¹ and its mean density around 910 g dm⁻³. It was then possible to simulate the reaction inside the batch reactor at lab scale, in isothermal conditions, for all the different runs of the experimental campaign.

Some results are reported in Figures 5 and 6.

By looking at Figure 5, it can be observed that the increase in BD production is non-linear with the excess methanol and at molar ratios 1:10 almost all the TG is anyway reacted after 20 min (being $[TG]_0 = 0.867$ mol dm⁻³, the maximum theoretical $[BD]_f = 2.601$ mol dm⁻³). The chosen operating condition is molar ratio TG:M = 1:5. By observing Figure 6, an operating temperature of 60 °C seems to assure a valid final biodiesel concentration for reacting times of about 30–40 min, paying attention that good mixing is assured. By taking into account energy savings and safety conditions (and excluding operating temperatures near the boiling temperature of methanol), this operating temperature seems to be a good choice for reacting times near to 40 min. Figure 6b shows the simulated reaction at molar TG:M = 1:5, $T = 60$ °C and error bars of the measured biodiesel concentration. In this case, at $[TG]_0 = 0.867$ mol dm³, the concentrations of waste oil and biodiesel after 30 min are respectively $[TG]_{30} = 0.149$ mol dm³ and $[BD]_{30} = 1.965$ mol dm³. This final condition corresponds to a conversion of TG equal to 84% and a yield of about 75%. Comparing these simulated data with the experimental data of Table 2 reporting a $[BD]_{30,exp} = 1.97$ mol dm³, it can be supposed that the kinetics parameters found by [36] for soybean oil at molar TG:M = 1:6 and $T = 50$ °C can be successfully extended to roughly predict the transesterification of the sunflower cooking oil.

Anyway, the next step of the study was to estimate the kinetic parameters from the collected experimental data. Ref. [24] proposed to use of a pseudo-first-order kinetic scheme when the process is carried out in high excess alcohol conditions, reserving the second-order kinetics scheme of Figure 1 (three-reactions scheme plus shunt-reaction scheme) at lower excess alcohol. We estimated kinetics by adopting both the global reversible reaction scheme reported in Figure 2 and its simplification to a pseudo-first-order irreversible reaction.

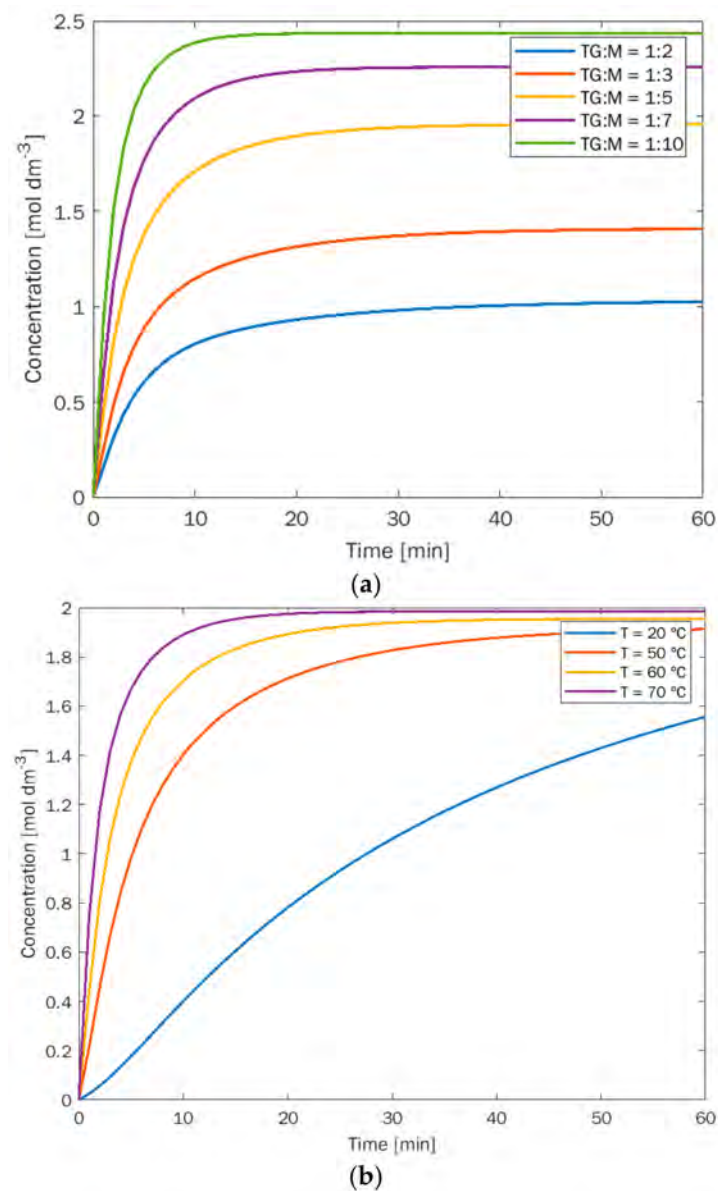


Figure 5. (a) Simulated biodiesel production from waste sunflower oil at changing reactants ratio, $T = 60\text{ }^{\circ}\text{C}$ and catalyst load = $0.6\% w/w$. (b) Simulated biodiesel production from waste sunflower oil at changing reaction temperature, TG:M = 1:5 molar and catalyst load = $0.6\% w/w$.

By considering the reaction scheme of Figure 2, we estimated the two constants k_7 and k_8 using the molar balances of the four components:

$$\frac{d[\text{TG}]}{dt} = -k_7 \cdot [\text{TG}] \cdot [\text{M}]^3 + k_8 [\text{G}] \cdot [\text{BD}]^3 \quad (8)$$

$$\frac{d[\text{M}]}{dt} = -3k_7 [\text{TG}] \cdot [\text{M}]^3 + 3k_8 [\text{G}] \cdot [\text{BD}]^3 \quad (9)$$

$$\frac{d[\text{G}]}{dt} = k_7 [\text{TG}] \cdot [\text{M}]^3 - k_8 [\text{G}] \times [\text{BD}]^3 \quad (10)$$

$$\frac{d[\text{BD}]}{dt} = 3k_7 [\text{TG}] \cdot [\text{M}]^3 - 3k_8 [\text{G}] \cdot [\text{BD}]^3 \quad (11)$$

to which two more equations (derived from experimental data) were added to solve the system. The former was the biodiesel concentration in time, obtained by fitting the measured biodiesel molar concentration at each fixed temperature with a time function of

the kind $[BD]_t = at^b$ (results are shown in Figure 7). The latter is the van't Hoff equation, used to compute the equilibrium constant at different temperatures from the enthalpy literature data. The system was solved numerically at different reactants ratios, for each tested temperature and at the different catalyst loads for all the tested oils. Arrhenius equation was finally used to estimate $E_{a,7}$, $E_{a,8}$, $k_{0,7}$, and $k_{0,8}$ from the calculated couples (k_7 ; k_8) at the different temperatures. The estimated values for the waste sunflower oil are $E_{a,7} = 35,661 \text{ J mol}^{-1}$, $E_{a,8} = 72,989 \text{ J mol}^{-1}$, $k_{0,7} = 9.7708 [\text{dm}^3 \text{ mol}^{-1}]^3 \text{ min}^{-1}$, and $k_{0,8} = 24,810 [\text{dm}^3 \text{ mol}^{-1}]^3 \text{ min}^{-1}$.

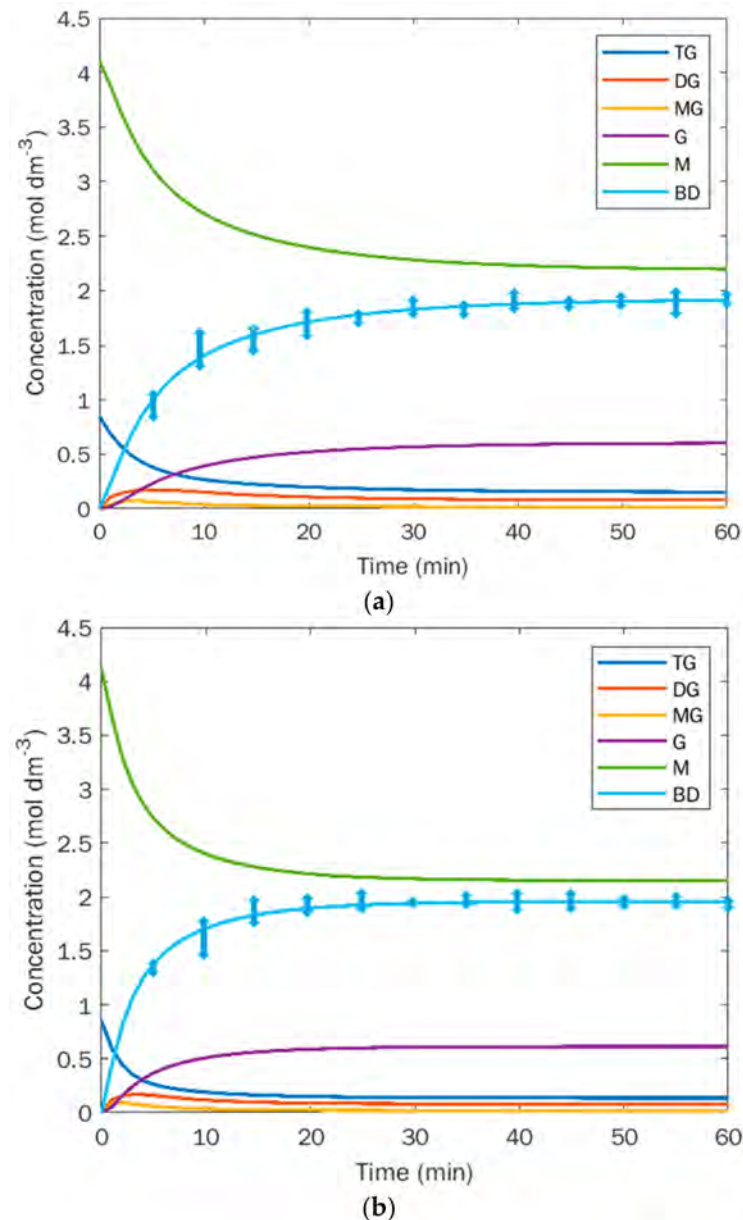


Figure 6. Simulated transesterification of waste sunflower oil at molar TG:M = 1:5 and catalyst load = 0.6% w/w. Comparison with experimental data at (a) $T = 50 \text{ }^\circ\text{C}$; (b) $T = 60 \text{ }^\circ\text{C}$.

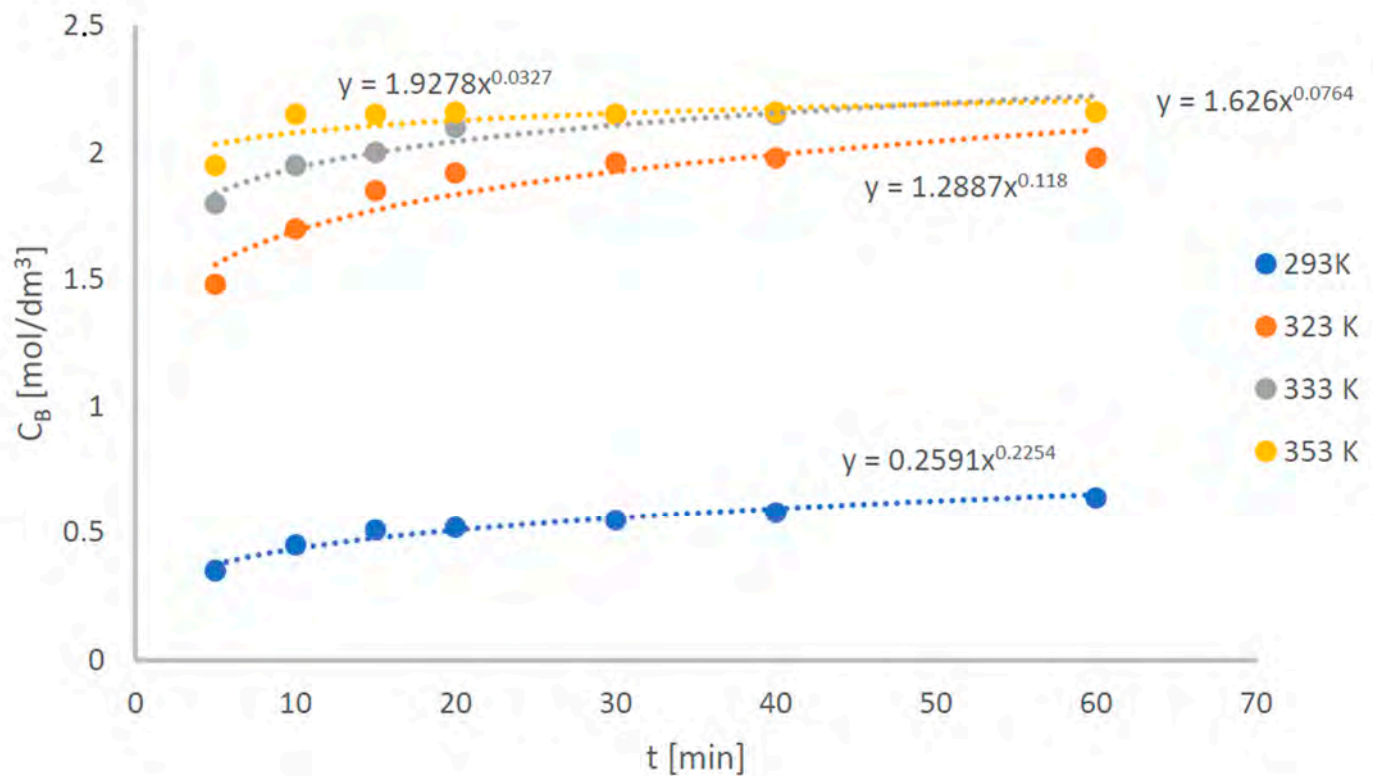


Figure 7. Biodiesel molar concentration curves estimated from experimental data at TG:M = 1:5 and different operating temperatures.

Since the hypothesis of irreversible reaction scheme seems to be in agreement with the experimental data at high excess methanol (see both Tables 2 and 3), by neglecting the backward reaction with constant k_8 , the TG molar balance can be written as

$$\frac{d[\text{TG}]}{dt} = -k_7[\text{TG}] \cdot [\text{M}]^3 \quad (12)$$

and, for molar TG:M greater than 1:5 (excess methanol), it can be supposed that the reaction rate mainly depends on the oil concentration. Being the remaining methanol concentration almost constant, the molar balance of TG reduces to

$$\frac{d[\text{TG}]}{dt} = -K[\text{TG}] \quad (13)$$

with $K = k_7[\text{M}]^3 = \text{const.}$

The analytical derivatives of the fitted biodiesel concentration in time (Figure 7) were calculated and directly used to compute K from the simplified system under the hypothesis of pseudo-first-order reaction rate for all the experimental runs.

The estimated parameter values for the sunflower waste oil with a high catalyst load are $E_a = 67,348 \text{ J mol}^{-1}$ and $k_0 = 2.157 \times 10^9 \text{ min}^{-1}$. The obtained activation energy is in agreement with [26], in which values in the range 33,600–84,000 J mol^{-1} are reported. The kinetics of virgin oils have also been estimated with the same approach and using the pseudo-first-order reaction scheme.

3.2. Biodiesel Production at the Pilot Scale

Biodiesel was produced at the pilot scale in discontinuous operating mode. Mean designed residence times of the main unit operations are 3, 50, 120, 50, 15, 120 min, respectively for methanol/NaOH mixing, first transesterification, first separation, second transesterification, bubble washing, and second separation. Chosen conditions were at

$T = 60\text{ }^{\circ}\text{C}$, molar reactants ratio (TG:M = 1:5) and catalyst load (1.2% w/w). Simulations were performed for continuous production using the estimated kinetics to design the correct unit volumes required. In Tables 5 and 6 the operating conditions and results at pilot-scale are summarized.

Table 5. Discontinuous transesterification of waste frying oil, main process data.

Waste Oil [kg]	Methanol [kg]	NaOH [kg]	Resid. Time in DSTRs [min]	Produced Biodiesel [kg]	Crude Glycerol [kg]
11	2	0.12	50	8.8–9	1.7–1.9
Wastewater to Microalgae [L]	Residual Glycerol [g L ⁻¹]	Residual Methanol [g L ⁻¹]	Residual NaOH [g L ⁻¹]	Soaps [g L ⁻¹]	Salts of Fatty Acids [g L ⁻¹]
1.2	4–12	1–2	3	1	2

Table 6. Continuous transesterification of waste frying oil, main process data.

Inlet Waste Oil [kg h ⁻¹]	Inlet Methanol [kg h ⁻¹]	Inlet NaOH [kg h ⁻¹]	Mean Resid. Time in CSTRs [min]	Inlet Oil Flow Rate [L h ⁻¹]
11	2	0.12	60	0.25
Produced Biodiesel [kg h ⁻¹]	Crude Glycerol [kg h ⁻¹]	Wastewater to Microalgae [L h ⁻¹]	Waste Oil Conversion 1st unit	Waste Oil Conversion 2nd unit
8.6–8.9	1.7–1.8	1–2	0.54	0.78

3.3. Biodiesel Quality Analysis

Biodiesel is defined by the notation BDxx (Bxx in the USA), where xx indicates the volume percentage of FAME content in the liquid. Pure biodiesel is generally not used as fuel being instead blended with petroleum diesel: common commercial biodiesel blends are BD2, BD5, and BD20 [39]. Biodiesel has to meet defined quality standards before it can be sold as fuel or blending stock. In Europe, the reference is EN 14214 standard, released by the European Committee of Standardization (CEN), while ASTM D6751 is valid in the US. In more detail, the specifications set the required properties as the Lower Heating Value (LHV), kinematic viscosity, density, Cetane number, and some important limits related to the biodiesel production process such as the maximum admitted residual glycerol, catalyst, and alcohol, the absence of free fatty acids (compliance with these characteristics is generally verified by GC-MS) or metals.

Generally, the principal factors influencing the quality of biodiesel obtained by transesterification are the quality of the added reactants and catalyst, the fatty acid composition of the parent vegetable oil or animal fat, and the post-production parameters.

In this work, we also investigated if other possible quality parameters of biodiesel, more than cetane number and LHV, can be defined to underlie differences between the biodiesels produced by virgin and waste sunflower oil.

3.3.1. FAME Distribution and Cetane Number

The results of the analysis are summarized in Table 7, in which the quantities of the most abundant FAMES inside the biodiesel produced are compared.

Table 7. Average cetane number (CN) of biodiesel produced from the different oils.

FAME Common Name	% Total	Average CN
Waste sunflower oil		
Linoleic Acid (di-unsaturated 9–12)	56.5–56.6%	39.0 ± 0.2
Oleic Acid (unsaturated)	28.2–28.4%	56.6 ± 0.2
Palmitic Acid (saturated)	7.4–7.6%	76.6 ± 0.3
Stearic Acid (saturated)	4.5–4.6%	85.9 ± 0.3
Tricosanoic Acid (saturated)	1.0–1.2%	71.6 ± 0.2
Total Cetane number		47.9 ± 0.1
Virgin sunflower oil		
Linoleic Acid (di-unsaturated 9–12)	58.3–58.5%	39.2 ± 0.1
Oleic Acid (unsaturated)	27.1–27.3%	56.6 ± 0.1
Palmitic Acid (saturated)	7.2–7.4%	76.6 ± 0.2
Stearic Acid (saturated)	4.4–4.6%	85.9 ± 0.3
Behenic acid (saturated)	1.0–1.1%	74.1 ± 0.1
Total Cetane number		47.8 ± 0.2
Olive oil		
Oleic Acid (unsaturated)	80.7–80.9%	56.6 ± 0.2
Palmitic Acid (saturated)	12.3–12.4%	76.6 ± 0.2
Stearic Acid (saturated)	3.6–3.8%	85.9 ± 0.3
Total Cetane number		58.4 ± 0.2
Rapeseed oil		
Oleic Acid (unsaturated)	78.4–78.5%	56.6 ± 0.2
Linoleic Acid (di-unsaturated 9–12)	8.4–8.6%	39.2 ± 0.3
Palmitic Acid (saturated)	5.0–5.2%	76.6 ± 0.3
Stearic Acid (saturated)	2.1–2.2%	85.9 ± 0.2
Eicosanoic acid (unsaturated)	1.7–1.8%	52.5 ± 0.2
Total Cetane number		53.4 ± 0.2

The percentage of FAME inside biodiesel depends on the initial composition of the oil, and vegetable oils characterized by a low content of residues of unsaturated fatty acids, such as oleic (monounsaturated) and linoleic (poly-unsaturated) acids, ensures ease of combustion and oxidative stability of the produced biodiesel. Moreover, the number and position of double bonds also influence the quality of biodiesel, since they contribute to destabilizing the fatty acid chain by reacting with oxygen forming hydro-peroxides. They attach to the fatty acid chain by degrading it and leading to chain polymerization, forming also insoluble gums.

By looking at Table 7, it can be observed that the biodiesel produced by the waste and the virgin sunflower oil contains about 85% of unsaturated fatty acids, and at very similar percentages. Since most of these methyl esters come from linoleic and oleic acid, this validates the hypothesis made for simulations that considers waste oil as pure oleic trioleate. Most probably, it could be also present a small additional quantity of linoleic acid that is not visible because of the peak of oleic acid which almost certainly encompasses it.

Cetane number (CN) is one of the most significant properties to specify the ignition quality of any fuel and, in particular, constitutes the prime indicator for diesel engines since it is related to the ignition delay time [40]. Various standards have been established worldwide for cetane number determination, with standard ISO 5165 by International Organization for Standardization (ISO). Value 100 of cetane number has been assigned to hexadecane $C_{16}H_{34}$, to represent the high-quality standard on the cetane scale [41]. Presuming that the CN of the methyl-esters mixture is a linear combination of the cetane numbers of the single FAMES [42] the average CNs are calculated from the results of the GC-MS analysis. CN is computed by considering the most abundant FAMES inside each produced biodiesel with a concentration greater than 1% weight). Results are reported in Table 7 (see the Supplementary Material, Figures S1–S4 and Tables S1–S9 for the complete

analysis of biodiesel produced in the chosen operating conditions of $T = 60\text{ }^{\circ}\text{C}$, TG:M = 1:5, catalyst load = 0.6% w/w).

3.3.2. Lower Heating Value

The lower heating values found for each biodiesel sample are reported in Table 8.

Table 8. LHV of the different types of biodiesel.

Biodiesel Source	LHV [kJ kg ⁻¹]
Olive oil	36,660 ± 120
Rapeseed oil	36,992 ± 110
Virgin sunflower oil	36,079 ± 110
Waste sunflower oil	36,557 ± 100

3.3.3. UV-VIS Analysis

Biodiesel produced from waste sunflower oil was compared to biodiesel produced from virgin sunflower oil, using the latter as a blank by spectrophotometric analysis. The observation of spectra in the visible length range shows no significant difference between biodiesel from fried sunflower and biodiesel from virgin sunflower oil, as reported in [43]. This fact lets us assume that cooking impurities do not remain inside biodiesel. On the contrary, some important differences are observed by comparing glycerol samples obtained by TE of waste and virgin sunflower oils. In this case, a peak was identified [43] (Appendix A, Figures A1 and A2), pointing out that most cooking residuals and other substances already present in waste oil are mostly dissolved in glycerol. This is probably due to the fact that glycerol is, albeit weakly, a polar compound. This is only a qualitative comparison analysis, the next step being a complete GC-MS analysis of crude glycerol samples.

3.4. The Recycling of Washing Wastewater

Waste-oil transesterification leads to the formation of glycerol, unprocessed methanol, catalyst, and small amounts of other chemicals. After bubble washing, crude glycerol is separated and destined for recycling or reuse after purification, while washing wastewater is usually sent to treatment units. In this work, we successfully used the produced wastewater rich in glycerol to feed *Chlorella vulgaris*, nurtured in photobioreactors in mixotrophic conditions.

At the lab scale, microalgae cultivation added with wastewater allowed to increase the biomass concentration inside the reactors of about 25% with respect to the control reactors in which microalgae were cultivated only in BBD medium [44]. The kinetic parameters of *Chlorella v.* growth were estimated by the model proposed in [29]:

$$c(t) = c_0 + \frac{k_a}{1 + \frac{k_a - c_0}{c_0} \exp(-k_1 t)} \quad (14)$$

where k_1 [d⁻¹] represents the specific rate constant, k_a [g L⁻¹ d⁻¹] is the maximum increase in the microalgae population, and c_0 [g L⁻¹] is the estimated initial microalgae concentration. In order to reduce errors due to microalgae acclimatization, concentration data collected in the first 24 h were neglected. Anyway, the experimental growing curve in fed-batch reactors at lab-scale partially contained a short lag-phase; for this reason, the initial concentration c_0 was estimated with appreciable uncertainty by the non-linear weighted least squares procedure, so as the values of k_1 . As a consequence, the values obtained for the determination coefficients were smaller than those calculated for the airlift reactors.

The obtained growing conditions are summarized in Table 9 for lab and pilot-scale.

Table 9. Kinetic parameters of growing conditions with wastewater (10% glycerol) and air enriched with 10% *v/v* carbon dioxide.

Operating Conditions	k_1 [d ⁻¹]	k_a [g L ⁻¹]	Average Biomass * [g L ⁻¹ d ⁻¹]	R ²
Outdoor, Fed-batch	Min: 0.382 ± 0.218	Min: 1.629 ± 0.073	233	0.742
	Max: 0.425 ± 0.253	Max: 1.775 ± 0.082	254	0.743
Outdoor, airlift	Min: 0.273 ± 0.066	Min: 1.493 ± 0.043	249	0.982
	Max: 0.326 ± 0.043	Max: 1.596 ± 0.050	266	0.953

* Biomass is extracted at day 7 in the fed-batch, and cyclically, every 6 days, in the airlift.

Standard deviations reported in Table 9 take into consideration the replicas and the results of the estimation procedure with the proposed model. Since we collected data at different wavelengths, and there is not an optimal wavelength to be used for indirectly measuring concentration when algae are fed with wastewater, we used different calibration curves. Hence, the parameter estimation procedure was carried out many times and with different methods and the results here reported are averaged (a complete set of calibration curves and best fits with Equation (13) for a growing test in fed-batch and outdoor light conditions can be found in the Supplementary Material, Figures S5–S7).

The ranges of the operating parameters were $T = (19–25) ^\circ\text{C}$, $\text{pH} = (7.5–9.5)$, $\text{DO}_x = (13–21) \text{ mg L}^{-1}$ and $\text{EC} = (400–820) \mu\text{S}$ (two time-series of one double cultivation test can be found in the Supplementary Material, Figure S8).

Mass balances and best-operating conditions at the pilot-scale are resumed in Table 10.

Table 10. Operating conditions of 12 airlift photobioreactors fed with recycling wastewater and flue-gas.

Light Conditions	Regime	Recycled Wastewater	Required Sparging	Recycled Flue-Gas	Extraction + Feeding	Brought to Volume
Summer daylight	Churn flow $\text{Re}_{\text{riser}} = 11,000$	28.8 L d ⁻¹	20 m ³ h ⁻¹ × 12	90 m ³ h ⁻¹	Day 3, 6 $V = 6.2 \text{ L} \times 12$	Day 1, 2, 4, 5
Winter daylight	Churn flow $\text{Re}_{\text{riser}} = 10,300$	28.8 L d ⁻¹	20 m ³ h ⁻¹ × 12	90 m ³ h ⁻¹	Day 6 $V = 9.1 \text{ L} \times 12$	Day 1, 2, 3, 4, 5
Night conditions	Bubble flow $\text{Re}_{\text{riser}} = 5200$	28.8 L d ⁻¹	3–4 m ³ h ⁻¹ × 12 air	Storage tank	-	-

3.5. The Potential of Glycerol, a By-Product of the Production of Bioesters

The weight composition of the produced crude glycerol is glycerol (55–60%), methanol (25–30%), water (8%), fatty acids esters (3%), salts of fatty acids (2% *w/w*), and sodium chloride (2%).

The simplest method of valorizing crude glycerol consists of its combustion, which does not require any purification. The LHV of glycerol is about two times lower than that of fossil fuels, and it is comparable to the heat of combustion of most types of biomass. Due to the high content of water in crude glycerol obtained as a by-product of oil transesterification, often a co-combustion of glycerol with other liquid fuels is performed in special burners [45]. Moreover, pyrolysis and gasification are used, and they are usually carried out at 650–800 °C under atmospheric pressure [46–48]. Steam reforming [49] is another valuable process to produce hydrogen from crude glycerol, and the most effective catalysts used are silica or (Al₂O₃)-supported nickel, cobalt, nickel/copper, platinum, and other metals such as Rh, Ir, Ru, Zr, Ce, La [50].

The authors [21,43] proposed to use crude glycerol in co-gasification with biomass, in the form of solid waste pellets, to heighten the LHV and improve the quality of the syngas. In this perspective, a pursuable possibility consists in designing a small-scale energy-generating plant hinging on an internal combustion engine running on biodiesel

coupled with a gasifier exploiting the biomass impregnated with glycerol. In this sense, some experimental tests were performed by comparing the syngas produced from wood chips, pellets, with that originating from the glycerol-imbued biowaste [43], confirming the increase of the combustion quality due to the addition of crude glycerol to the organic wastes, manifested in both the increase of the flues gas temperature, gauged at the end of the combustion zone, and in a better flue gas chemical composition. Nevertheless, it would be advisable to carry out further testing in order to optimize the gasification process.

On a broader note, additional ongoing studies are focused on enlarging the concept of the proposed tending-to-zero-waste TE process. In particular, the authors are analyzing photocatalysis and bioconversion to directly produce hydrogen.

4. Discussion

4.1. Kinetics of Transesterification

The kinetics of biodiesel production from waste sunflower oil was deeply investigated by adopting three different reaction schemes. The first scheme considered a three reversible second-order reaction network, the second scheme took into consideration one fourth-order reversible reaction while the third scheme was based on one pseudo-first-order irreversible reaction. We computed the activation energies and the pre-exponential factors for the first scheme from kinetic data reported in [36] that seem to better describe our experimental data among the possible kinetics found in the literature. Furthermore, we estimated the kinetic parameters from our experimental data, using both the second and the third reaction schemes.

Regarding the waste sunflower oil, the obtained values with the fourth-order reversible reaction are $E_{a,direct} = 35,661 \text{ J mol}^{-1}$, $E_{a,reverse} = 72,989 \text{ J mol}^{-1}$, $k_{0,direct} = 9.7708 [\text{dm}^3 \text{ mol}^{-1}]^3 \text{ min}^{-1}$, and $k_{0,reverse} = 24,810 [\text{dm}^3 \text{ mol}^{-1}]^3 \text{ min}^{-1}$.

The obtained values for the pseudo-first-order transesterification of waste sunflower oil are $E_a = 67,348 \text{ J mol}^{-1} \text{ K}^{-1}$ and $k_0 = 2.157 \times 10^9 \text{ min}^{-1}$, in agreement with [26], in which values in the range $33,600\text{--}84,000 \text{ J mol}^{-1} \text{ K}^{-1}$ are reported, in agreement with [51] where values of $E_a = 79,000 \text{ J mol}^{-1} \text{ K}^{-1}$ and $k_0 = 2.98 \times 10^{10} \text{ min}^{-1}$ are found for soybean waste oil and a catalyst based on snail shell.

By comparing kinetics estimated for the different oils, the most important obtained results can be summarized as follows:

1. Similar yields in biodiesel are obtained for the waste and the virgin sunflower oil using for the former an almost doubled quantity of sodium hydroxide catalyst; the probable reason being that in the exhaust oil more free fatty acids are present;
2. A pseudo-first-order reaction scheme can be adopted to model the transesterification reaction of different kinds of oils in the chosen temperature ranges, only paying attention that the system is reaction controlled for almost all the reaction time. This condition is assured at $Re = 8000$ at lab-scale;
3. The values of the activation energy estimated from the experimental data do not differ a lot by changing the type of oil (sunflower, rapeseed, and olive oil) and this is in agreement with the literature, where similar values of activation energy are found for different waste/virgin oils. Conversely, the estimated values of the pre-exponential factor highly depend on oil type and operating conditions.

4.2. Quality of Biodiesel

Since glycerol and alcohol excess pose major fuel contamination issues, washing of biodiesel was carried out by bubble washing before separation.

We computed the cetane number from GC-MS analysis. Generally, for values as high as 55 to 60, an improvement in biodiesel emissions is witnessed, although exceeding that threshold showed only a slight improvement. The cetane number of biodiesel from olive oil, which amounted to 58.4 ± 0.2 , is then inside the optimal range, while that of biodiesel from rapeseed oil is reasonably near (53.4). EN 14214 prescribes a minimum of 51, while ASTM D975 (American Standard Specification for Diesel Fuel Oils) set the minimum value

of cetane number at 47; this means that biodiesel produced from both new (47.8 ± 0.2) and used (47.9 ± 0.2) sunflower oil does not conform to the European standards but only for the American ones. The dissimilarities in cetane numbers are caused by the chain length (cetane number increases with it), the number and location of double bonds (cetane number is inversely proportional to the number of double bonds), and changes with various locations of the carbonyl group. Moreover, through GC investigations potential instability issues have emerged: peaks on chromatograms of every biodiesel sample were identified in correspondence with the retention time of the oleic acid and possibly linoleic acid, and the degree of saturation of the fatty acid chains tends to be correlated with stability.

The energy content of biodiesel was measured by LHV. Generally, the fuel consumption is proportional to the volumetric energy density of the fuel based on the lower heating value. In order to provide a comparison, petroleum diesel contains about $30,473 \text{ [kJ kg}^{-1}\text{]}$, while biodiesel accounts for about $27,177 \text{ [kJ kg}^{-1}\text{]}$ [52]. The results of the tested biodiesel samples are then excellent since they all surpass $36,000 \text{ [kJ kg}^{-1}\text{]}$.

4.3. Microalgae Using Washing Wastewater

Even if the washing wastewater contained residuals of methanol, catalyst and salts of fatty acids (Table 5), as well as glycerol, the experiments confirmed that microalgal strain *Chlorella v.* has a good capability of using this wastewater to grow in outdoor and in mixotrophic conditions. The experiments confirmed interesting results in terms of average biomass production, ranging from 233 to $266 \text{ g L}^{-1} \text{ d}^{-1}$. Moreover, the yields obtained are about 25% greater than those obtained with the same strain in control conditions. The first results about the percentage of lipids extracted are in agreement with the literature. Finally, the chemical analysis of the wastewater in exit confirm that glycerol conversion by microalgae is in the range 60–80%.

4.4. Feasibility of the Coupled Process

The energetic performance of the NaOH-activated TE process can be estimated by computing the energy equivalents of the reactants and products and by adding the direct energy consumption. A net energy production of 13.70 MJ L^{-1} , of which about 78% is renewable, was calculated in [53] by considering a value of 37.25 MJ L^{-1} for the produced biodiesel (in agreement with our LHV = $36,600 \text{ MJ kg}^{-1}$). Usually, the cost of wastewater treatment is not considered in these calculations, so the use of washing wastewater as a feed to microalgae can increase the global techno-economic performance of this biodiesel production, even if a classical catalyst as NaOH is used.

Calculated absolute yields at TG:M = 1:5, cat = 1.2% w/w, residence time (t_R) = 60 min and $T = 60 \text{ }^\circ\text{C}$ varied from $77 \pm 3\%$ and $82 \pm 2\%$. They are in agreement with the results obtained in previous works even if lower, as expected, than those obtained with higher TG:M ratios, as reported in [54] where for TG:M = 1:12, cat = 2% w/w, $t_R = 70$ min and $T = 55 \text{ }^\circ\text{C}$ a yield of 94% is obtained. In [55], NaOH catalyst was used for the TE of waste sunflower oil added with oil extracted from microalgae and a calculated yield of 95% in the optimal conditions of TG:M = 4:6 v/v, $t_R = 130$ min and $T = 50 \text{ }^\circ\text{C}$ is reported. Other studies show yields greater than 90% only with very high TG:M ratios and high residence times. Many research activities have recently been devoted to the development of heterogeneous catalysts, as acid-supported porous or metal-supported polymers. In [56] a maximum yield of 95% is reached with TG:M = 1:15, 4% w/w catalyst, $t_R = 240$ min and $T = 120 \text{ }^\circ\text{C}$ and using a MgAl-LDH-geopolymer. The operating conditions proposed in this work with yields reaching 82% are in our opinion the correct ones for the scale-up of the process to a continuous production. Higher yields, obtained at laboratory scale with residence times of more than two hours and high TG:M ratios could not be convenient for integrating this process into industrial scale biorefineries, since the increase of t_R needs high reactor volumes (with possible mixing problems) or lower flow rates so decreasing production rate, and the increase of TG:M requires higher costs for methanol recycling.

5. Conclusions

In this work we proposed to improve the feasibility of a second-generation biodiesel production process by coupling it with microalgae cultivation. The transesterification of waste sunflower oil with methanol and NaOH as catalyst is analyzed. This TE process needs a significant washing step since appreciable quantities of soaps can be formed, but if the wastewater in exit from the washing unit, rich in glycerol, is used as a feed to the microalgae, it becomes very interesting at industrial scale. At TG:M = 1:5 *w/w*, $T = 50\text{--}60\text{ }^{\circ}\text{C}$, catalyst load = 1.2% *w/w* and residence time = 60 min, the yield at the pilot scale reached 82%. Three different kinetic schemes were investigated, and the study showed that a pseudo-first order scheme well represents the reaction mechanism at the chosen conditions. Results were also reported about growing tests of *Chlorella vulgaris* cultivated in outdoor and in mixotrophic conditions, fed with washing wastewater. An increase of 25% in microalgae production was obtained. Finally, the mass balance of the designed continuous pilot-scale coupled process was reported, showing that a cyclical feeding can be designed in order to recycle all the washing wastewater produced.

Supplementary Materials: The following are available online at <https://www.mdpi.com/article/10.3390/su14010273/s1>, Table S1: GC device technical conditions and specifics; Figure S1: GC of biodiesel from virgin olive oil ($T = 60\text{ }^{\circ}\text{C}$, molar TG:M = 1:5, cat = 0.6 *w/w*); Table S2: GC data of biodiesel from virgin olive oil ($T = 60\text{ }^{\circ}\text{C}$, molar TG:M = 1:5, cat = 0.6 *w/w*); Table S3: FAME in biodiesel produced from virgin olive oil ($T = 60\text{ }^{\circ}\text{C}$, molar TG:M = 1:5, cat = 0.6 *w/w*); Figure S2: GC of biodiesel from virgin rapeseed oil ($T = 60\text{ }^{\circ}\text{C}$, molar TG:M = 1:5, cat = 0.6 *w/w*); Table S4: GC data of biodiesel from virgin rapeseed oil ($T = 60\text{ }^{\circ}\text{C}$, molar TG:M = 1:5, cat = 0.6 *w/w*); Table S5: FAMES in biodiesel produced from virgin rapeseed oil ($T = 60\text{ }^{\circ}\text{C}$, molar TG:M = 1:5, cat = 0.6 *w/w*); Figure S3: GC of biodiesel from virgin sunflower oil ($T = 60\text{ }^{\circ}\text{C}$, molar TG:M = 1:5, cat = 0.6 *w/w*); Table S6: GC data of biodiesel from virgin sunflower oil ($T = 60\text{ }^{\circ}\text{C}$, molar TG:M = 1:5, cat = 0.6 *w/w*); Table S7: FAMES in biodiesel produced from virgin sunflower oil ($T = 60\text{ }^{\circ}\text{C}$, molar TG:M = 1:5, cat = 0.6 *w/w*); Figure S4: GC of biodiesel from waste sunflower oil ($T = 60\text{ }^{\circ}\text{C}$, molar TG:M = 1:5, cat = 1.2 *w/w* repl. 1/3); Table S8: GC data of biodiesel from waste sunflower oil ($T = 60\text{ }^{\circ}\text{C}$, molar TG:M = 1:5, cat = 1.2 *w/w*, repl. 1/3); Table S9: FAMES in biodiesel produced from waste sunflower oil ($T = 60\text{ }^{\circ}\text{C}$, molar TG:M = 1:5, cat = 1.2 *w/w*, repl. 1/3); Figure S5: Calibration curves at (a) 430 nm; (b) 480 nm; (c) 650 nm; (d) 680 nm; ϵ overall, at steps of 10 nm; Figure S6: Estimated growth curves at different wavelengths (each own calibration curve), (a) 430 nm, (b) 480 nm, (c) 650 nm, (d) 680 nm; Figure S7: Estimated growth curves at different wavelengths (using the overall calibration curve), (a) 430 nm, (b) 480 nm, (c) 650 nm, (d) 680 ϵ (e) global spectrum; Figure S8: Online measured parameters during microalgae cultivation.

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Appendix A

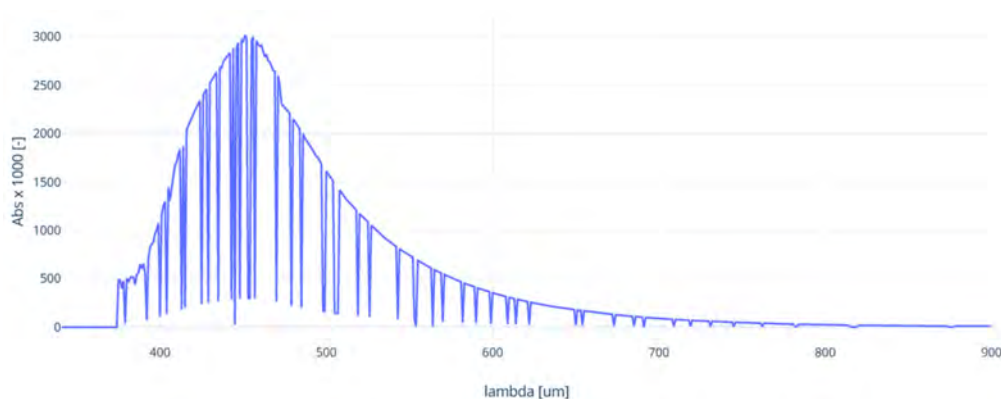


Figure A1. Spectrum of glycerol obtained by transesterification of waste sunflower oil, using glycerol from virgin sunflower oil as a blank.

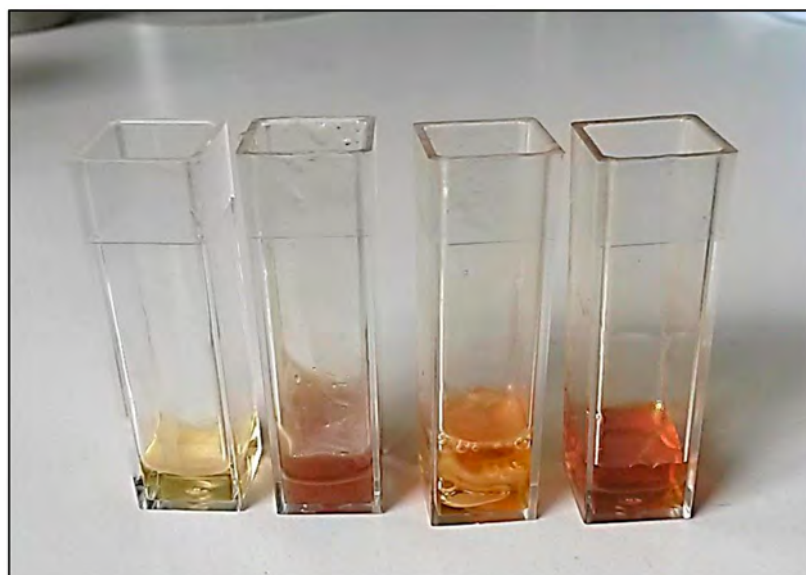


Figure A2. Glycerol samples obtained (starting from left to right) from olive oil, rapeseed oil, virgin sunflower oil, and waste sunflower oil.

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